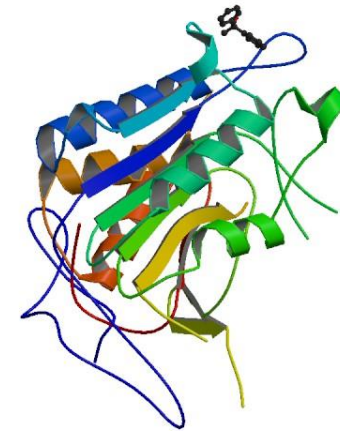
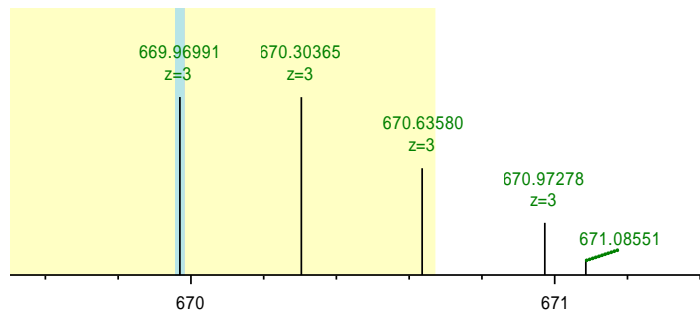




Production of omics data: Proteomics

Thibaut Léger, PhD

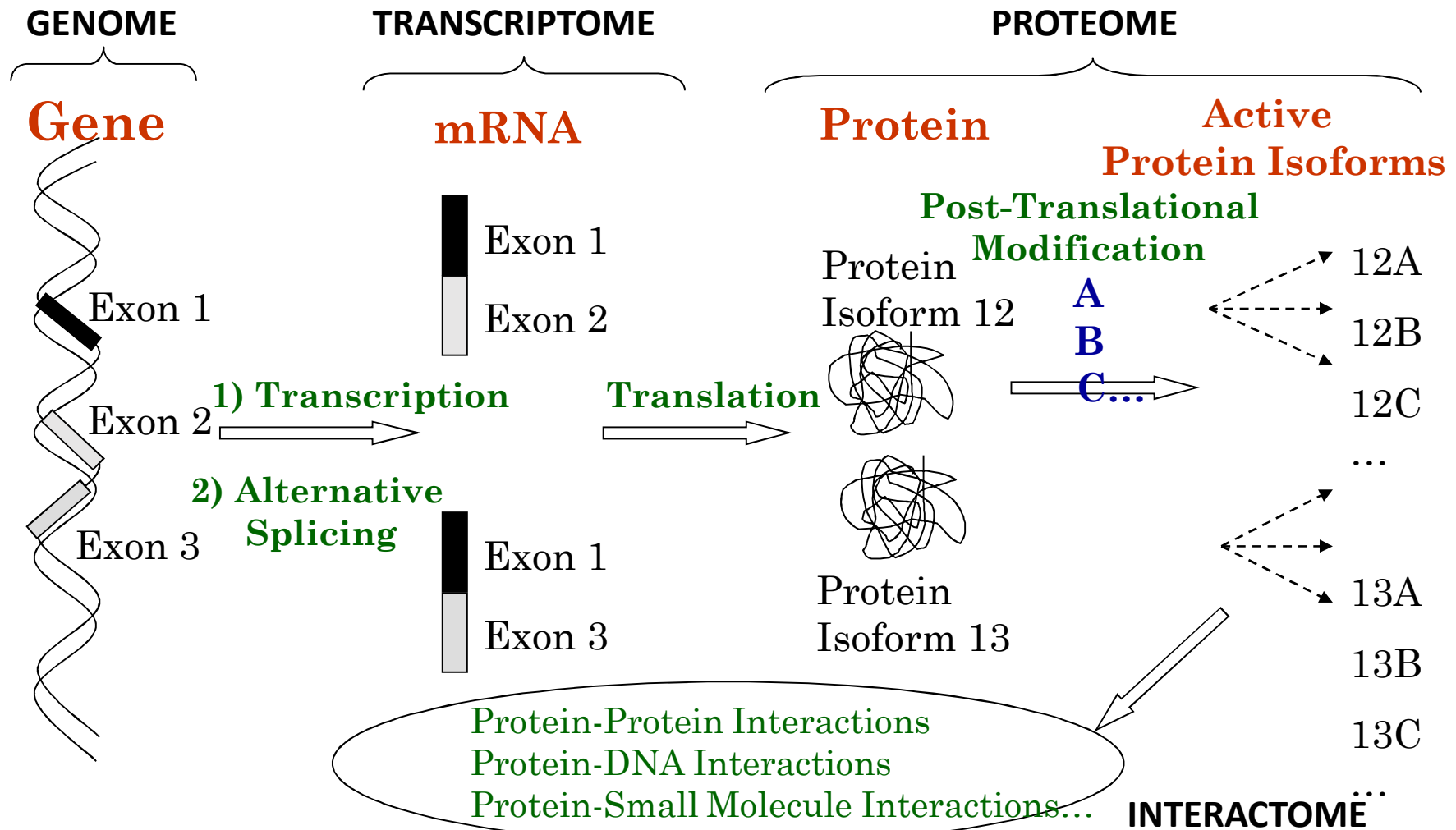
LERES, EHESP



DUBii

10th march 2021

CONCEPT



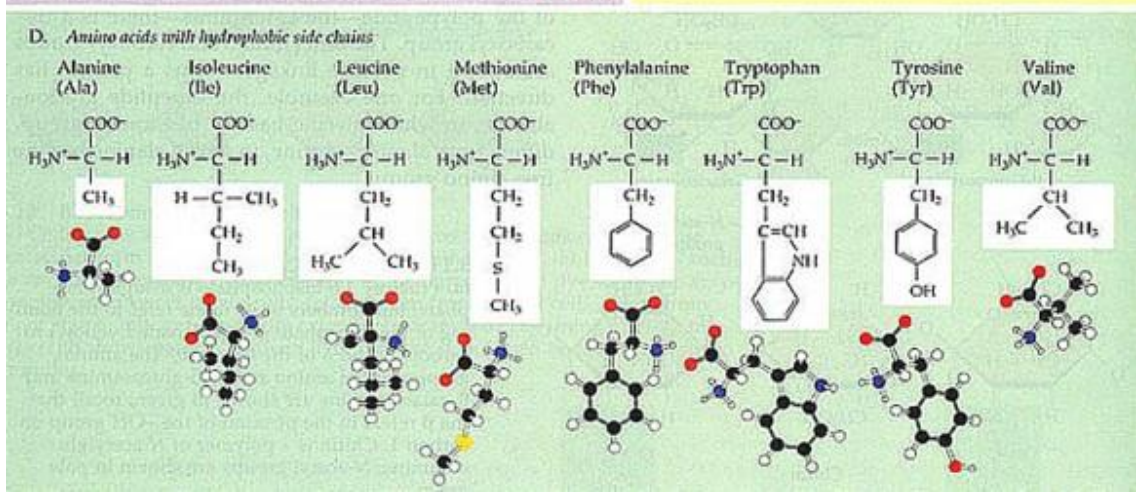
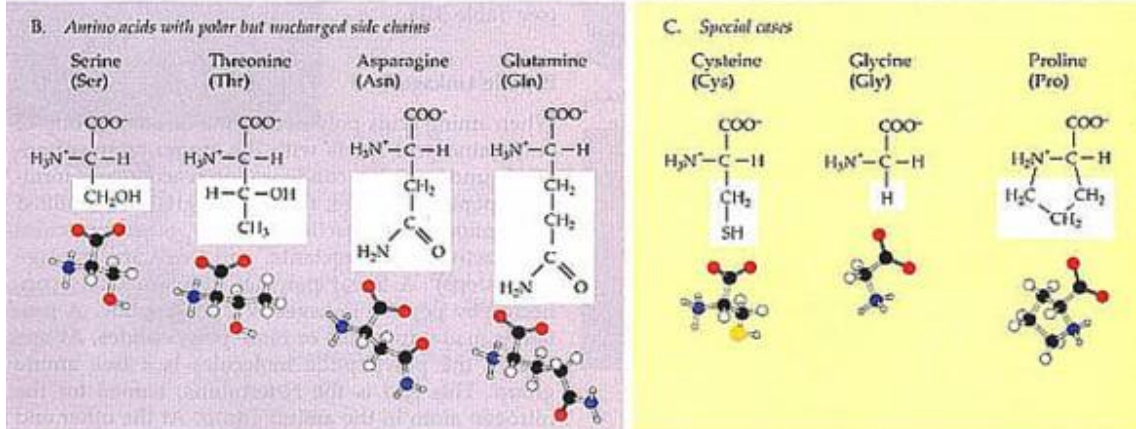
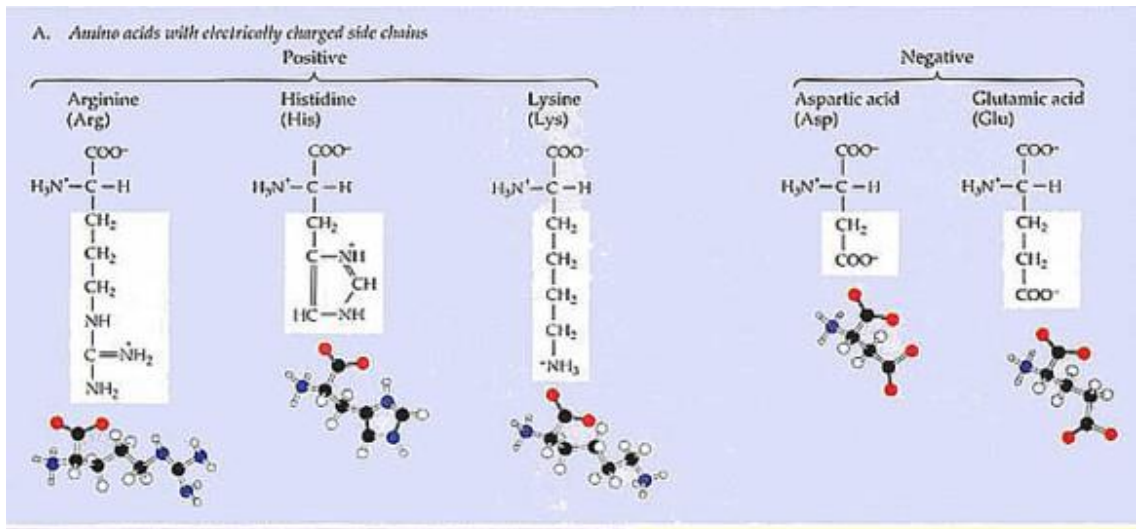
GENOMICS vs PROTEOMICS

Genome (DNA)

- “ Static (no change with time)
- “ Can be amplified (PCR)
- “ Little sample complexity
- “ *(4 base pairs, very similar, same order of concentration)*
- “ Good solubility

Proteome (proteins)

- “ Dynamic
- “ *(highly variable with time; many proteomes for one genome)*
- “ Cannot be amplified
- “ High sample complexity (wide variety of physical and chemical properties; concentrations can differ by 9 orders of magnitude)
- “ Various solubility; some proteins are insoluble in water



Alanine	A, Ala	71.079
Arginine	R, Arg	156.188
Asparagine	N, Asn	114.104
Aspartic acid	D, Asp	115.089
Cysteine	C, Cys	103.145
Glutamine	Q, Gln	128.131
Glutamic acid	E, Glu	129.116
Glycine	G, Gly	57.052
Histidine	H, His	137.141
Isoleucine	I, Ile	113.160
Leucine	L, Leu	113.160
Lysine	K, Lys	128.17
Methionine	M, Met	131.199
Phenylalanine	F, Phe	147.177
Proline	P, Pro	97.117
Serine	S, Ser	87.078
Threonine	T, Thr	101.105
Tryptophan	W, Trp	186.213
Tyrosine	Y, Tyr	163.176
Valine	V, Val	99.133

Proteomics?

- “ Proteomics is the large-scale study of proteomes, it means all proteins from a cell, an organelle, a tissue, an organ or from an organism at a one point, under specific conditions.
- “ Proteomics is at the crossroads of biochemistry, analytical chemistry and bioinformatics.
- ⇒ Proteins can be modified by different biological or chemical processes; The different variants of proteins are called now:

Proteoforms

Nat Methods. 2013 Mar;10(3):186-7. doi: 10.1038/nmeth.2369.

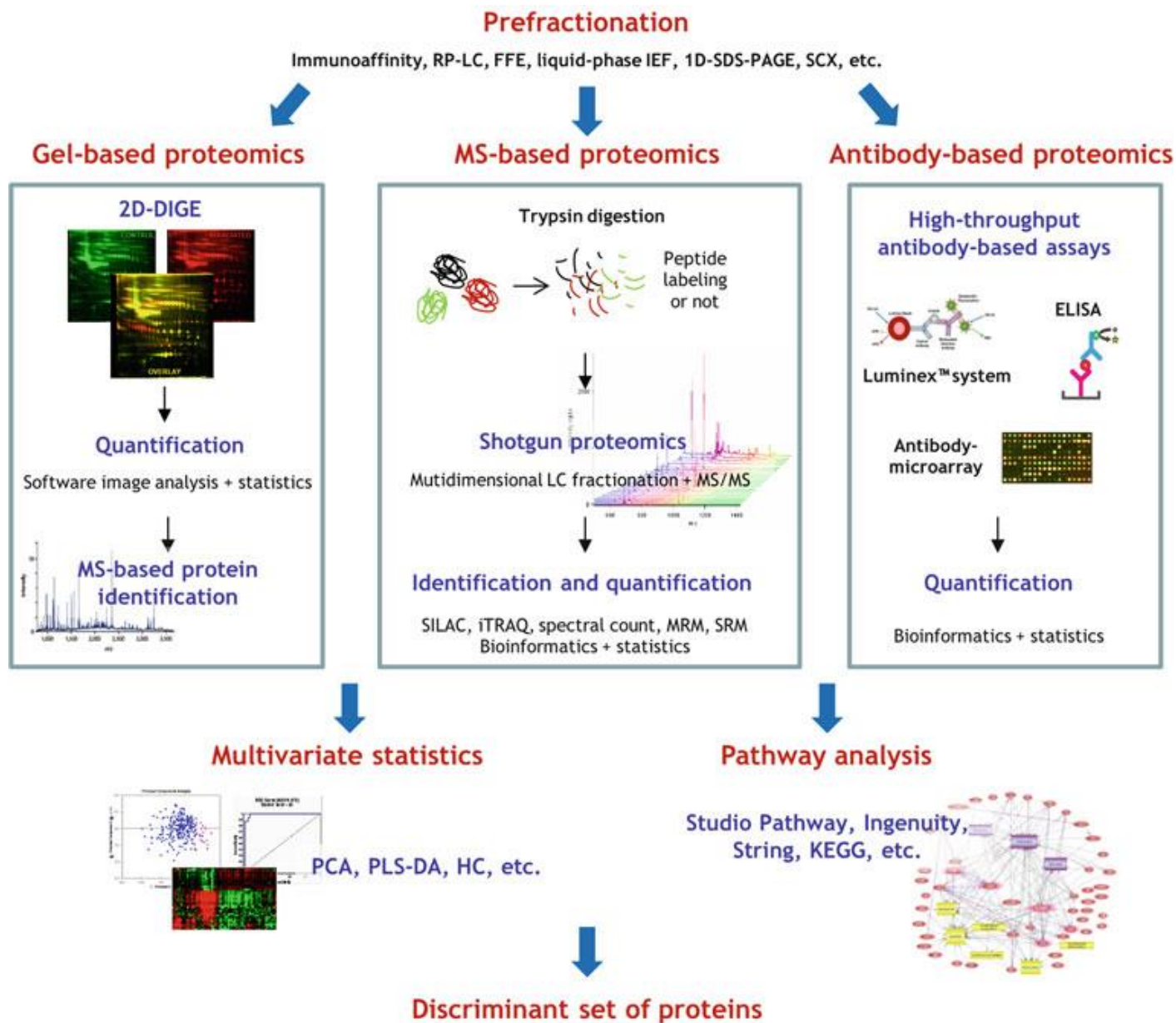
Proteoform: a single term describing protein complexity.

Smith LM, Kelleher NL; Consortium for Top Down Proteomics

PROTEOMICS GOALS

- “ Identification of all proteins in a proteome
- “ Search for new, hypothetical or predicted proteins
- “ Analysis of differential expression between 2,3,... different conditions (protein up- or downregulation)
- “ Identification of post-translational modifications
- “ Characterization of proteins by function, pathway, cellular location, etc.
- “ Study of protein-protein interactions

Proteomics techniques





HUMAN PROTEOME MAP

Statistics

Organs/cell types	30
Genes identified	17,294
Proteins identified	30,057
Peptide sequences	293,700
N-terminal peptides	4,297
Splice junctional peptides	66,947
Samples	85
Adult tissues	17
Fetal tissues	7
Cell types	6

Status

Human Proteome

Coverage:	80%
Proteins:	15721 of 19629
Isoforms:	11353 of 86771
Unique Peptides (Isoform):	113944
Unique Peptides (Gene):	455289
Spectra:	43237800

Repository

Registered Users:	533
Projects:	75
Experiments:	397
Files:	19459
Data Volume:	7.84 TB

Welcome to ProteomicsDB!

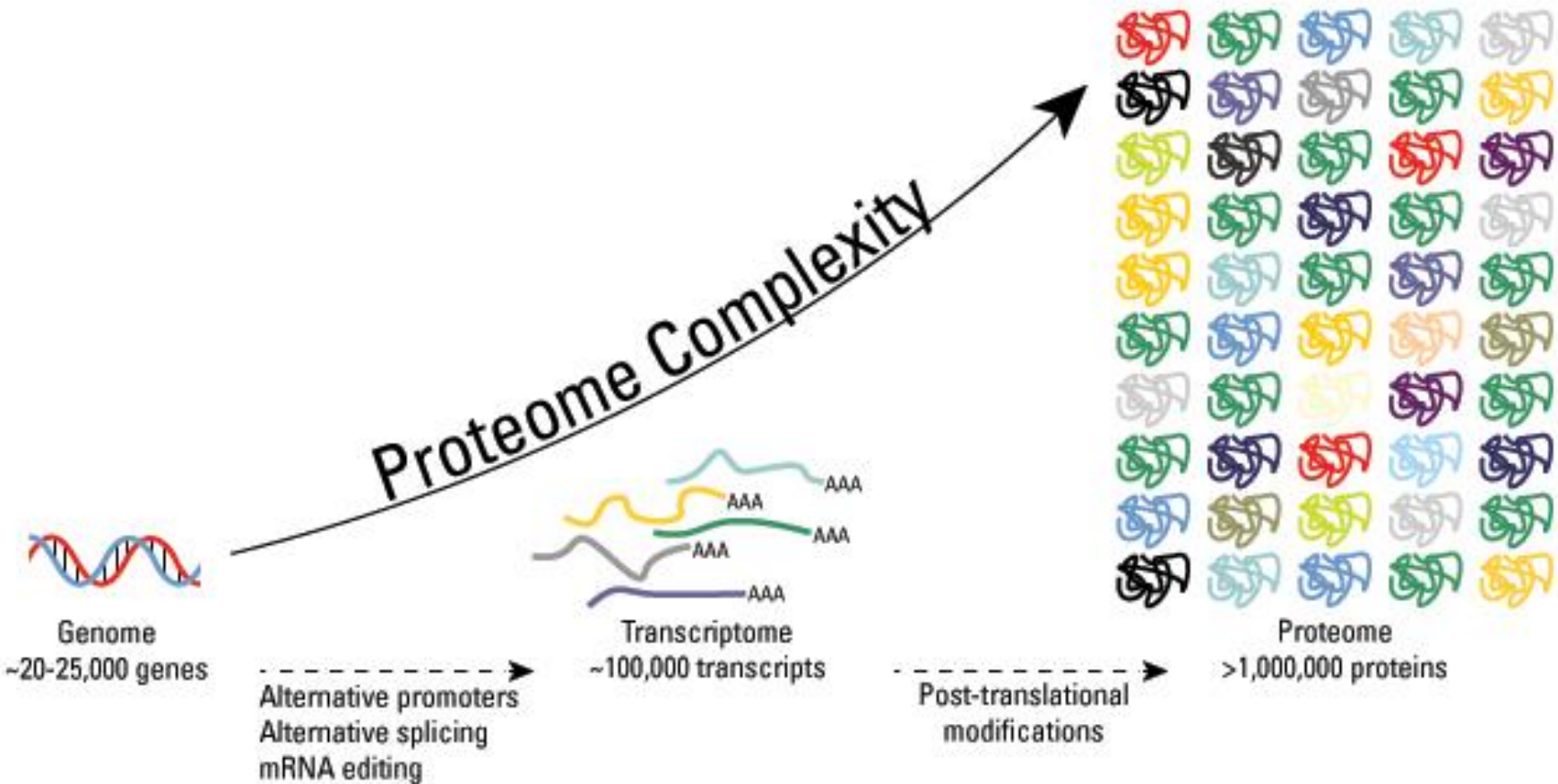
ProteomicsDB is a joint effort of the Technische Universität München (TUM) and the Max Planck Institute of Biochemistry (MPC) to map the human proteome and its use across the scientific community.



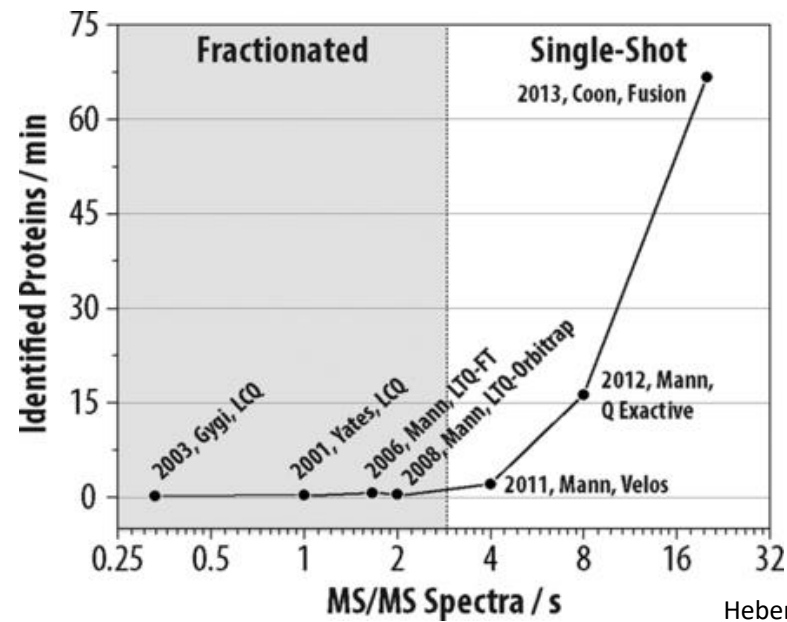
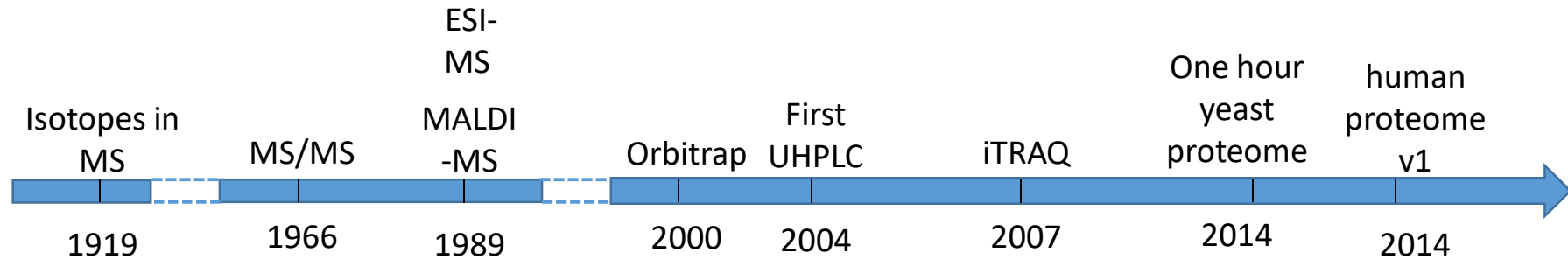
Browse proteins

Explore the human proteome protein by protein.

Problem of proteome complexity

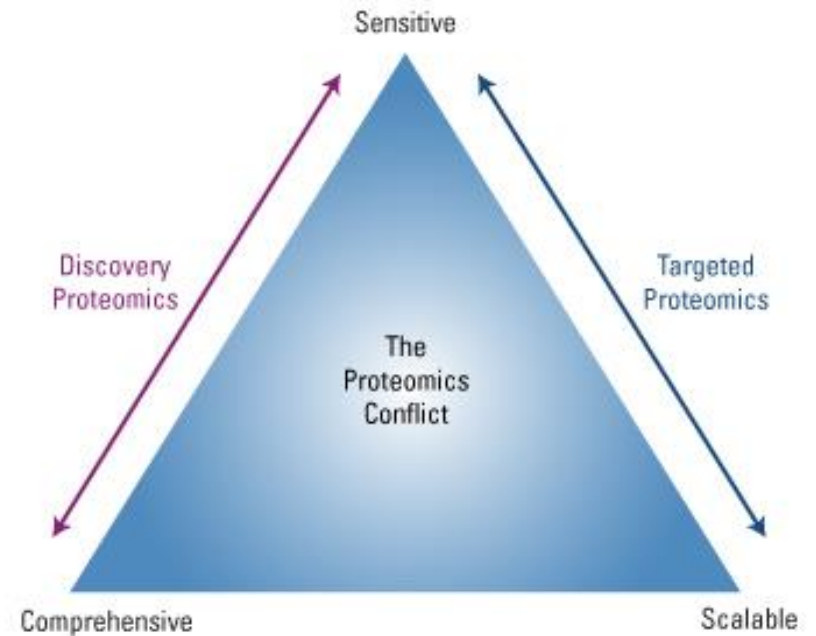
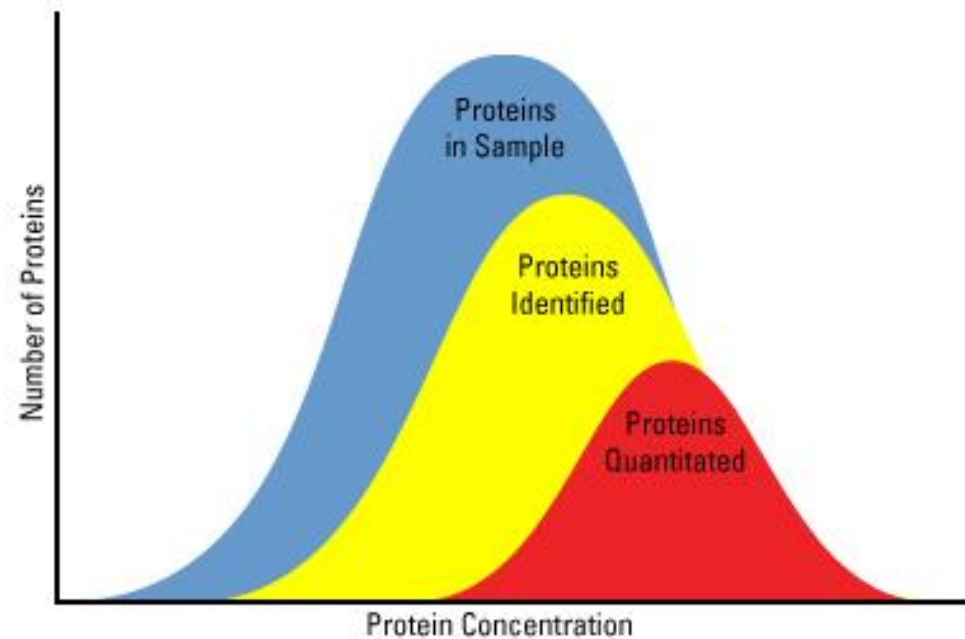


Evolution of proteomics performances



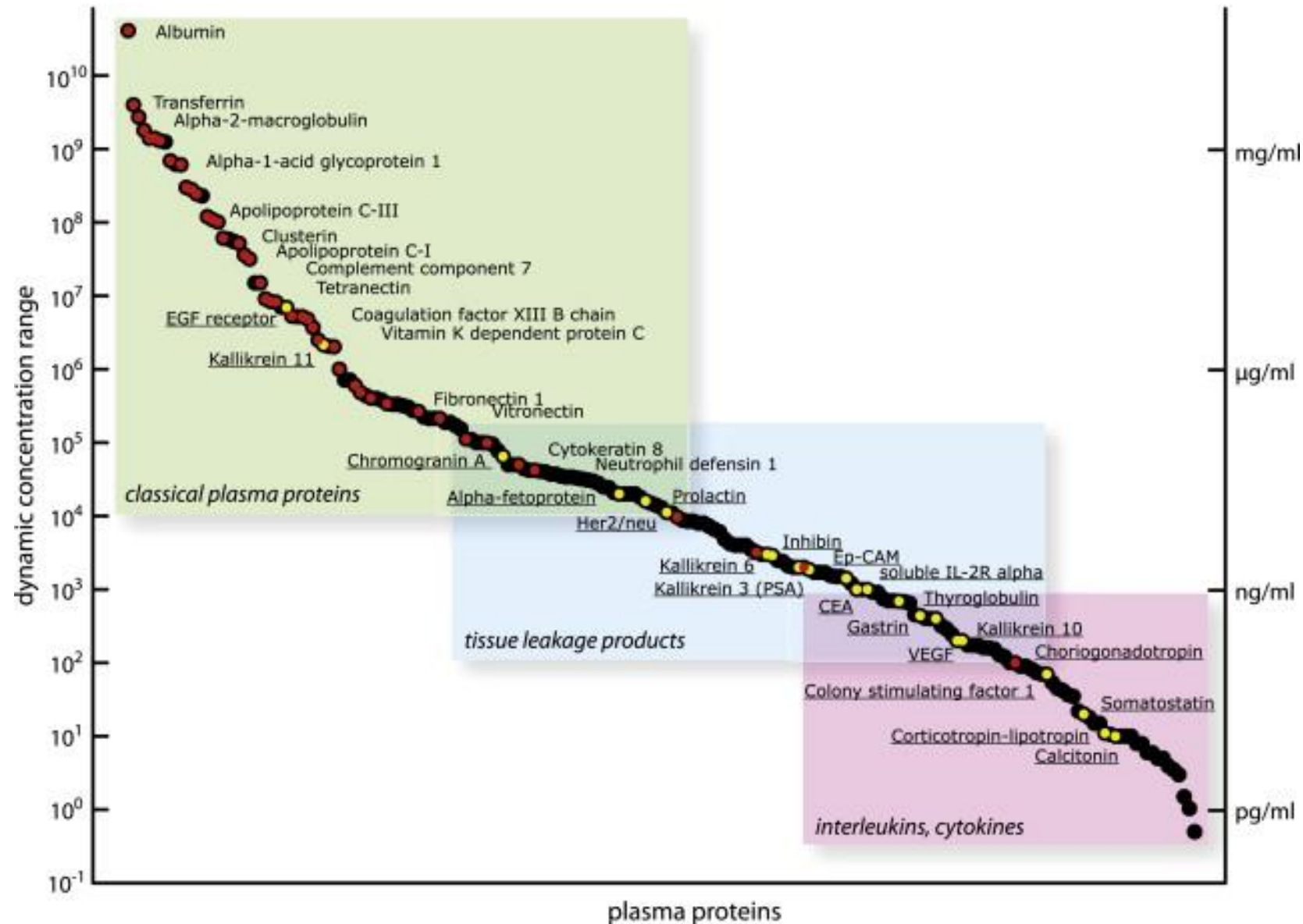
Hebert *et al.* 2009

Inherent dilemma linked to proteomics

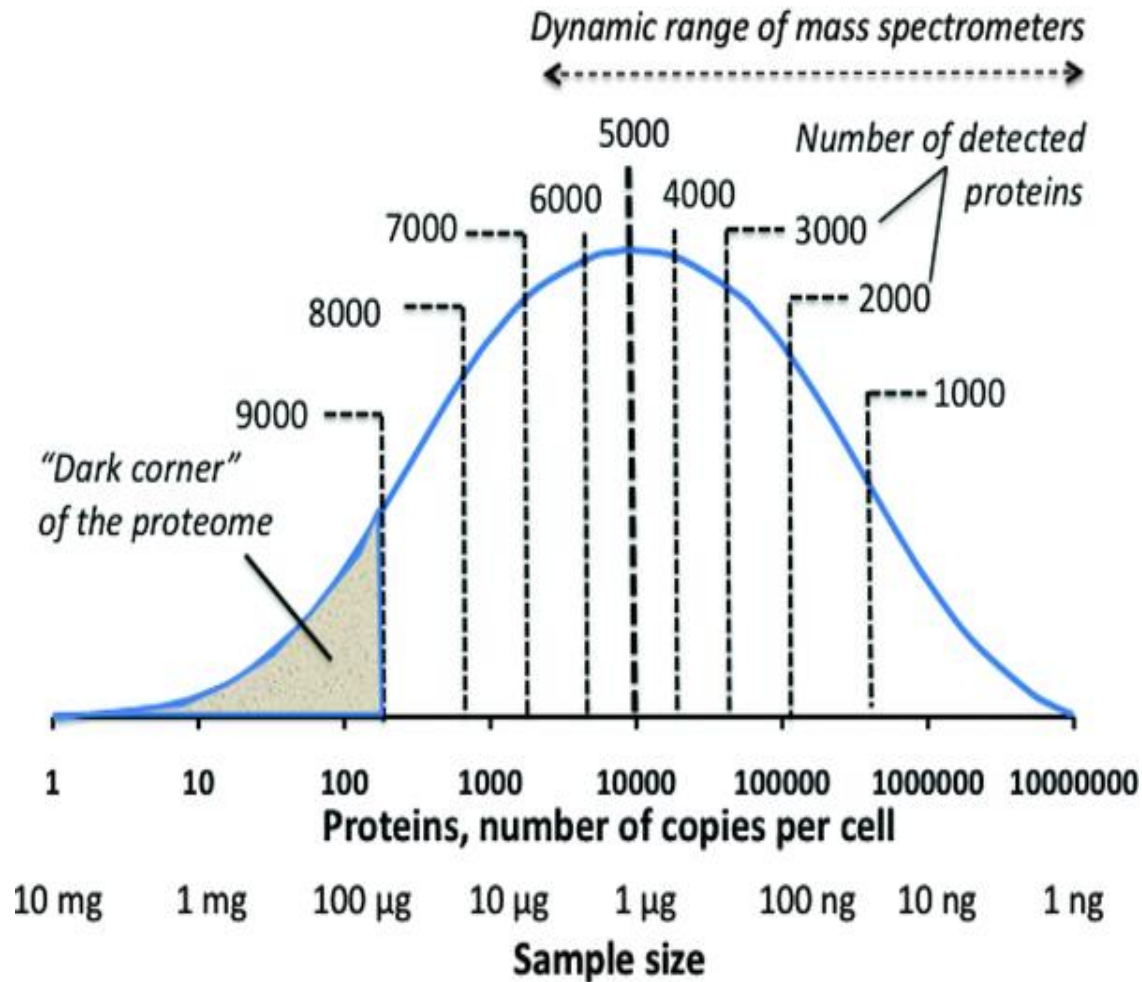


<http://www.piercenet.com/method/quantitative-proteomics>

Dynamic range in human plasma



Proteomics and proteome coverage



Zubarev et al. 2013

Dynamic range of proteomes

Key questions in proteomics



- “ What is the protein content of my biological sample?
=> problem of **identification**
- “ What is the abundance of my protein of interest?
=> **quantification**
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ What are the partners of my protein of interest?
- “ **Are there any signature proteins related to a particular biological process?**

=> **biomarkers identifications and quantifications**

Instrumentations

Micro/nanoHPLC



U3000 micro



EASY-nLC 1000



U3000 nano



EASY-nLC II

MALDI



4800 TOF-TOF

ESI



ORBITRAP-Velos-ETD:
Routine



Qexactive+:
Routine
Quantification



Fusion Tribrid:
TMT Quantification ciblée
R&D



NanoMate:
Infusion

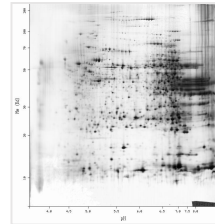
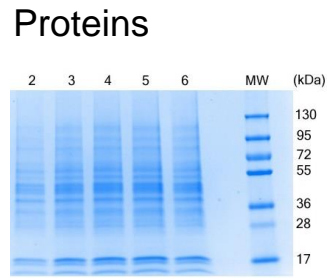
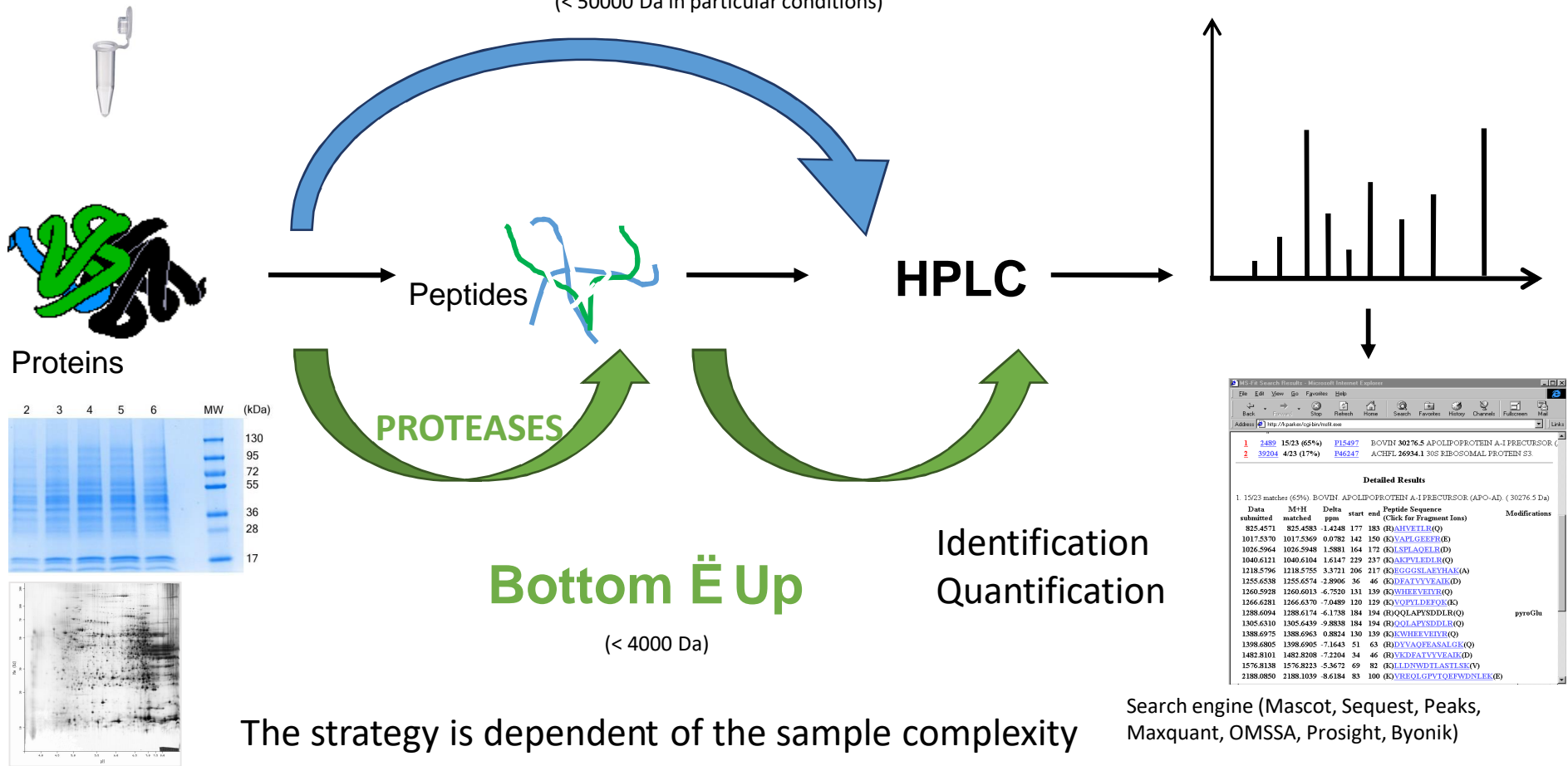
Proteomics workflows

Top Æ Down

(< 50000 Da in particular conditions)

MS acquisition

- ~ MALDI TOF/TOF
- ~ Orbitrap



The strategy is dependent of the sample complexity

Bottom Æ Up

(< 4000 Da)

Identification
Quantification

Search engine (Mascot, Sequest, Peaks, Maxquant, OMSSA, ProSight, Byonik)

BOTTOM-UP PROTEOMICS: PRO'S AND CON'S

Advantages

- “ Less sophisticated instrumentation and expertise
- “ High throughput
- “ More info about proteins with “extreme” phys.-chem. properties (hydrophobic, Hi/Low MW, acidic/basic)

Disadvantages

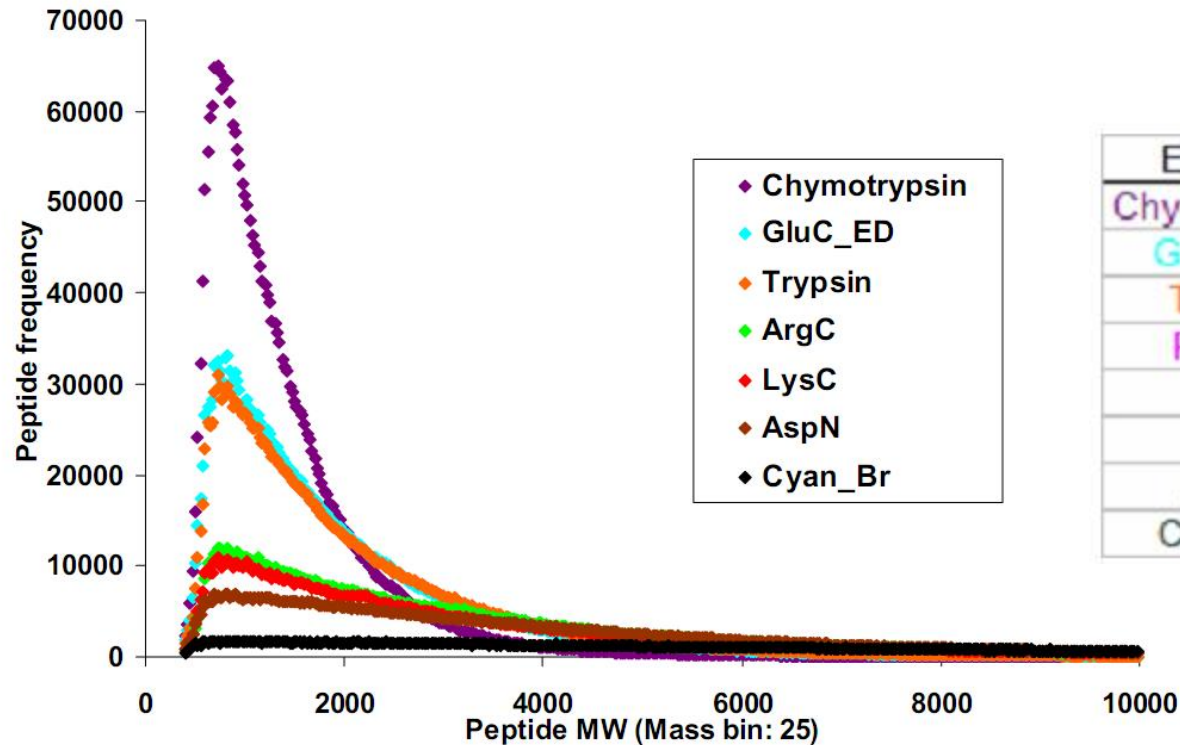
- “ Confidence in protein ID strongly depends on restriction criteria (subjective; potential bias)
- “ Since protein ID is often done by 1-2 peptides, PTM and isoform information is often lost

Cleavage rules of proteases

Enzyme or Reagent	Cleaves where?	Exceptions
Trypsin	C-terminal side of K or R	if P is C-term to K or R
Trypsin	(C-term to K/R, even before P)	C-terminal side of K or R
Trypsin (higher specificity)	C-terminal side of K or R	if P is C-term to K or R; after K in CKY, DKD, CKH, CKD, KKR; after R in RRH, RRR, CRK, DRD, RRF, KRR
Lys C	C-terminal side of K	
CNBr	C-terminal side of M	
Arg C	C-terminal side of R	if P is C-term to R
Asp N	N-terminal side of D	
Asp N + N-terminal Glu	N-terminal side of D or E	
Glu C (bicarbonate)	C-terminal side of E	if P is C-term to E, or if E is C-term to E
Glu C (phosphate)	C-terminal side of D or E	if P is C-term to D or E, or if E is C-term to D or E
Chymotrypsin	(C-term to F/Y/W/M/L, not before P, not after Y if P is C-term to Y) C-terminal side of F, L, M, W, Y	if P is C-term to F, L, M, W, Y, if P is N-term to Y
Chymotrypsin (C-term to F/Y/W/, not before P, not after Y if P is C-term to Y)	C-terminal side of F, Y, W	if P is C-term to F, Y, W, if P is N-term to Y
Trypsin/Chymotrypsin (C-term to K/R/F/Y/W, not before P, not after Y if P is C-term to Y)	C-terminal side of K, R, F, Y, W	if P is C-term to K, R, F, Y, W, if P is N-term to Y
Pepsin (pH 1.3)	C-terminal side of F, L	
Pepsin (pH > 2)	C-terminal side of F, L, W, Y, A, E, Q	
Proteinase K	C-terminal side of A, C, G, M, F, S, Y, W	

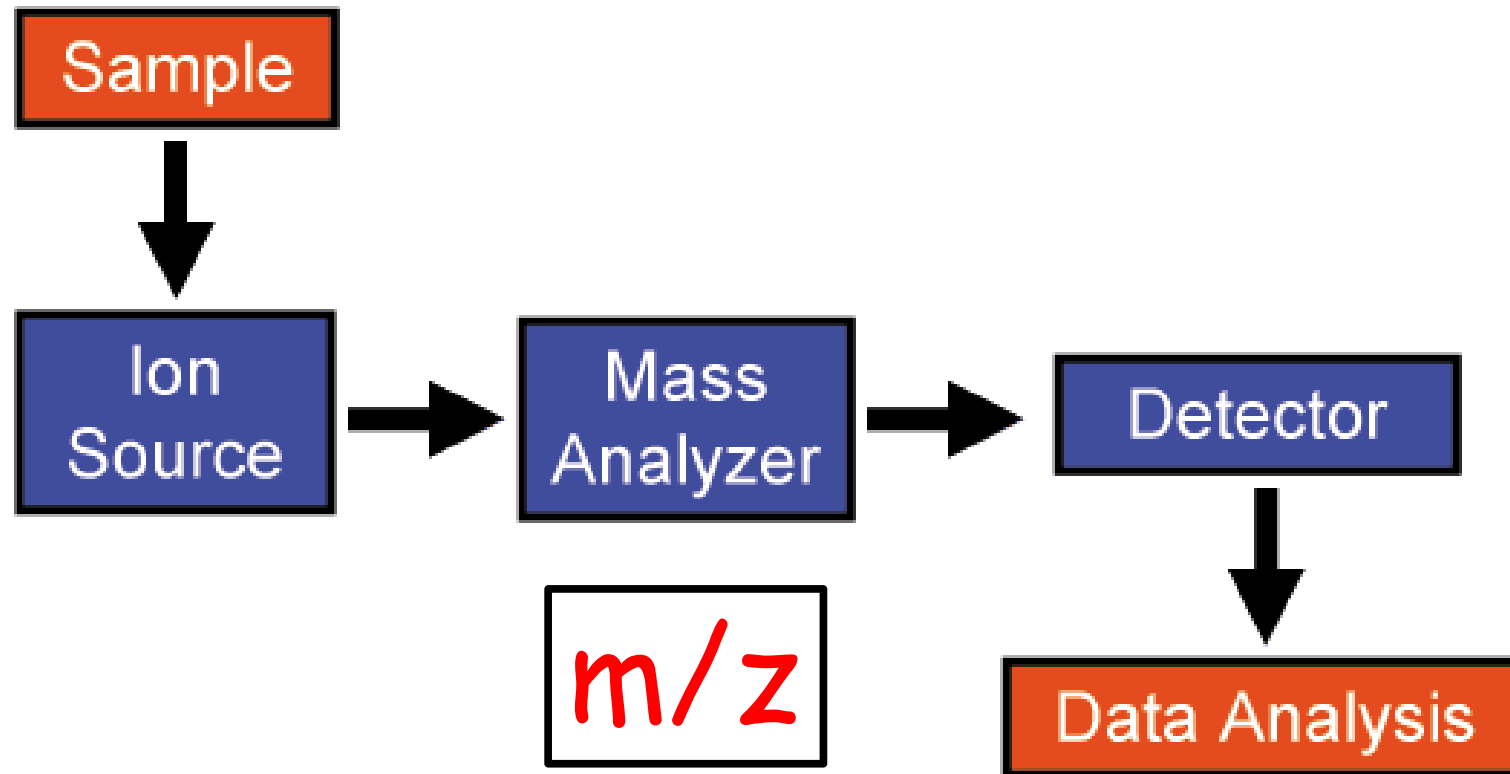
PEPTIDE LENGTH AND NUMBER OF PEPTIDES GENERATED DEPENDING ON ENZYME USED FOR DIGESTION

Other enzymes with more or less specific cleavage:

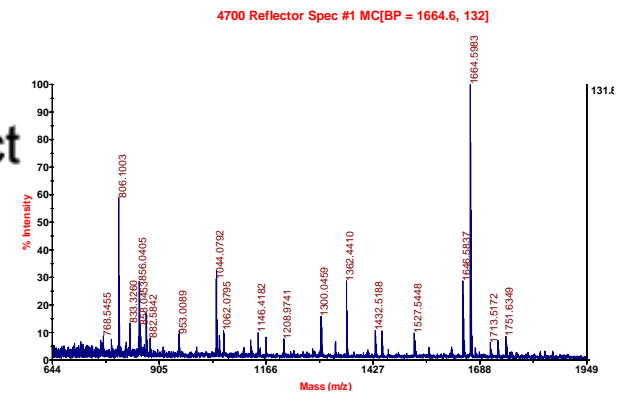
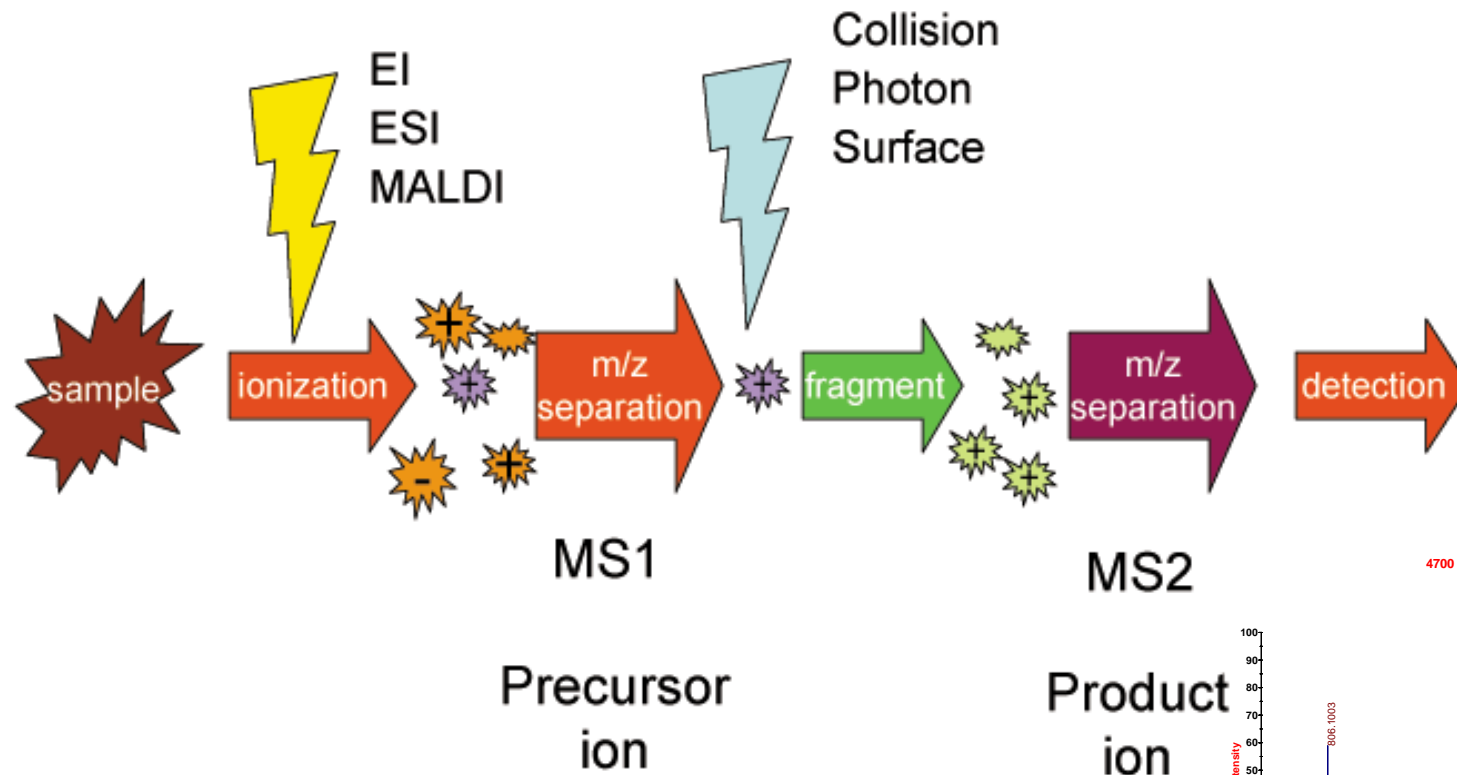


Advantages of a new proteomic approach that uses accurate mass measurements, LC retention time, isoelectric point and dual enzymatic digestion. Petritis K. et. al., Biological Sciences Division, Pacific Northwest National Laboratory, Richland, WA 99352; ASMS'2007 poster presentation http://www.chem.agilent.com/Library/posters/Public/Petritis_ASMS_2007.pdf

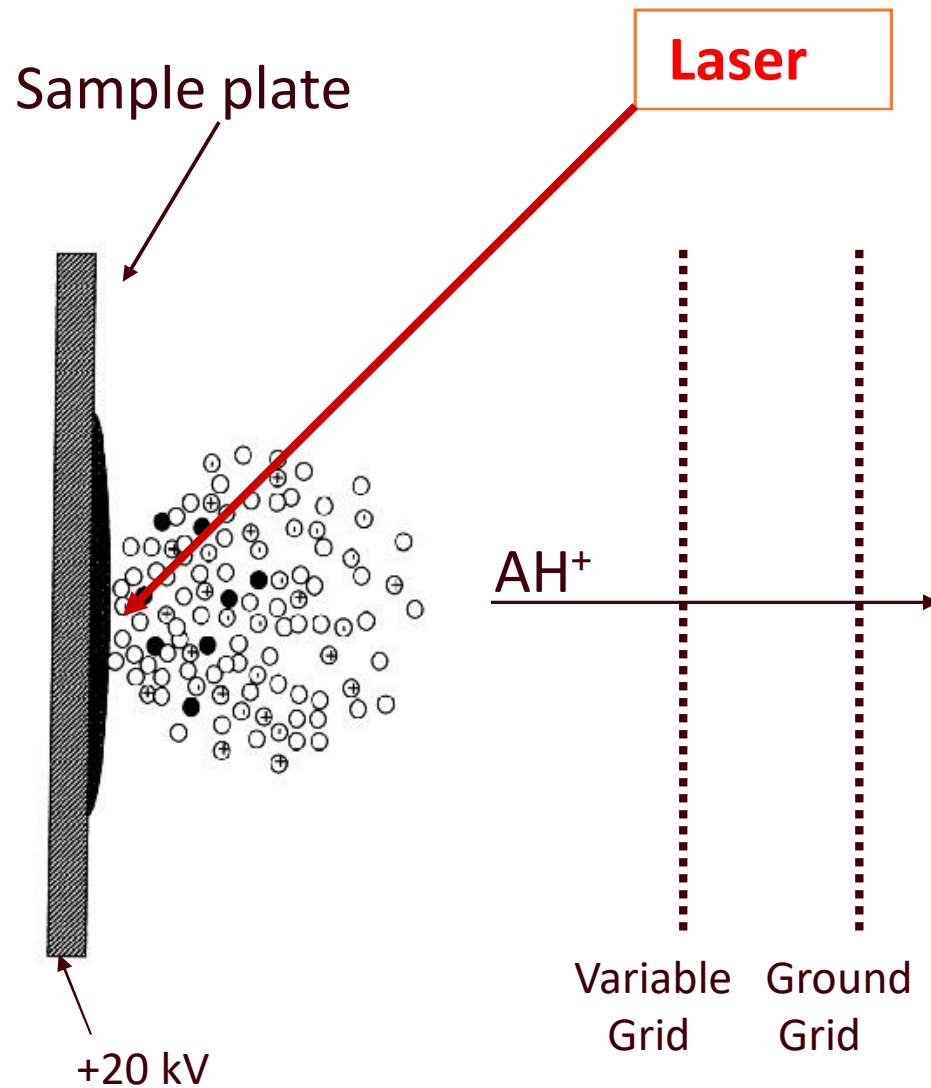
What is MS?



MS and MS/MS



MALDI ionization (Matrix Assisted Laser Desorption Ionization)



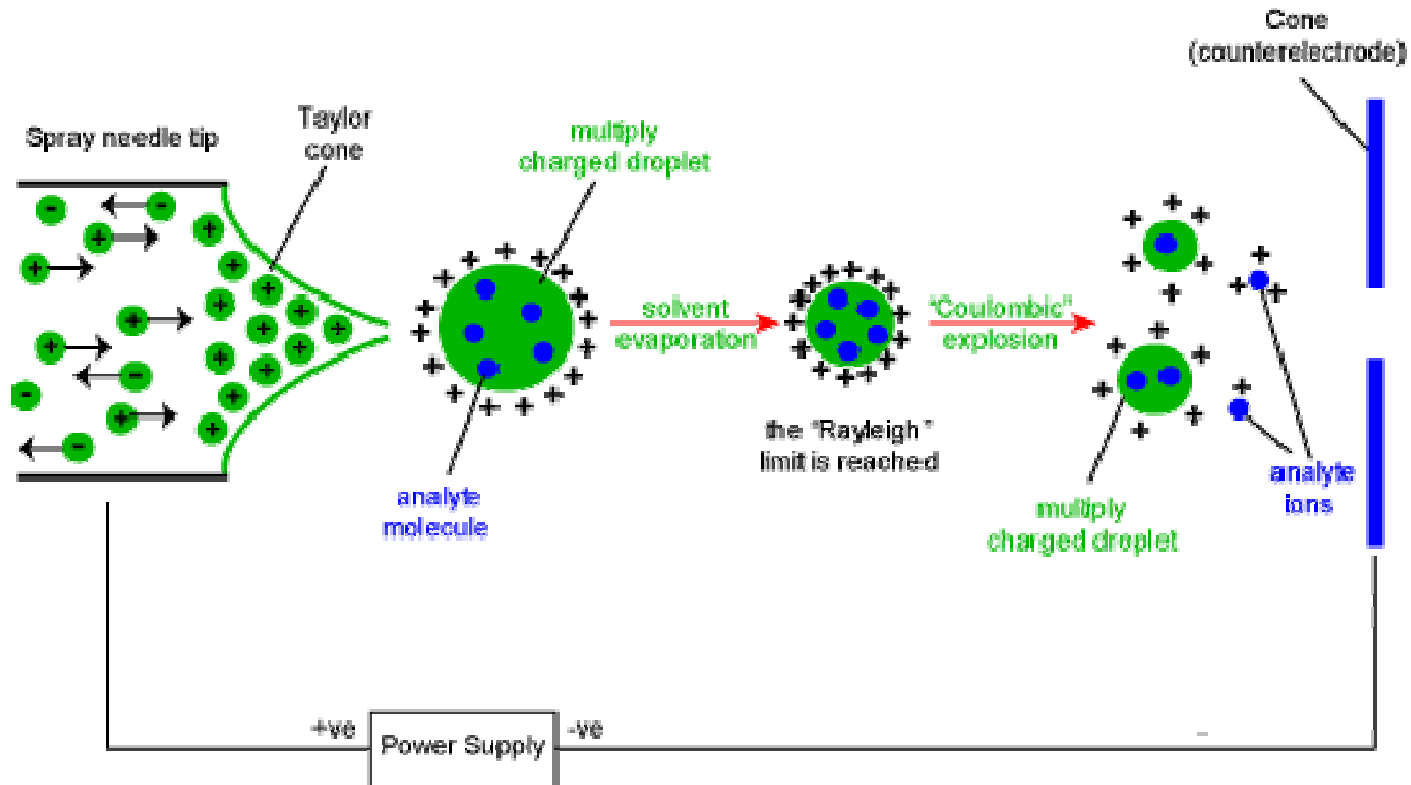
1. L'échantillon (A) est mélangé avec un excès de matrice (M) et séché sur la plaque MALDI

2. Le flash Laser ionise les molécules de matrice

3. Les molécules d'échantillon sont ionisées par transfert de protons de la matrice:



Ionization by electrospray

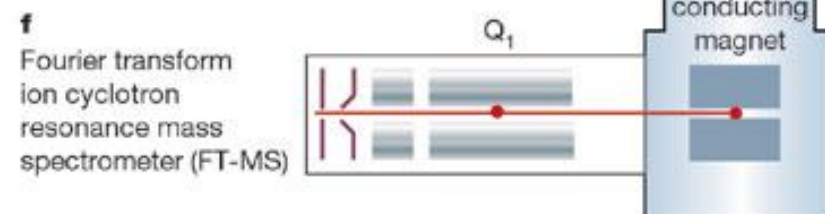
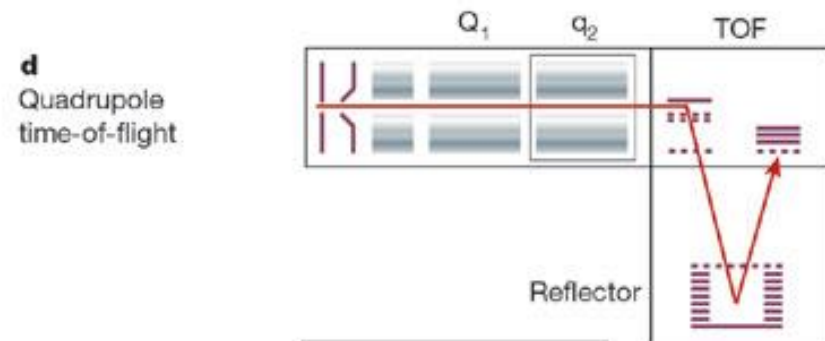
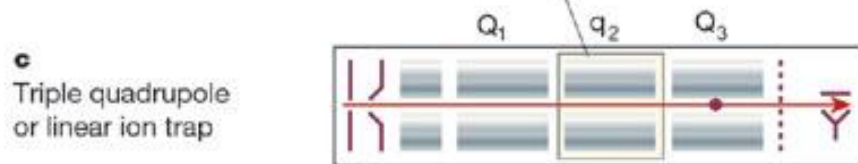
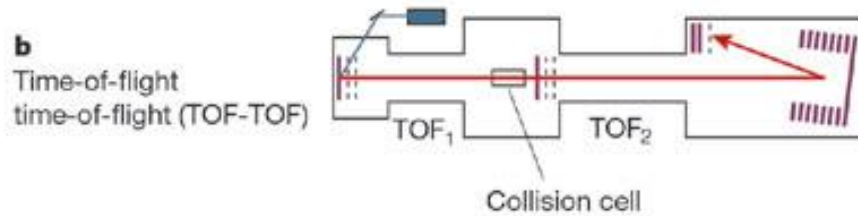
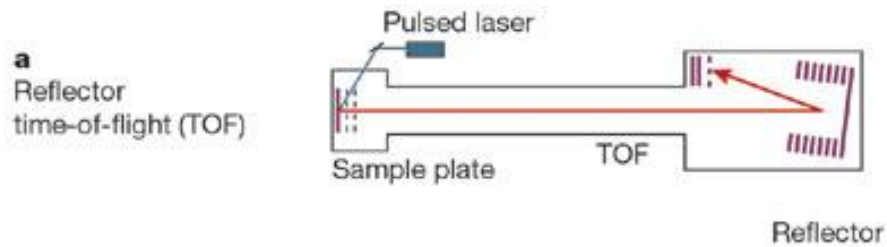
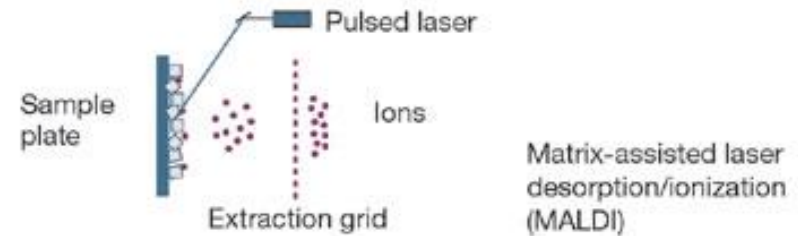
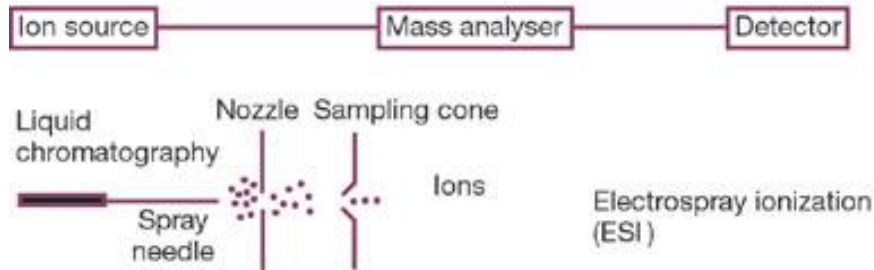


Electrospray

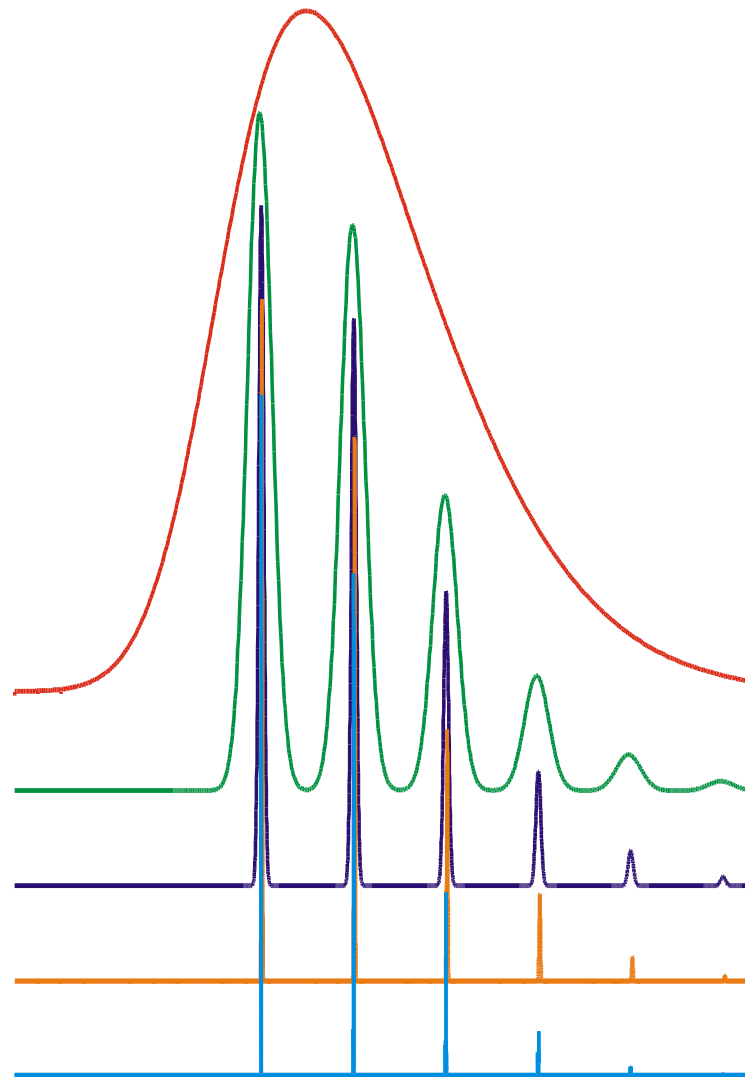


nanospray

Different instrumental design



Importance of spectral resolution

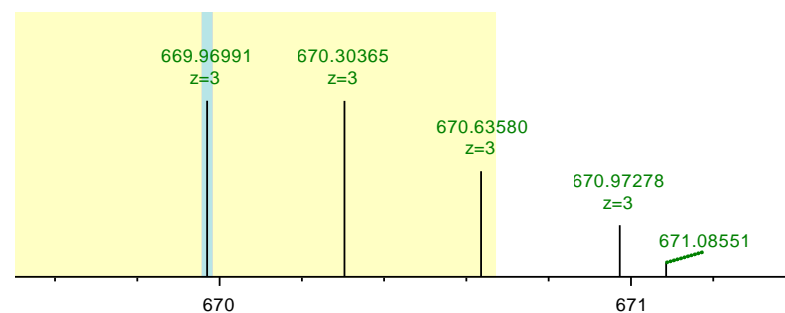


Resolution

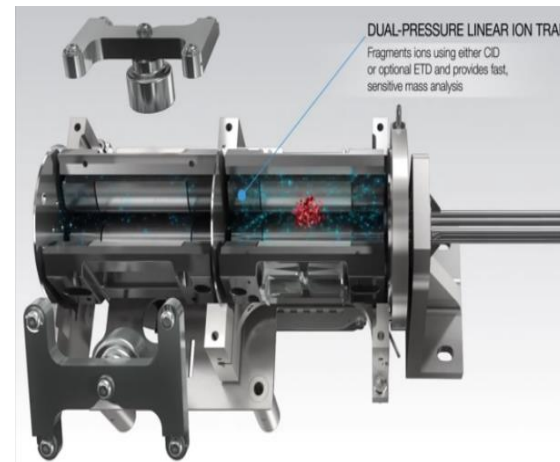
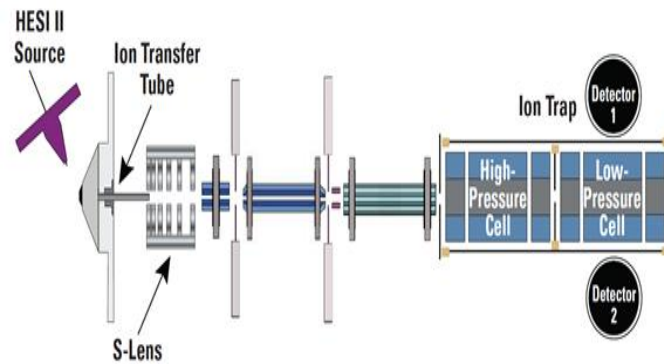
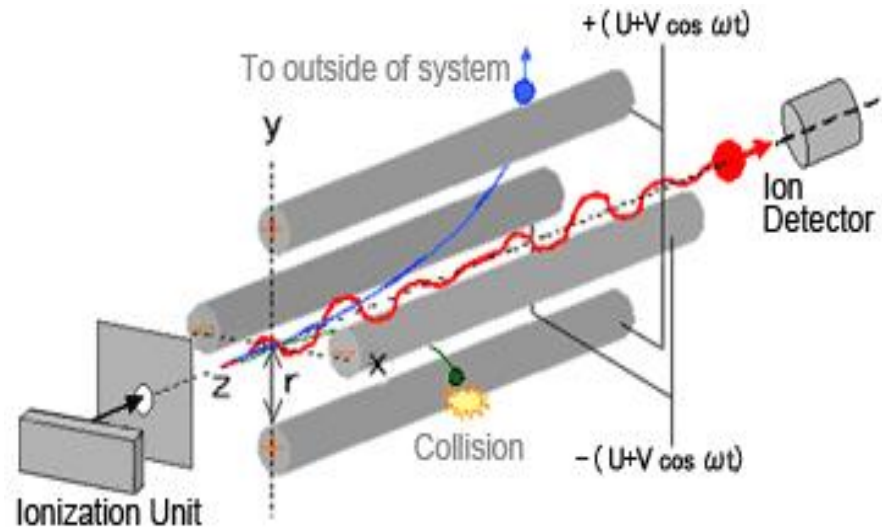
1.000	linear TOF w/o DE
5.000	reflector TOF w/o DE
25.000	reflector TOF with DE
125.000	FTMS wideband mode
1.000.000	FTMS high-res mode

Natural abundance of atoms isotopes in proteins

Name	Symbol	Mass (Da)	Abundance (%)
Hydrogen	H	1.007825	99.9885
Deuterium	H	2.014102	0.0115
Carbon	C	12.000000	98.9300
	C	13.003355	1.0700
Nitrogen	N	14.003074	99.6320
	N	15.000109	0.3680
Oxygen	O	15.994915	99.7570
	O	16.999132	0.0380
	O	17.999160	0.2050
Phosphorus	P	30.973762	100.0000
Sulfur	S	31.973762	94.9300
	S	32.971458	0.7600
	S	33.967867	4.2900
	S	35.967081	0.0200

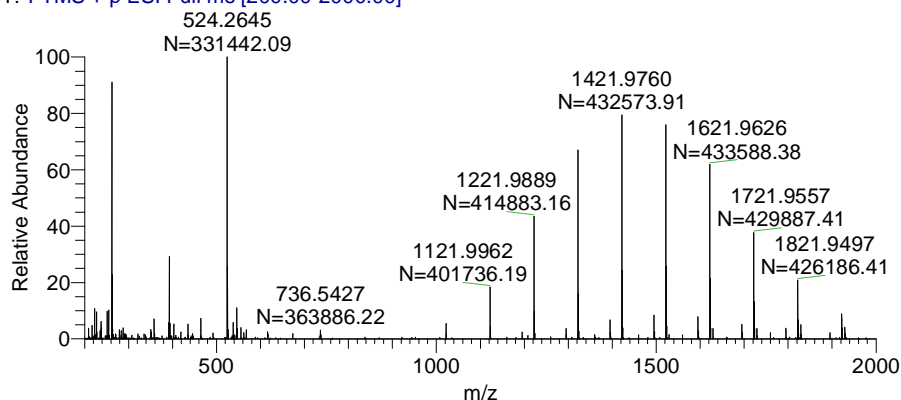


Different instrumental design



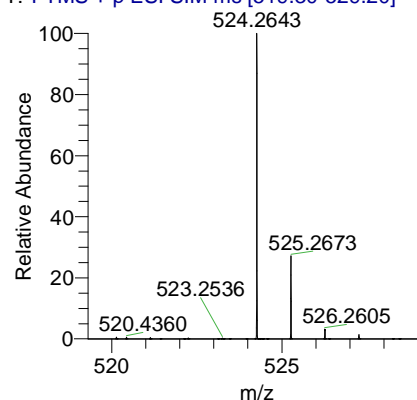
Isolation Width: Full MS to 1 amu (MRFA)

110511_Transmission_comp_all_1e5_01 #11 RT: 0.05 AV: 1 NL: 1.33E8
T: FTMS + p ESI Full ms [200.00-2000.00]



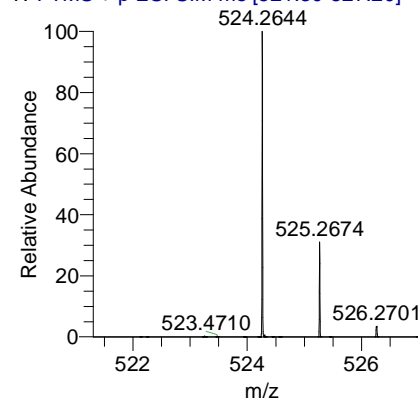
IsoW= 1800

110511_Transmission_comp_all_1e5_01 #70
T: FTMS + p ESI SIM ms [519.30-529.20]



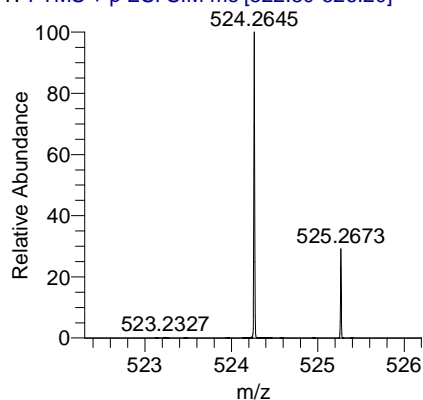
IsoW= 10

110511_Transmission_comp_all_1e5_01 #109
T: FTMS + p ESI SIM ms [521.30-527.20]



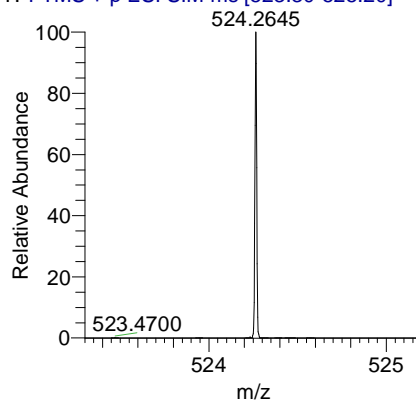
IsoW= 6

110511_Transmission_comp_all_1e5_01 #160
T: FTMS + p ESI SIM ms [522.30-526.20]



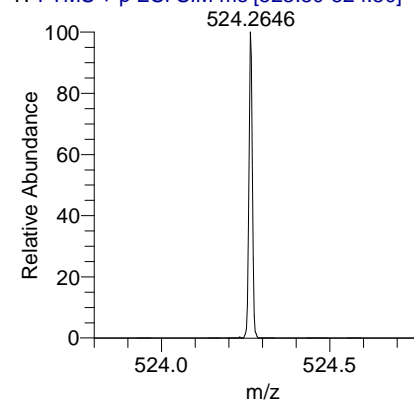
IsoW= 4

110511_Transmission_comp_all_1e5_01 #209
T: FTMS + p ESI SIM ms [523.30-525.20]



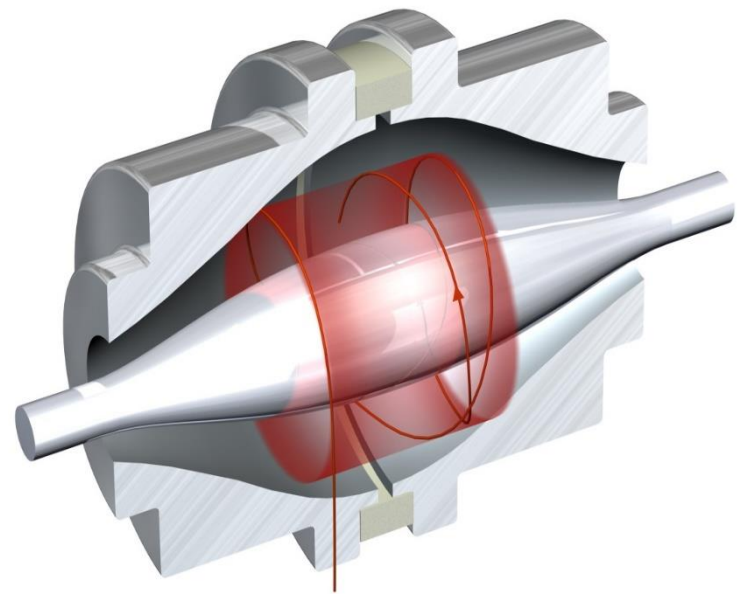
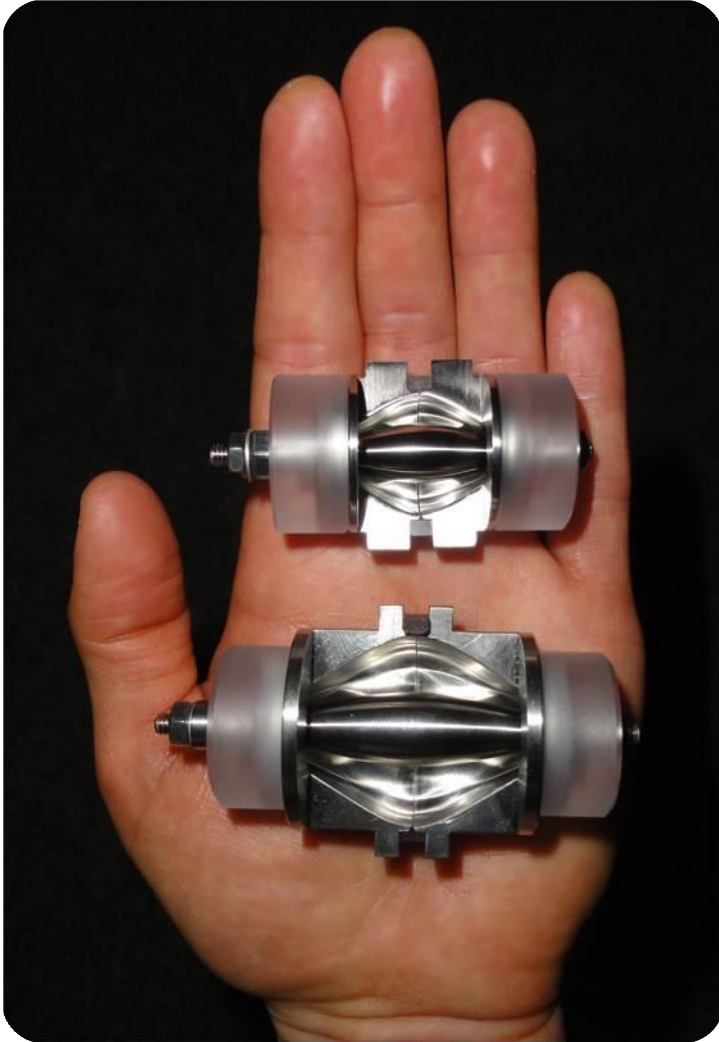
IsoW= 2

110511_Transmission_comp_all_1e5_01 #245
T: FTMS + p ESI SIM ms [523.80-524.80]



IsoW= 1

the orbitrap cell



$$\omega_z = \sqrt{\frac{k}{m/z}}$$

Developpement of the orbitrap family

2007

LTQ Orbitrap XL and Discovery

2008

LTQ Orbitrap XL ETD

2009

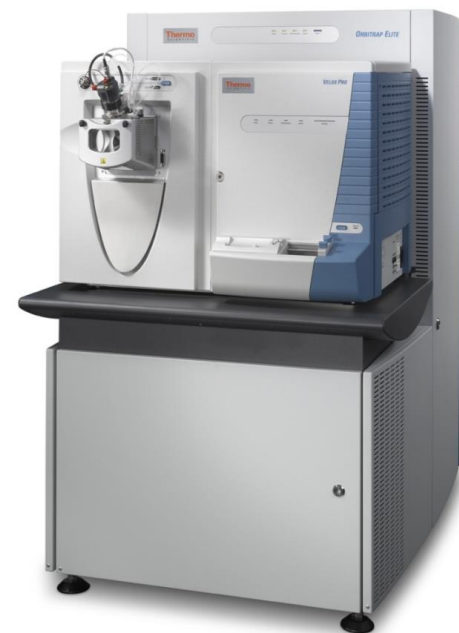
LTQ Orbitrap Velos

2011

LTQ Orbitrap Velos Pro
Orbitrap Elite

2013

Orbitrap Fusion Tribrid



Instrumentations

Micro/nanoHPLC



U3000 micro



EASY-nLC 1000



U3000 nano



EASY-nLC II

MALDI



4800 TOF-TOF

ESI



ORBITRAP-Velos-ETD:
Routine



Qexactive+:
Routine
Quantification

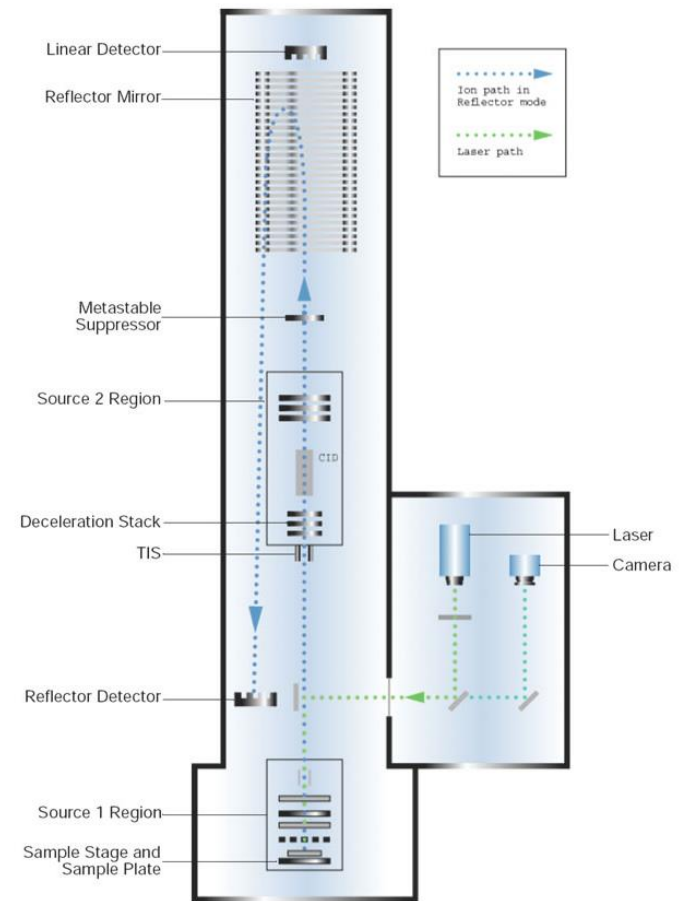
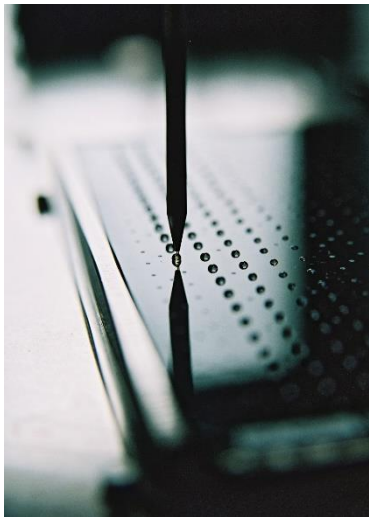


Fusion Tribrid:
TMT Quantification ciblée
R&D

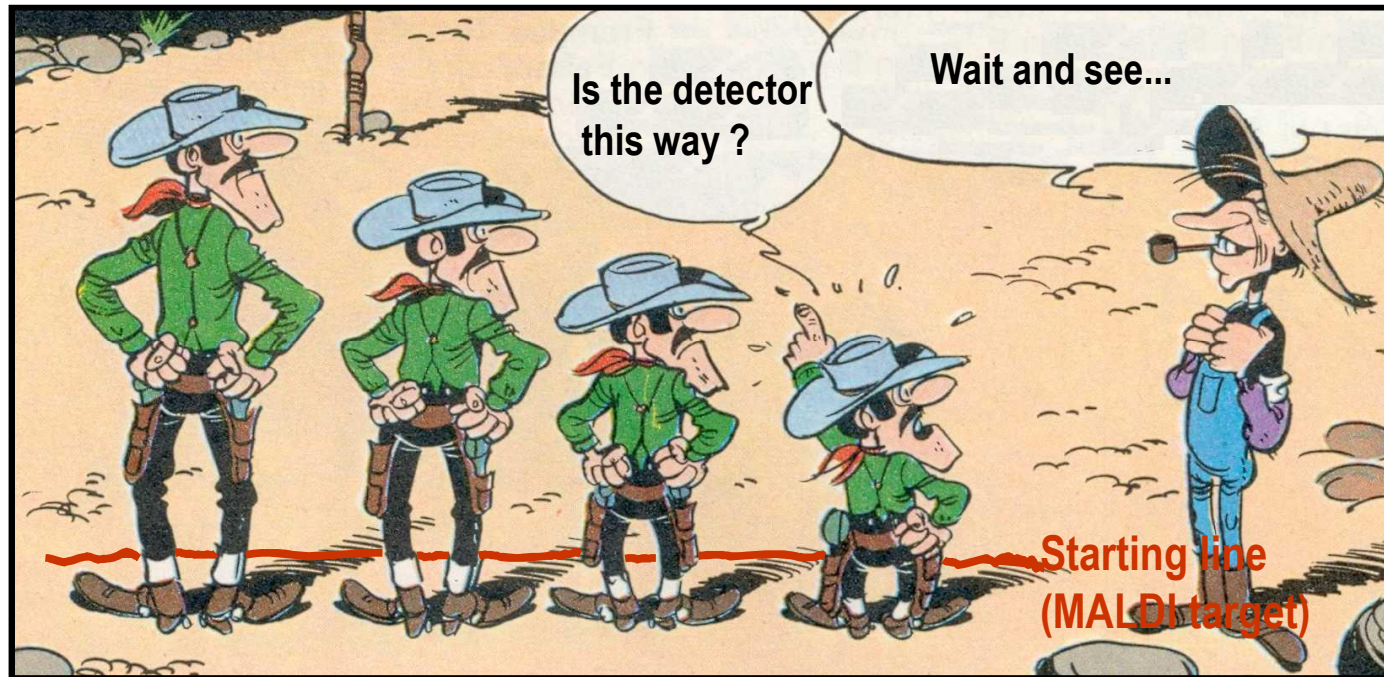


NanoMate:
Infusion

MALDI-TOF/TOF



Time of flight – principles (TOF)

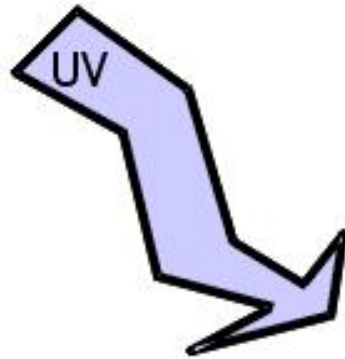


Remember : Mass of an ion is measured in the Dalton units !

Start !

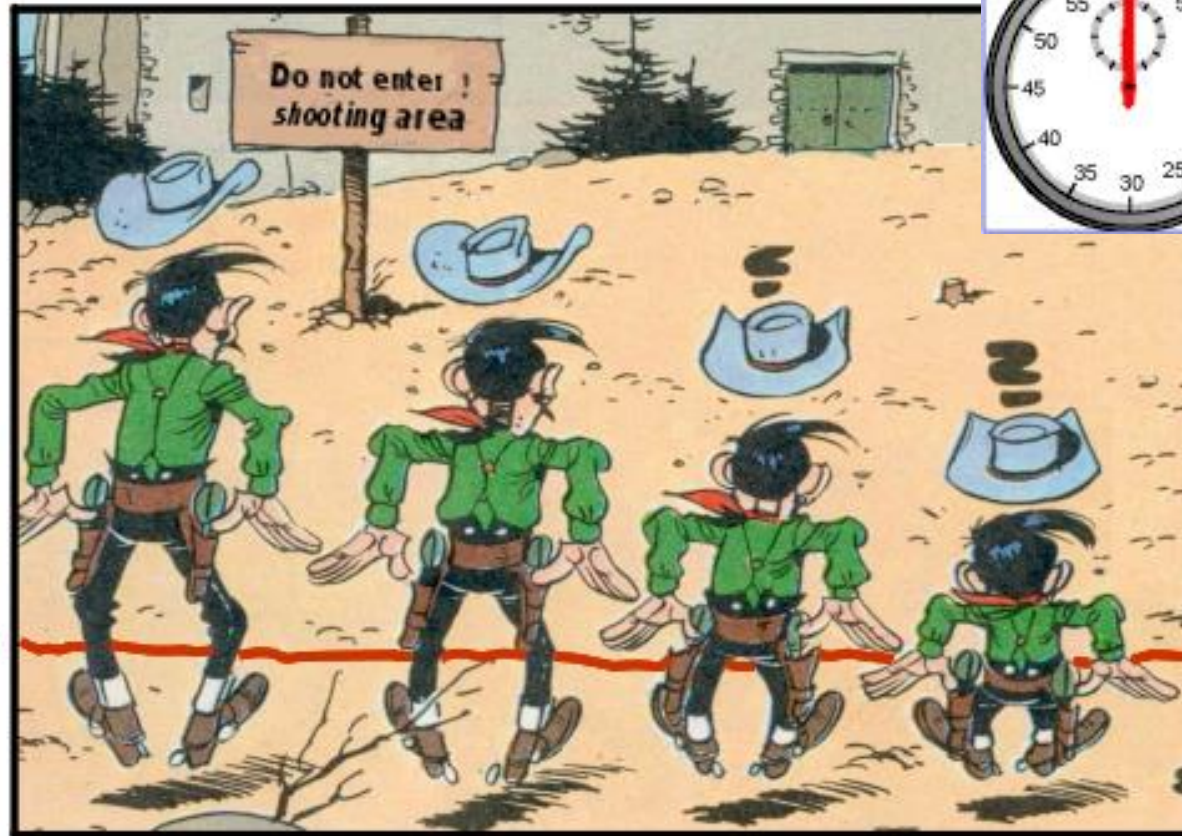
Laser

The desorptionevent



Induced by the laser impuls

Start →



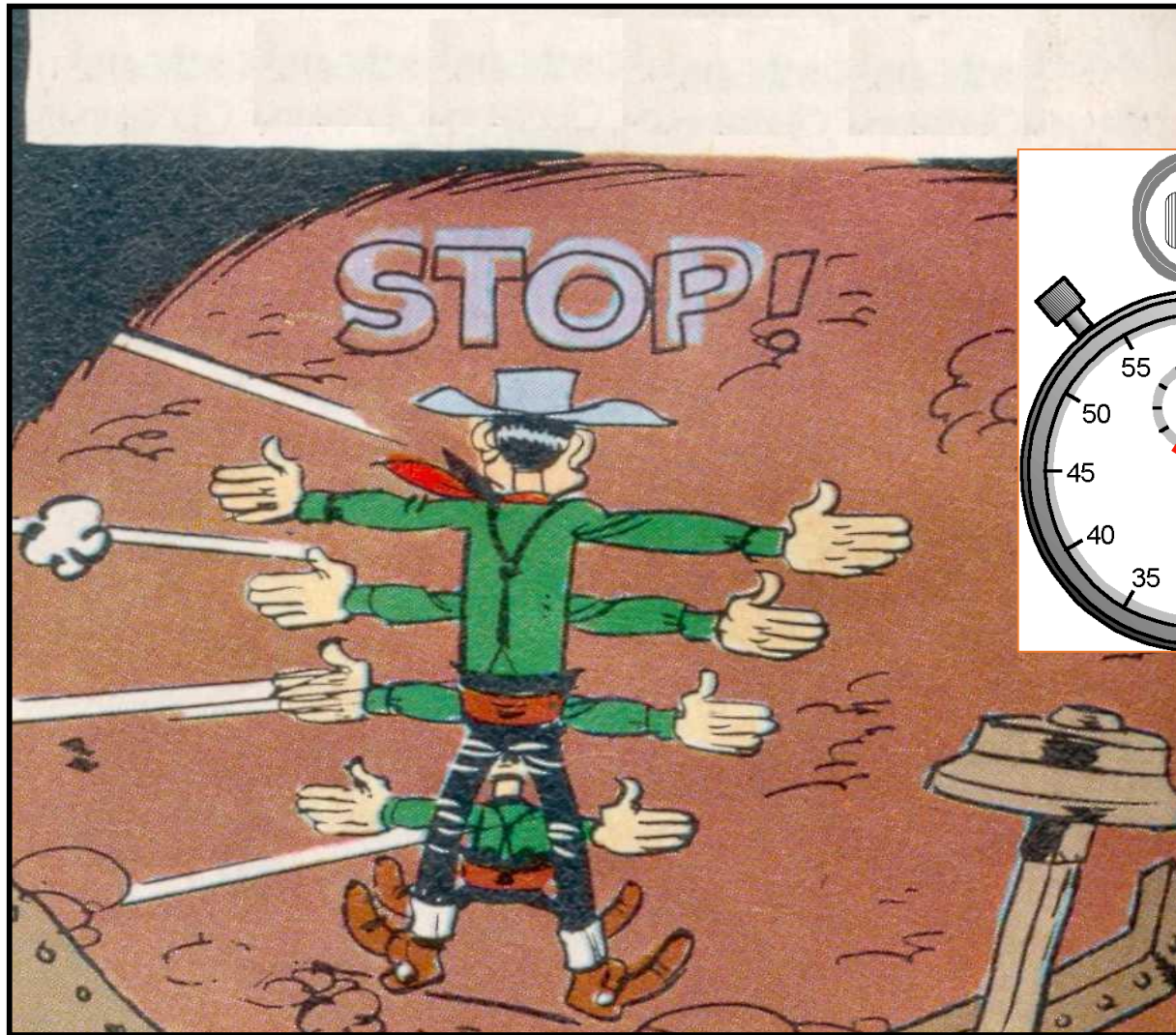
Ions in the time of flight (TOF)



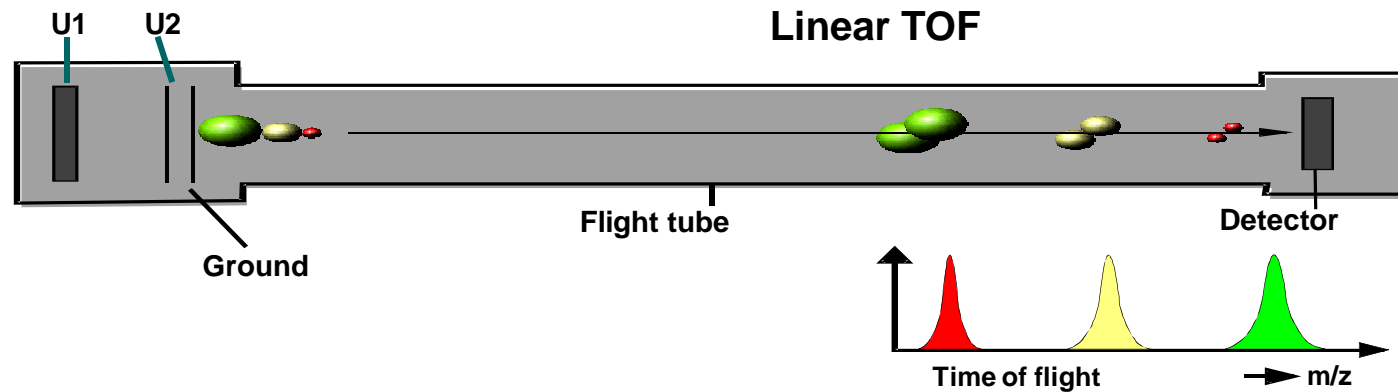
Increasing MW



Ions in the detector



Ions analysis in linear mode



Electric field : $E_c = qU = 1/2 mV^2$ Identical for all ions

($V=L/t$ L : tube length)

⇒ Simple relation $t^2 = mL^2/2qU = \text{Constante} \times m/z$

Light Corrections :

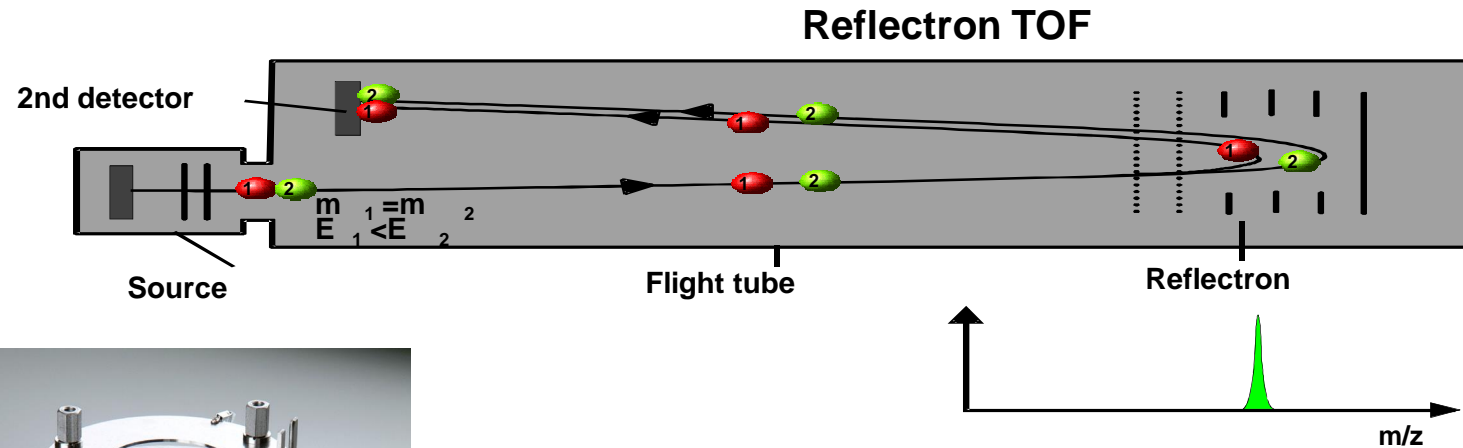
$$t^2 = Am^2 + Bm + C$$

(A : initial desorption E_c

C : Extraction Delay)

⇒ Simple Quadratic equation

Ions analysis in reflectron mode



Reflectron : 2 effects on resolution :

- Increasing flight path (better separation of particles of different masses) (equivalent 3m flight tube)
- Focusing effect for particles with same mass

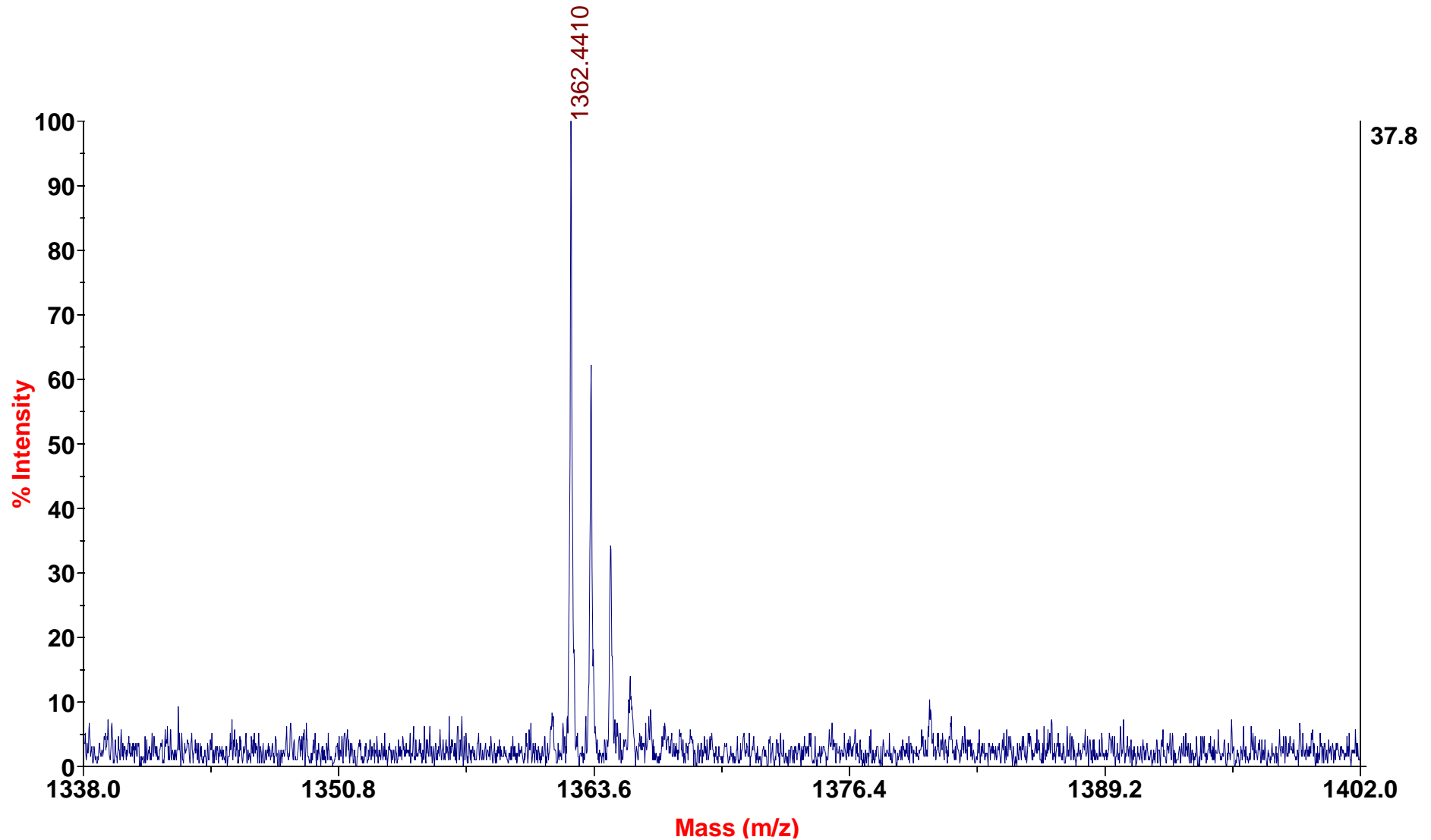
Key questions in proteomics

- “ What is the protein content of my biological sample?
=> problem of **identification**
- “ What is the abundance of my protein of interest?
=> quantification problem
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ What are the partners of my protein of interest?
- “ Are there any signature proteins related to a particular biological process?

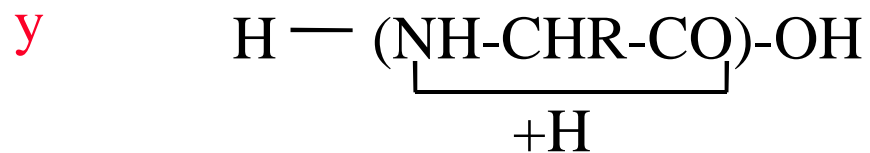
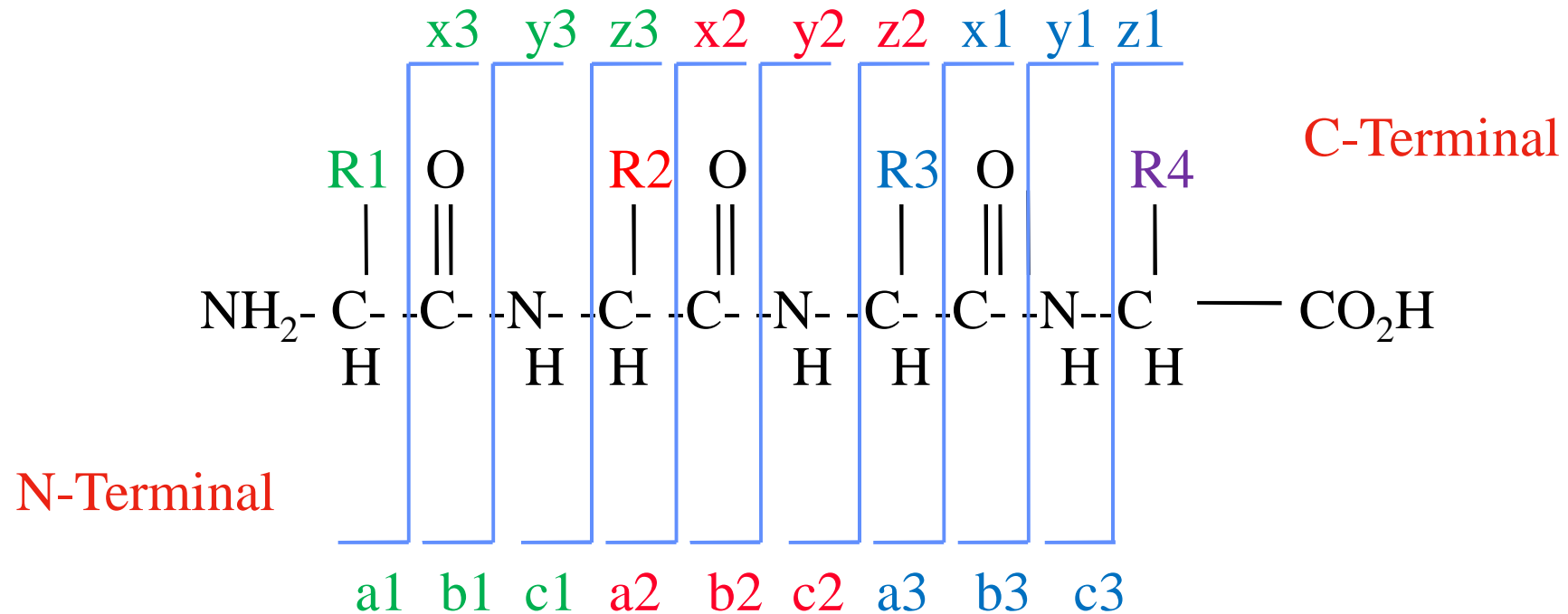
=> biomarkers identifications and quantifications

Ion precursor selection

4700 Reflector Spec #1 MC[BP = 1664.6, 132]



MS/MS fragmentation for peptides



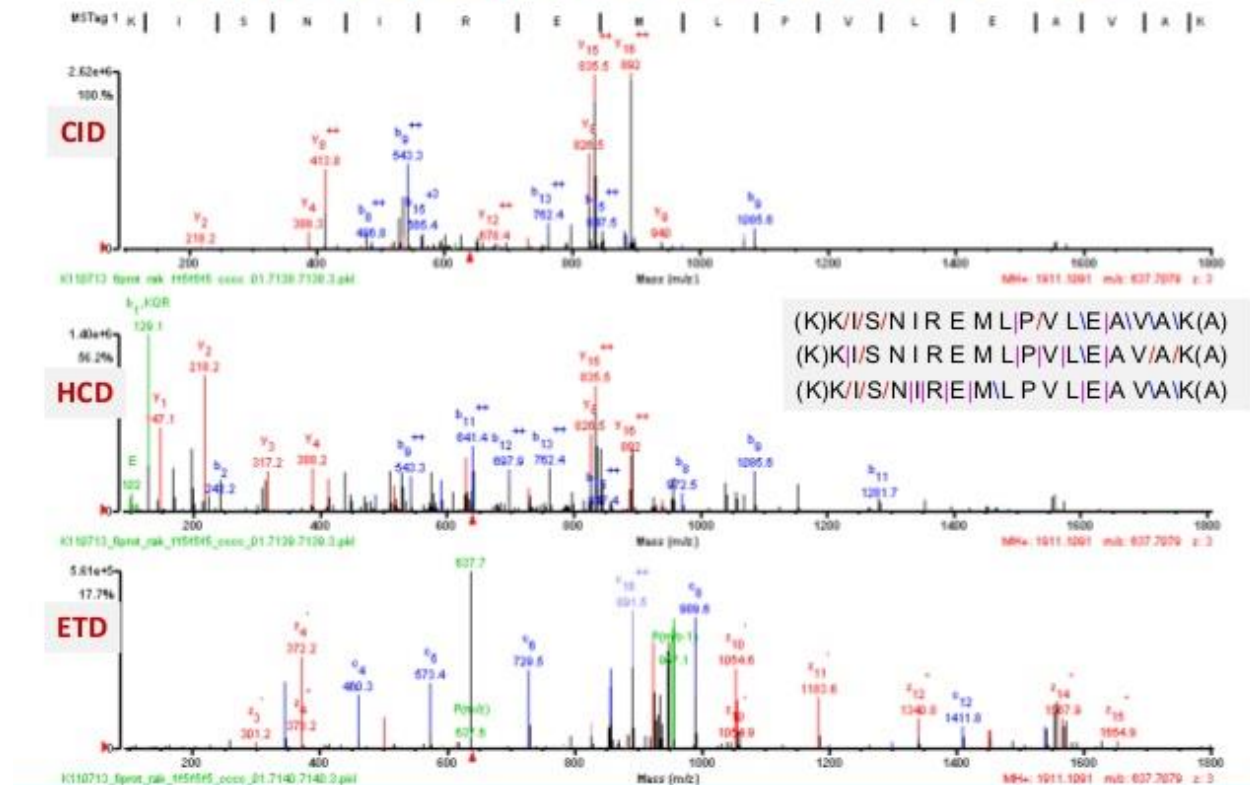
MS/MS fragmentation for peptides

DISSOCIATION INDUITE PAR COLLISION (CID)

HIGHER ENERGY COLLISIONAL DISSOCIATION (HCD)

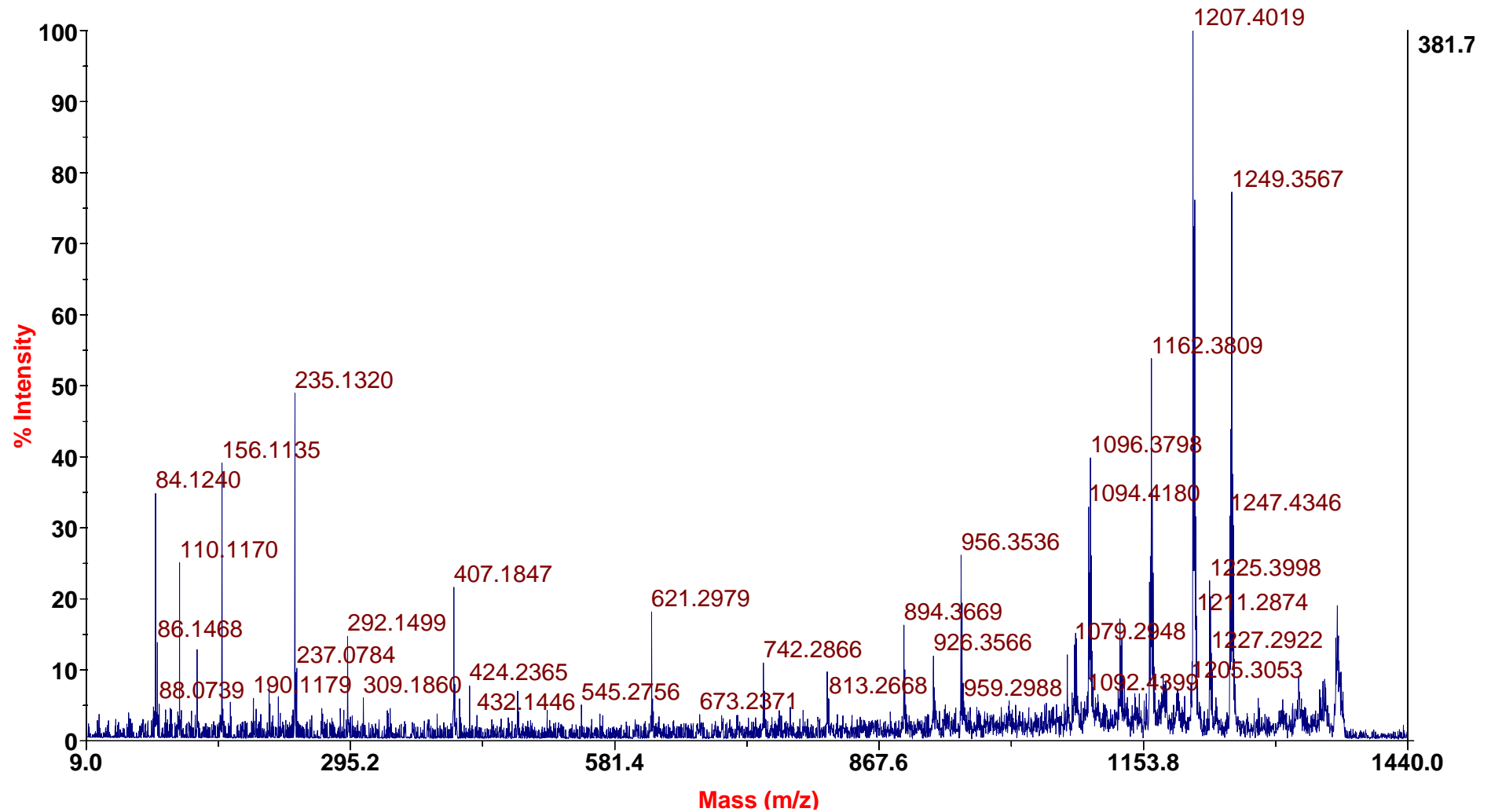
ELECTRON TRANSFER DISSOCIATION (ETD)

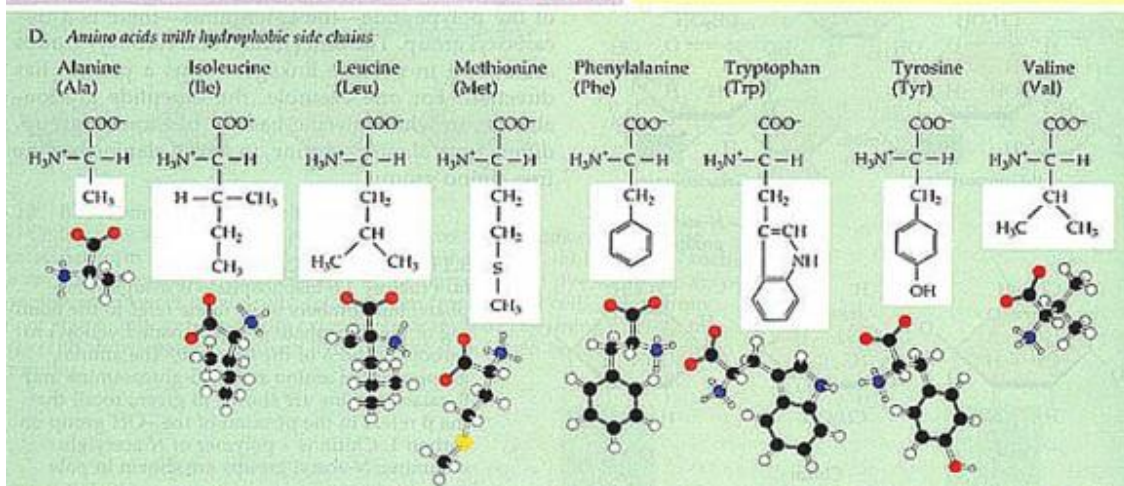
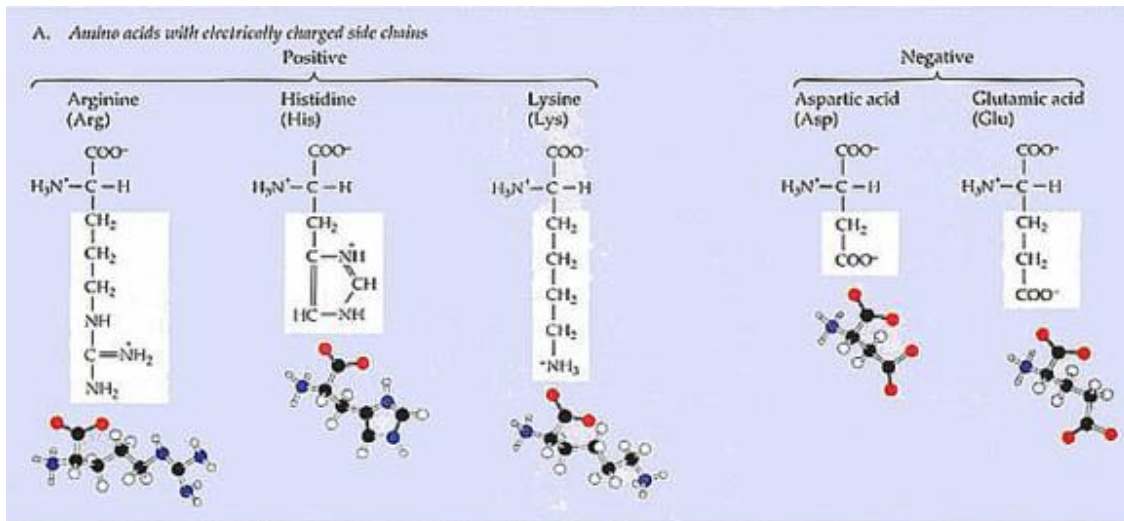
CID/HCD/ETD triplets on same precursor $z=3$



MS/MS spectrum of the precursor 1362.44 m/z

4700 MS/MS Precursor 1362.44 Spec #1 MC[BP = 1207.3, 382]





Alanine	A, Ala	71.079
Arginine	R, Arg	156.188
Asparagine	N, Asn	114.104
Aspartic acid	D, Asp	115.089
Cysteine	C, Cys	103.145
Glutamine	Q, Gln	128.131
Glutamic acid	E, Glu	129.116
Glycine	G, Gly	57.052
Histidine	H, His	137.141
Isoleucine	I, Ile	113.160
Leucine	L, Leu	113.160
Lysine	K, Lys	128.17
Methionine	M, Met	131.199
Phenylalanine	F, Phe	147.177
Proline	P, Pro	97.117
Serine	S, Ser	87.078
Threonine	T, Thr	101.105
Tryptophan	W, Trp	186.213
Tyrosine	Y, Tyr	163.176
Valine	V, Val	99.133

MS/MS spectra interpretation

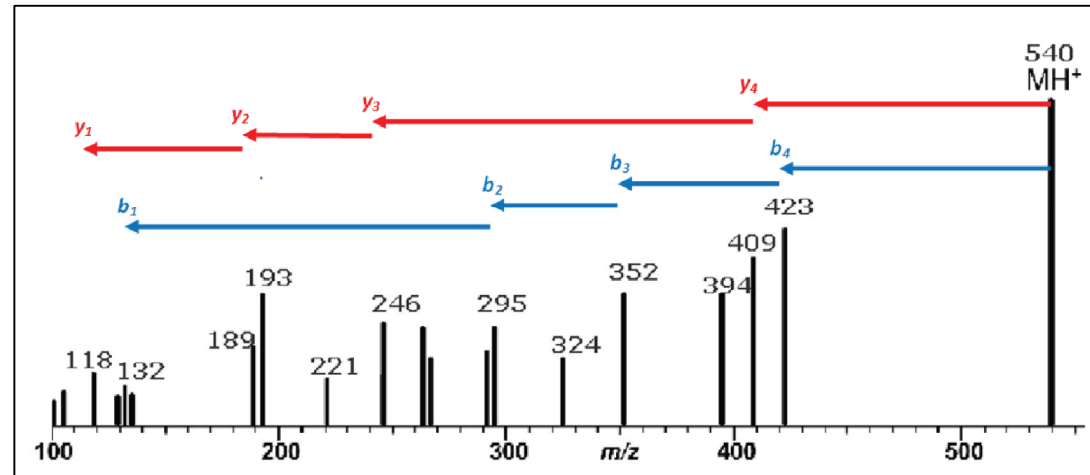


Table 1

Ion	m/z	Neutral loss (from previous ion in the series)	Amino Acid Residue
Precursor $[M+H]^+$	540		
y_4	409	131	M
y_3	246	163	Y
y_2	189	57	G
y_1	118	71	A
b_4	423	117 (99+18)	V
b_3	352	71	A
b_2	295	57	G
b_1	132	163	Y
a_4	395?		
a_3	324		
a_2	267		
a_1	104		

MYGAV

User AA Formula 1: C2 H3 N1 O1

Elemental Composition: C24 H38 N5 O7 S1

MH+1(av) MH+1(mono)

540.6627 540.2486

[–] Main Sequence Ions

b			y	
---	1	M	5	---
295.1111	2	Y	4	409.2082
352.1326	3	G	3	246.1448
423.1697	4	A	2	189.1234
---	5	V	1	118.0863

Current post-translational modifications (PTMs)

Acids & amides (E/D/Q/N)	Pyroglutamic acid (Q)	-17.0306	Deamidation (Q/N)	+0.9847
	Carboxylation (E/D)	+44.0098		

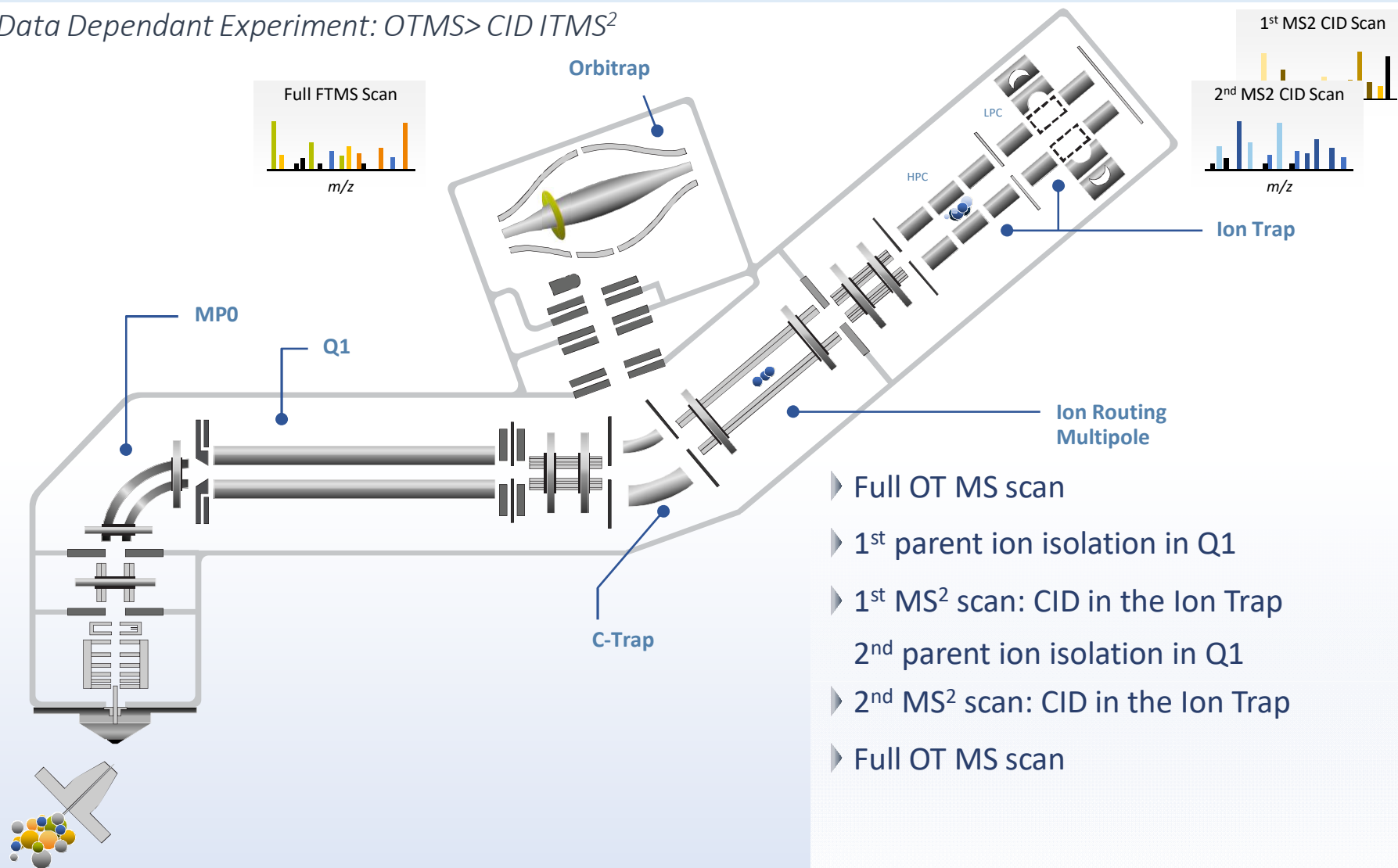
Hydroxyl groups (S/T/Y)	Phosphorylation	+79.9799	Sulphation	+80.0642
------------------------------------	-----------------	----------	------------	----------

Carbohydrates (S/T/N)	Pentoses	+132.1161	Deoxyhexoses	+146.1430
	Hexosamines	+161.1577	Hexoses	+162.1424
	N-acetylhexosamines	+203.1950	Sialic acid	+291.2579

Sulphydryls (C)	Disulphide bond	-2.0159	Oxidation	+15.9994
	Cysteinylation	+119.1442	Glutathionylation	+305.3117

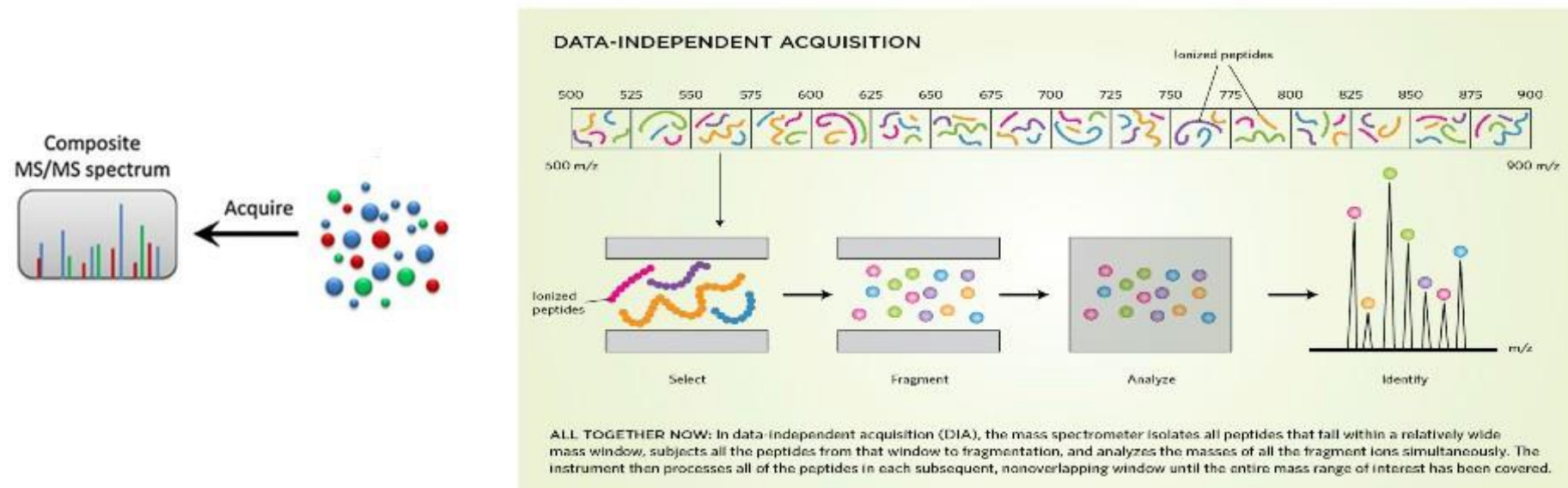
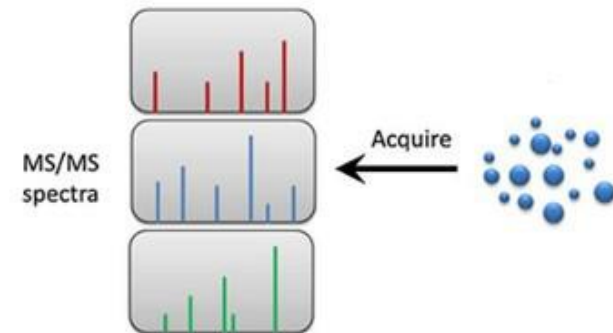
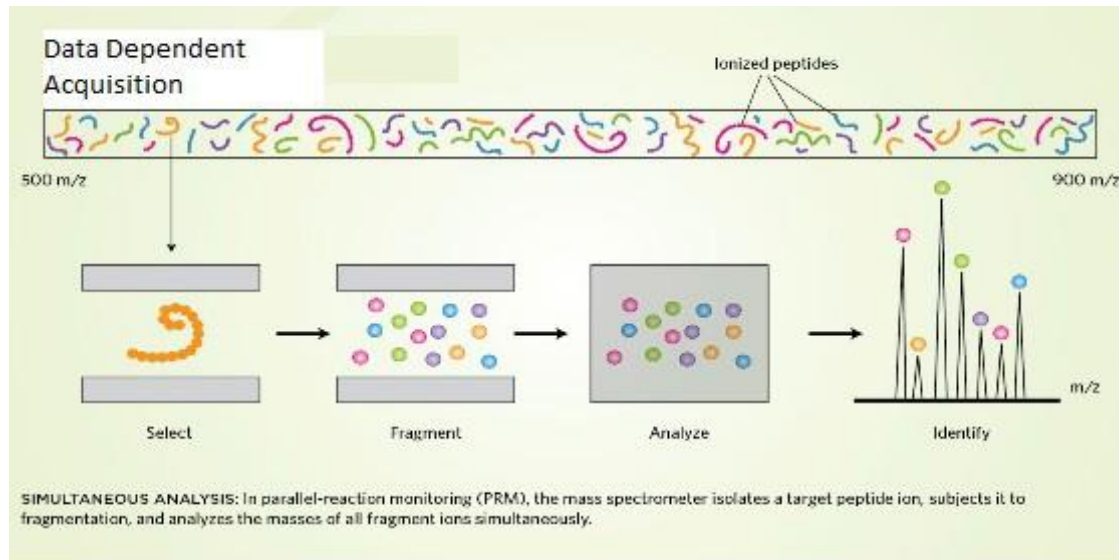
MS and MS/MS spectra generation

Data Dependant Experiment: OTMS > CID ITMS²



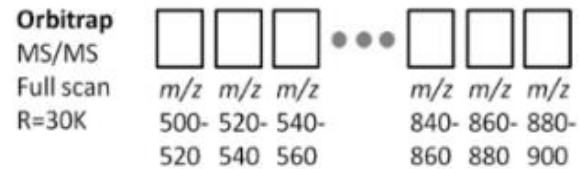
- ▶ Full OT MS scan
- ▶ 1st parent ion isolation in Q1
- ▶ 1st MS² scan: CID in the Ion Trap
- ▶ 2nd parent ion isolation in Q1
- ▶ 2nd MS² scan: CID in the Ion Trap
- ▶ Full OT MS scan

DDA versus DIA

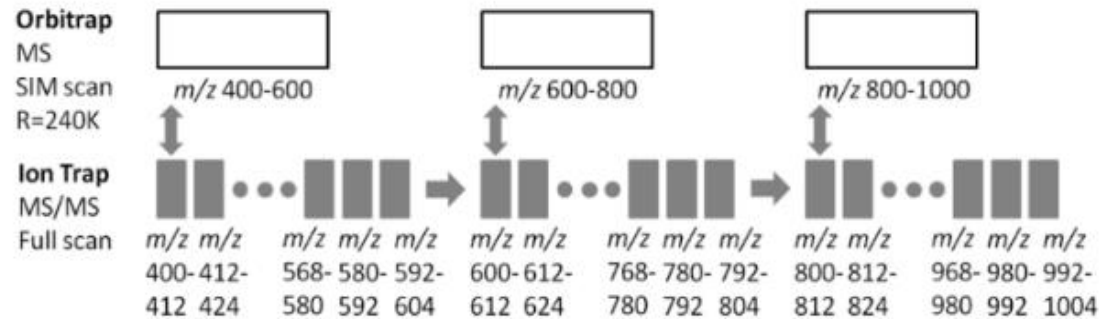


Data Independent Acquisition: DIA

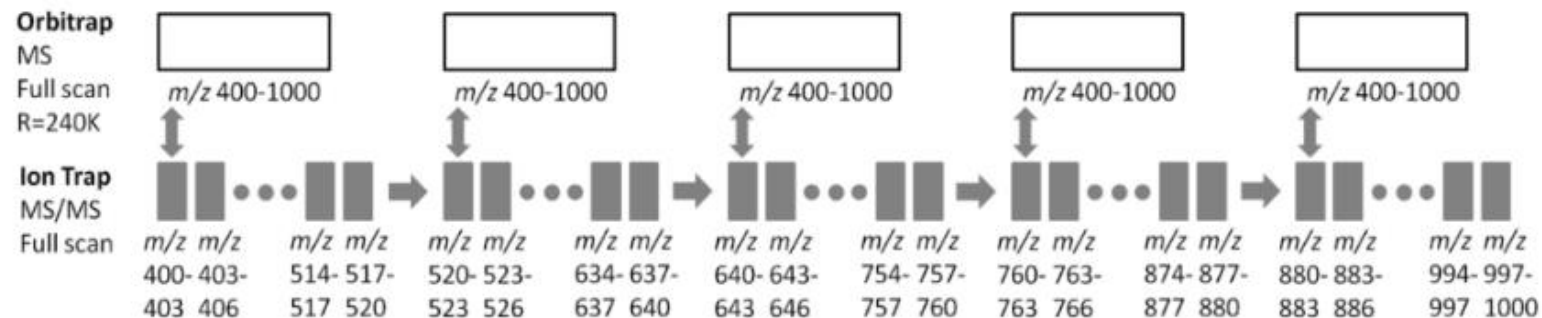
(A) DIA



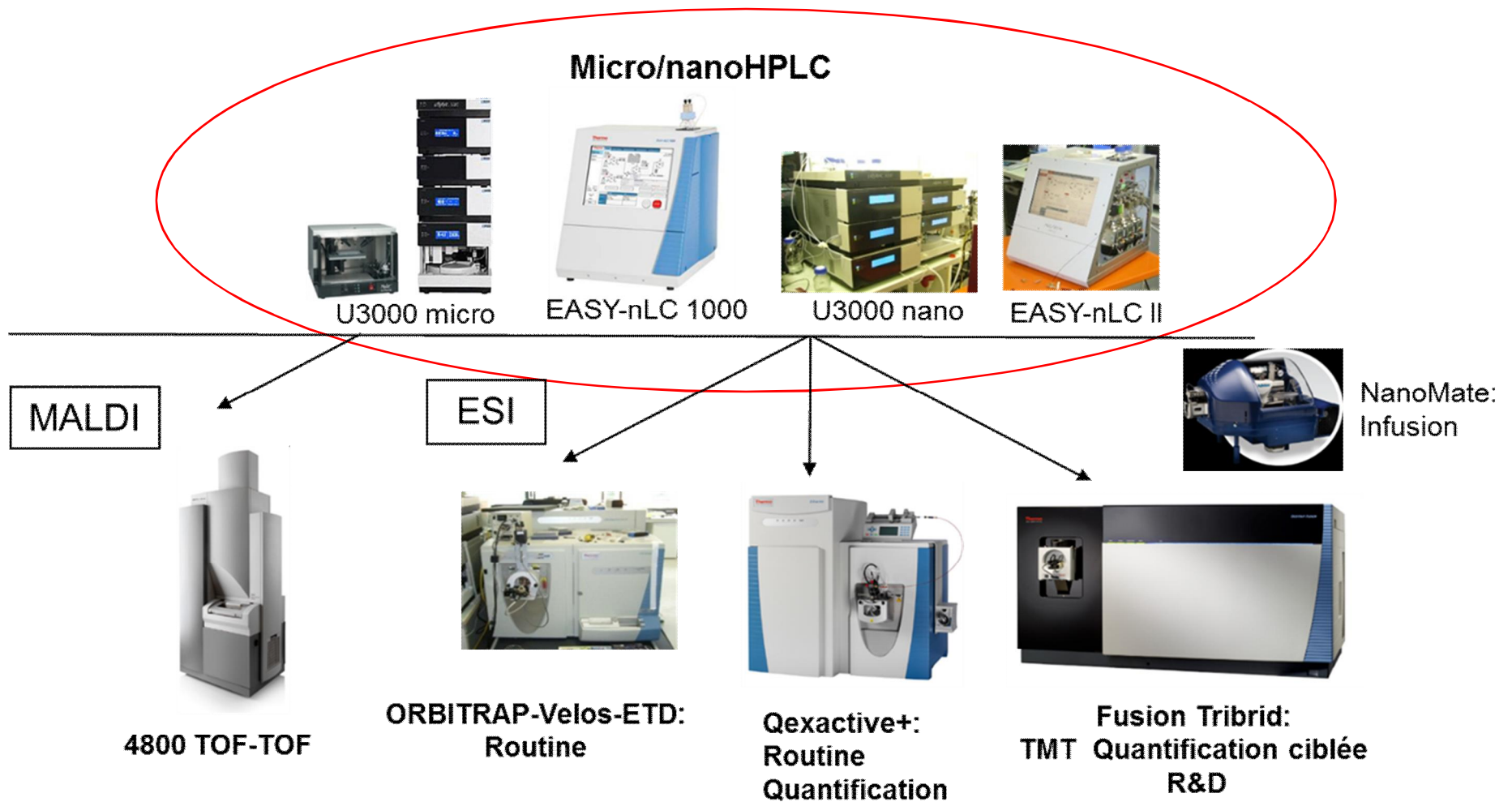
(B) WiSIM-DIA



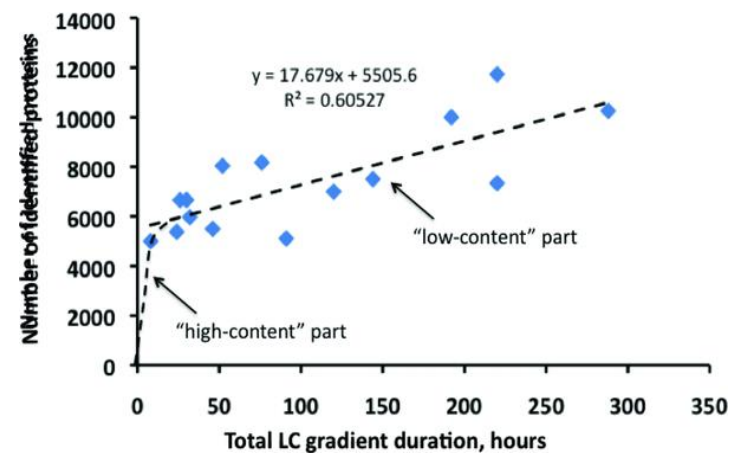
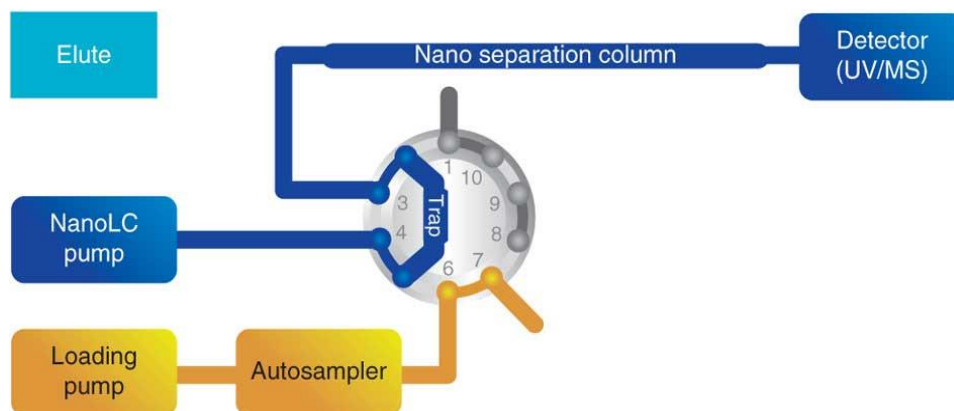
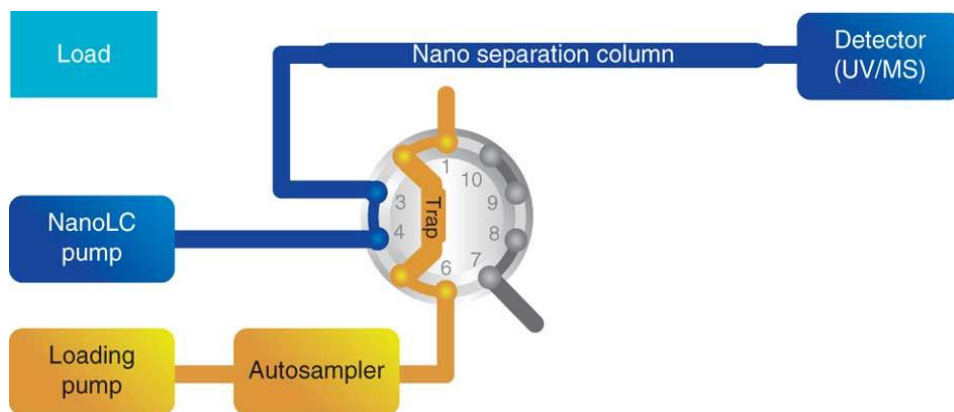
(C) Full MS-DIA



Contribution of nano-HPLC



Peptides separation by nano-LC

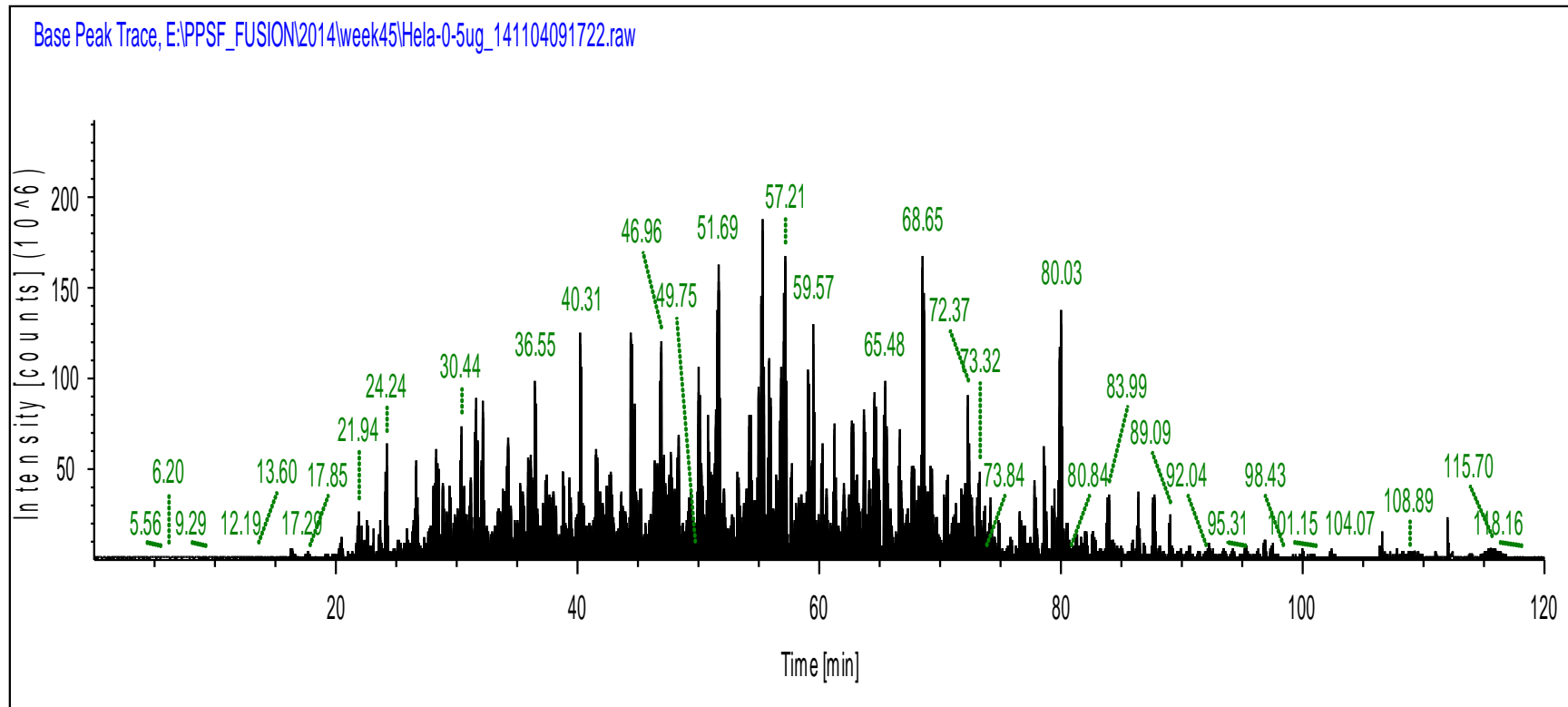


Peptides nano-LC separation

- It is impossible to resolve all species in a proteomics sample using only one separation method
- Multidimensional separation - two or more independent (orthogonal) separation techniques coupled together for the analysis of a single sample.

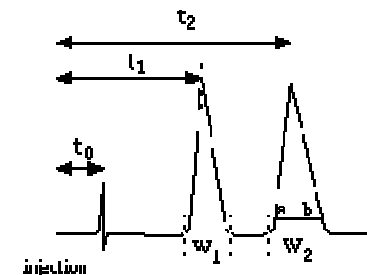
Separation method	Separation by:
Reversed phase	Hydrophobicity
Ion exchange, IsoElectroFocusing (IEF)	Net charge, Isoelectric point
Size exclusion, SDS Gel Electrophoresis	Size, molecular weight
Affinity chromatography	Specific functional groups

Total ion current (TIC) HeLa tryptic digest (0.5 μg of total proteins)



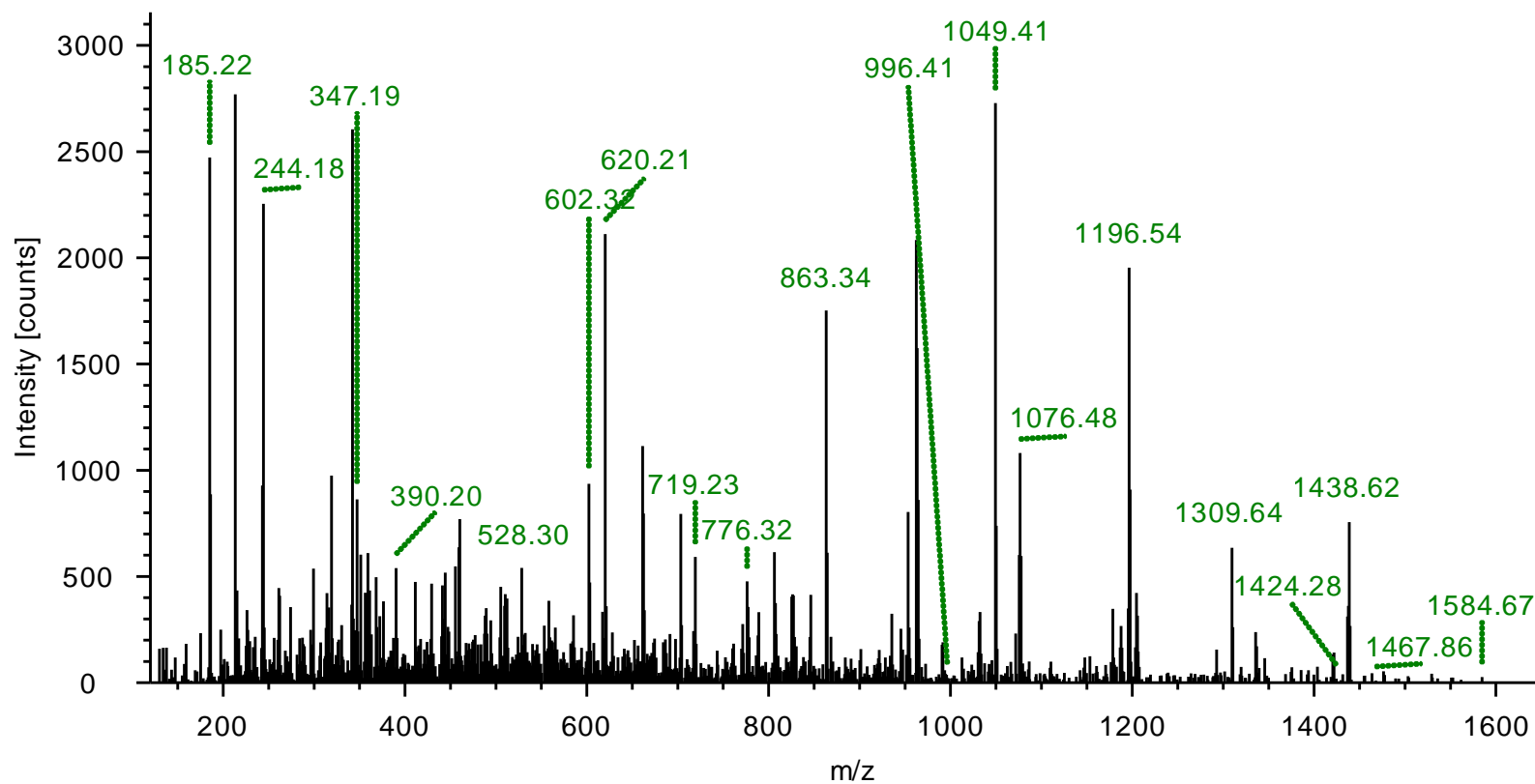
$$R = \frac{2(t_2 - t_1)}{(w_2 - w_1)}$$

with t_1 and t_2 the retention time and w_1 and w_2 peak widths at mid-height



90000 MSMS in 2h gradient (C18 RPC)

E:\PPSF_FUSION2014\week45\Hela-0-5ug_141104091722.raw #48756, RT=63.18 min
ITMS, HCD, Precursor: z=+2, Mono m/z=825.90680 Da, MH+=1650.80632 Da



Proprietary MS data formats

Company	Extension	File type
Agilent	.D (folder)	Agilent MassHunter, Agilent ChemStation, or Bruker BAF/YEP/TDF data format
Bruker	.YEP	instrument data format
Agilent/Bruker	.BAF	instrument data format
Bruker	.FID	instrument data format
Bruker	.TDF	timsTOF instrument data format
ABI/Sciex	.WIFF	instrument data format
ABI/Sciex	.t2d	4700 and 4800 file format
Waters	.PKL	MassLynx peak list format
Thermo	.RAW*	Thermo Xcalibur
PerkinElmer	.RAW* (folder)	PerkinElmer TurboMass
Micromass**/Waters		Waters MassLynx
Chromtech		Finnigan ITDS file format; MAT95 instrument data format
Finnigan***	.DAT	MassLab data format
VG		ITS40 instrument data format
Finnigan***	.MS	GCMSSolution format
Shimadzu	.QGD	instrument data format
Shimadzu	.qgd	instrument data format
Shimadzu	.lcd	QQQ/QTOF instrument data format
Shimadzu	.spc	library data format
Bruker/Varian	.SMS	instrument data format
Bruker/Varian	.XMS	instrument data format
ION-TOF	.itm	raw measurement data
ION-TOF	.ita	analysis data
Physical Electronics/ULVAC-PHI	.raw*	raw measurement data
Physical Electronics/ULVAC-PHI	.tdc	spectrum data

Open MS data formats

JCAMP-DX

This format was one of the earliest attempts to supply a standardized file format for data exchange in mass spectrometry. [JCAMP-DX](#) was initially developed for infrared spectrometry. JCAMP was officially released in 1988. JCAMP was found impractical for today's large MS data sets, but it is still used for exchanging moderate numbers of spectra.

ANDI-MS or netCDF

The Analytical Data Interchange Format for Mass Spectrometry is a format for exchanging data. ANDI was initially developed for chromatography-MS data and therefore was not used in the [proteomics](#) gold rush where new formats based on [XML](#) were developed.

mzData

mzData was the first attempt by the [Proteomics Standards Initiative](#) (PSI) from the [Human Proteome Organization](#) (HUPO) to create a standardized format for Mass Spectrometry data. This format is now deprecated, and replaced by mzML.

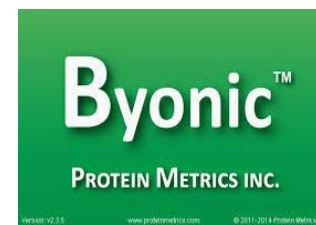
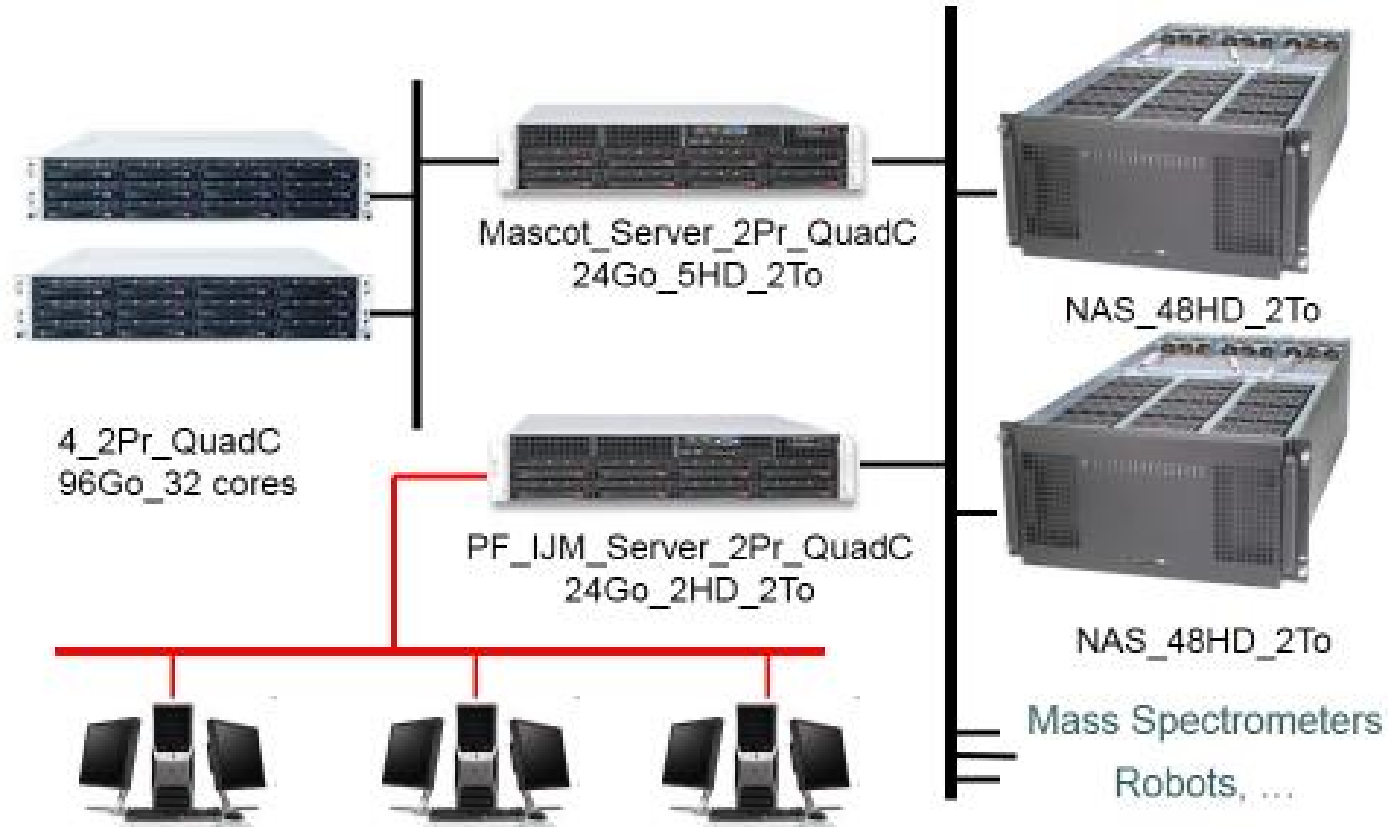
mzXML

mzXML is a [XML](#) (eXtensible Markup Language) based common file format for [proteomics](#) mass spectrometric data. This format was developed at the Seattle Proteome Center/Institute for Systems Biology while the HUPO-PSI was trying to specify the standardized mzData format, and is still in use in the proteomics community.

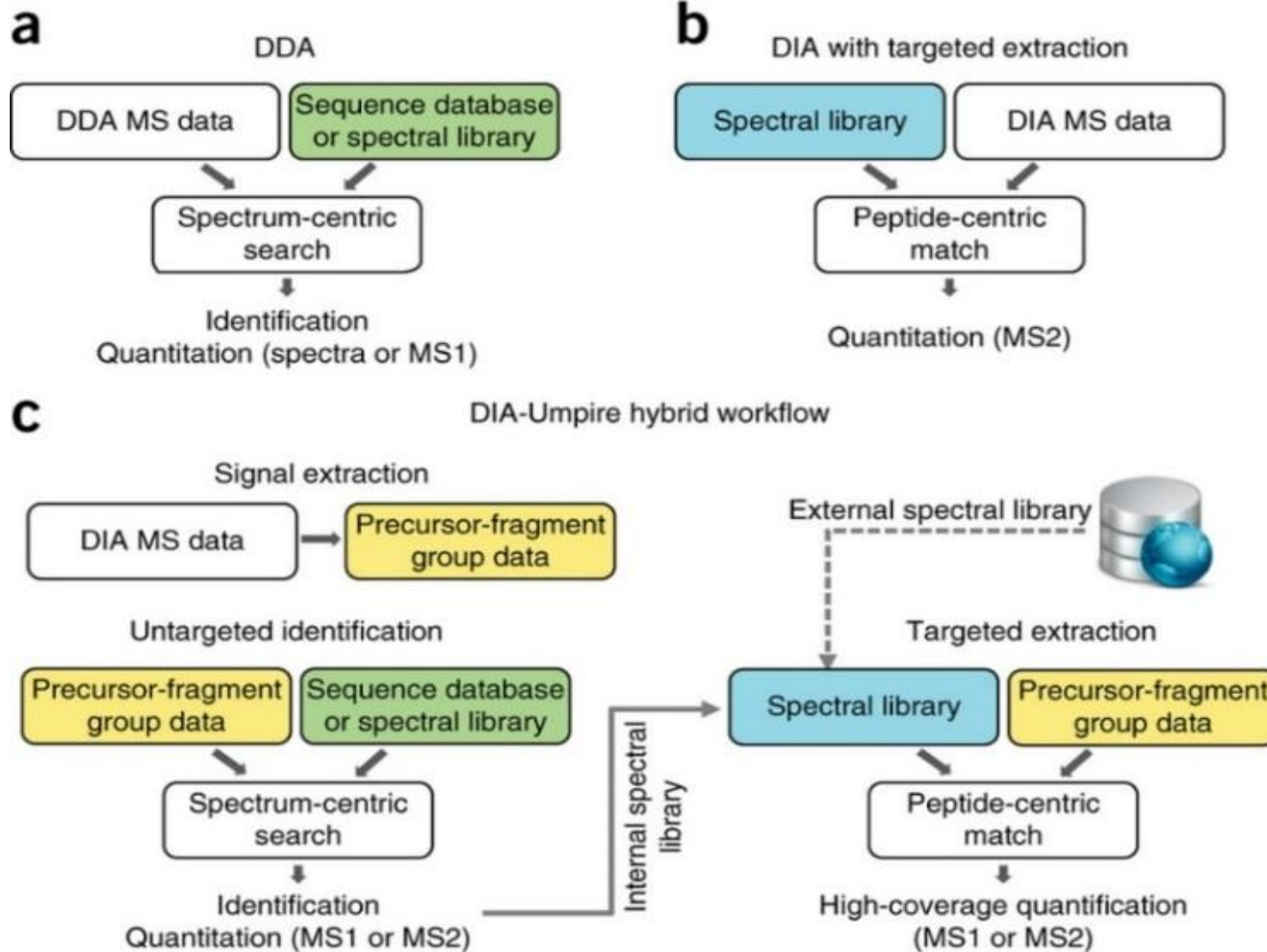
mzML

As two formats (mzData and mzXML) for representing the same information is an undesirable state, a joint effort was set by HUPO-PSI, the SPC/ISB and instrument vendors to create a unified standard borrowing the best aspects of both mzData and mzXML, and intended to replace them. The first specification was published in June 2008. This format was officially released at the 2008 [American Society for Mass Spectrometry](#) Meeting, and is since then relatively stable with very few updates. On 1 June 2009, mzML 1.1.0 was released. There are no planned further changes as of 2013.

Saving data and servers



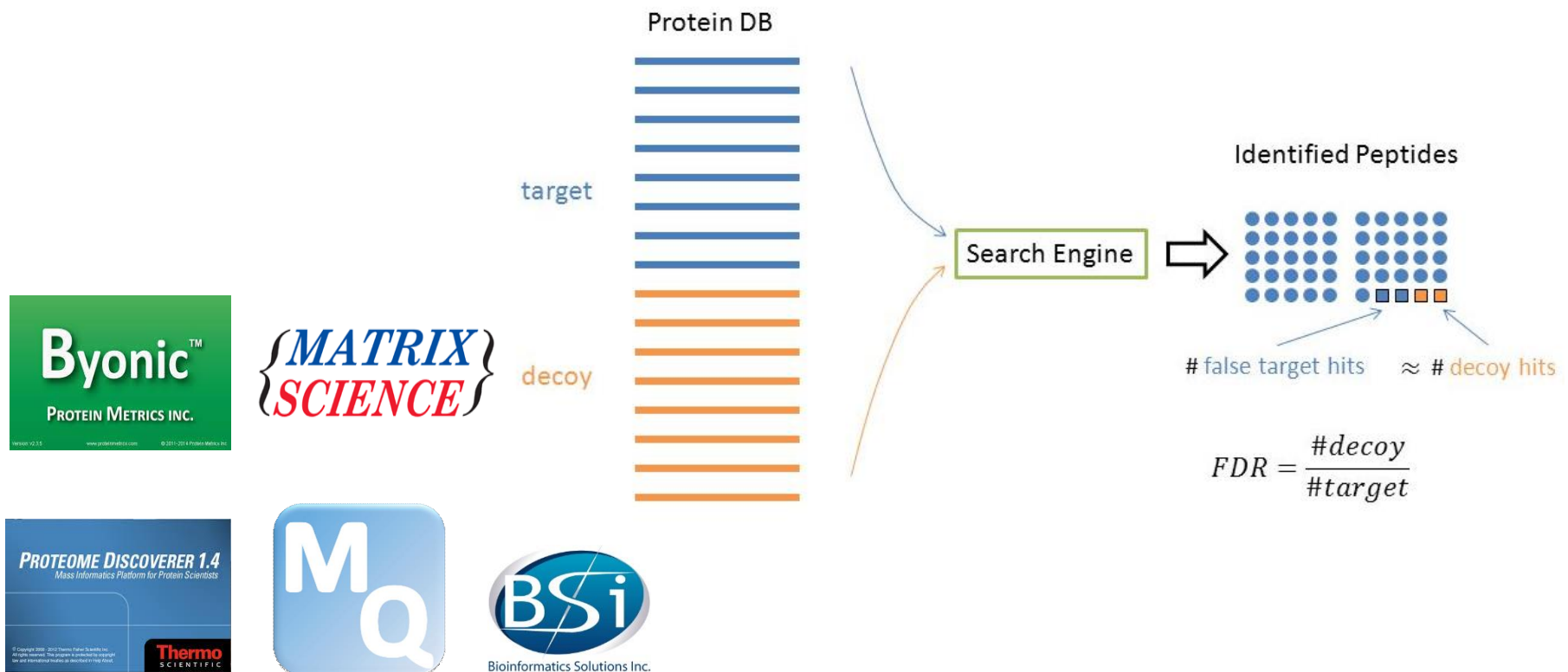
Search engine



Search engines and validation of peptides and proteins identifications

$$FDR (\%) = \frac{(\# \text{ false target hits}) \times 2}{(\# \text{ false target hits}) + (\# \text{ target hits})}$$

FDR Estimation



Critical importance of mass accuracy for database searches

Expressed as Da or as ppm (10 ppm = 0,001% 1 ppm = 0,0001%)

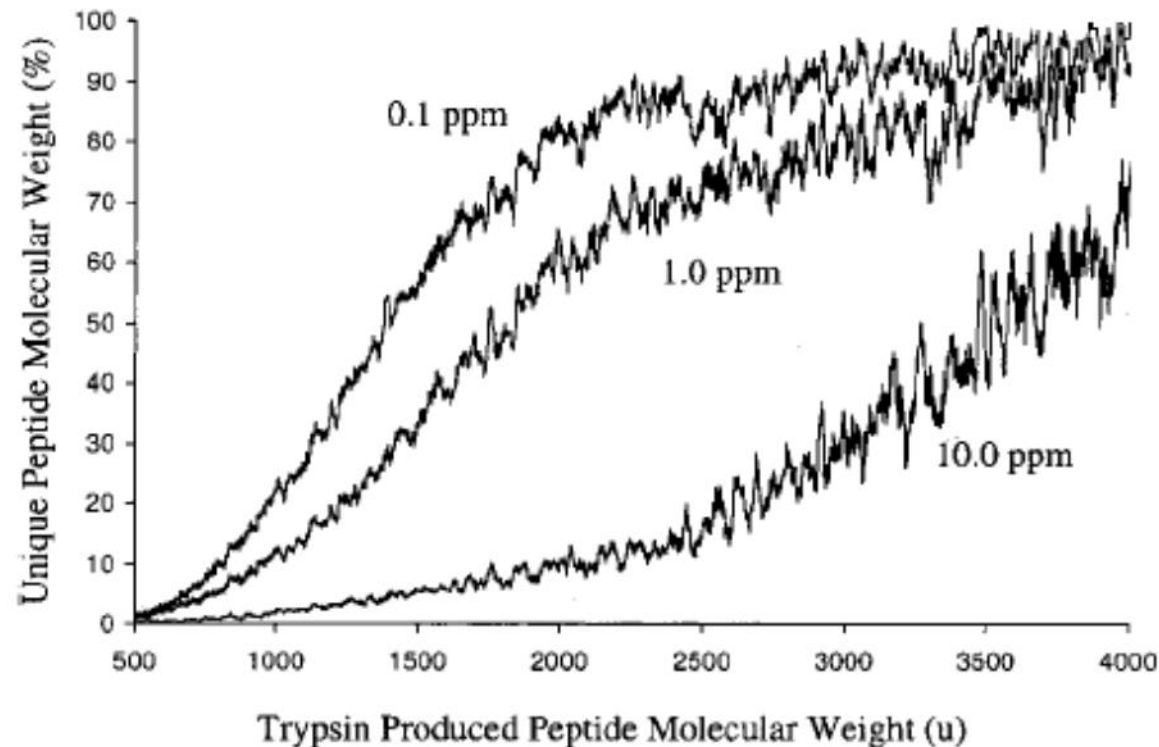



Figure 1. All possible unique peptide molecular weights after digestion of all yeast proteins in the National Center for Biotechnology Information at a mass accuracy of 0.1, 1.0, and 10.0 ppm.

A database search engine : Mascot

Home Mascot database search Products Technical support Training News Blog Contact

[Access Mascot Server](#) | [Database search help](#)

Mascot database search > Access Mascot Server > MS/MS Ions Search



MASCOT MS/MS Ions Search

Your name **Email**

Search title

Database(s) Invertebrates_EST Human_EST Fungi_EST Environmental_EST SwissProt

Enzyme Trypsin

Allow up to 1 missed cleavages

Quantitation None

Taxonomy All entries

Fixed modifications --- none selected ---

Display all modifications

Variable modifications --- none selected ---

- Acetyl (K)
- Acetyl (N-term)
- Acetyl (Protein N-term)
- Amidated (C-term)
- Amidated (Protein C-term)
- Ammonia-loss (N-term C)
- Biotin (K)
- Biotin (N-term)
- Carbamidomethyl (C)
- Carbamyl (K)
- Carbamyl (N-term)

Peptide tol. ± 1.2 Da # ¹³C 0 **MS/MS tol.** ± 0.6 Da

Peptide charge 2+ **Monoisotopic** **Average**

Data file Parcourir... Aucun fichier sélectionné.

Data format Mascot generic **Precursor** m/z

Instrument Default **Error tolerant**

Decoy **Report top** AUTO hits

MASCOT MS/MS Ions Search

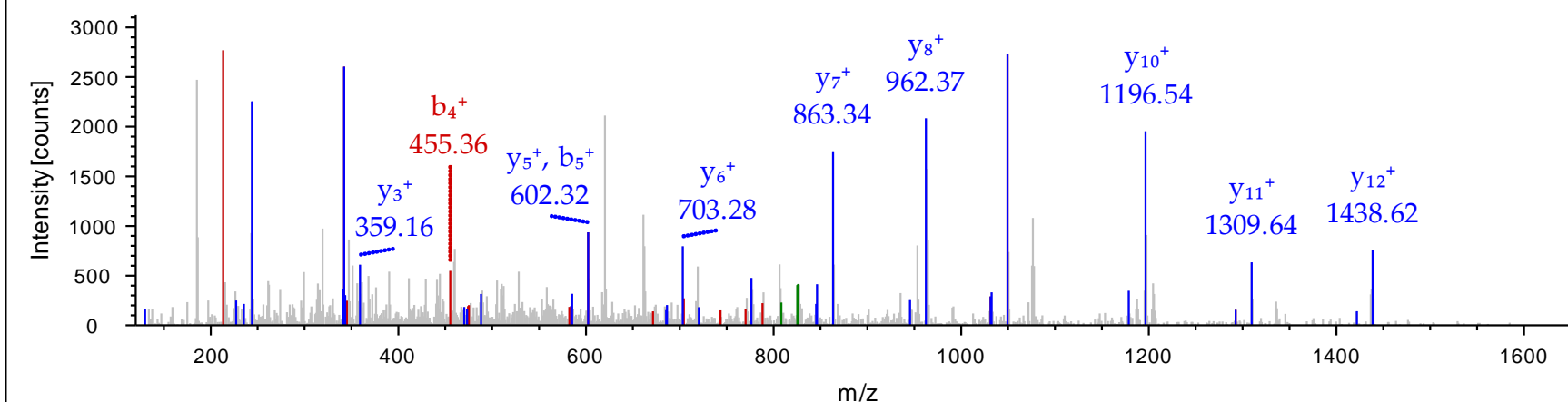
<u>Your name</u>	<input type="text"/>	<u>Email</u>	<input type="text"/>
<u>Search title</u>	<input type="text"/>		
<u>Database(s)</u>	<div style="border: 1px solid black; padding: 2px;"><ul style="list-style-type: none">Invertebrates_ESTHuman_ESTFungi_ESTEnvironmental_EST<li style="background-color: #0070C0; color: white;">SwissProt</div>	<u>Enzyme</u>	Trypsin <input type="text"/>
<u>Taxonomy</u>	<div style="border: 1px solid black; padding: 2px;"><ul style="list-style-type: none">All entries<li style="background-color: #0070C0; color: white;">All entries</div>	<u>Allow up to</u>	1 missed cleavages
<u>Fixed modifications</u>	<div style="border: 1px solid black; padding: 2px;"><ul style="list-style-type: none">.. Archaea (Archaeobacteria).. Eukaryota (eucaryotes)... Alveolata (alveolates)..... Plasmodium falciparum (malaria parasite)..... Other Alveolata... Metazoa (Animals)..... Caenorhabditis elegans..... Drosophila (fruit flies)..... Chordata (vertebrates and relatives)..... bony vertebrates..... lobe-finned fish and tetrapod clade..... Mammalia (mammals)..... Primates..... Homo sapiens (human)..... Other primates..... Rodentia (Rodents)..... Mus...... Mus musculus (house mouse)..... Rattus</div>	<u>Quantitation</u>	None <input type="text"/>
<u>Variable modifications</u>			<div style="border: 1px solid black; padding: 2px;"><ul style="list-style-type: none">Acetyl (K)Acetyl (N-term)Acetyl (Protein N-term)Amidated (C-term)Amidated (Protein C-term)Ammonia-loss (N-term C)Biotin (K)Biotin (N-term)Carbamidomethyl (C)Carbamyl (K)Carbamyl (N-term)</div>
<u>Peptide tol. ±</u>			0.6 Da <input type="text"/>
<u>Peptide charge</u>			<input checked="" type="radio"/> Average <input type="radio"/>
<u>Data file</u>			<input type="text"/> m/z
<u>Data format</u>			
<u>Instrument</u>	Default <input type="text"/>	<u>Error tolerant</u>	<input type="checkbox"/>
<u>Decoy</u>	<input type="checkbox"/>	<u>Report top</u>	AUTO hits
<input type="button" value="Start Search ..."/>		<input type="button" value="Reset Form"/>	

Search engine output formats

File name	File content
Processed peak lists	Heavily processed form of mass spectrometry data, usually derived from raw data files via various (semi-) automatic steps, e.g.: centroiding, deisotoping and charge deconvolution. These files are formatted in plain text, with typical formats like dta , pkl , ms2 or mgf .
Search engine output files	<p>These files contain the data and metadata generated by the software (called search engines) used for performing the identification and quantification of peptides and proteins. Each search engine has its own specific output file format. The outputs are typically formatted in either plain text or XML.</p> <p>mzIdentML - provides a common format for the export of identification results from any search engine.</p> <p>mzQuantML - provides a common format for the export of quantification results from any search engine.</p> <p>mzTab - represents both identification and basic quantification results.</p> <p>To allow a full representation of the processed results in the PRIDE database and in the PX tool, the search engine output files need to be converted to PRIDE XML. PRIDE Converter and PRIDE Converter 2 are the two tools developed by the PRIDE team to make this conversion possible.</p>
Protein/peptide identifications	Proteomics mass spectra can be matched to peptides or proteins, resulting in identifications for those spectra. Typically a spectrum is considered to have been identified if the score attributed to a peptide or protein match qualifies against an <i>a priori</i> or <i>a posteriori</i> defined threshold. In the case of fragmentation spectra, the initial identification will consist of a peptide sequence; subsequent steps will derive a list of proteins from the identified peptides. The protein assembly step can be a discernible process with its own input and output files, or it can be implicit in the overall identification software.

31700 MS/MS spectra interpreted!!!!

Extracted from: E:\PPSF_FUSION\2014\week45\Hela-0-5ug_141104091722.raw #48756 RT: 63.18
ITMS, HCD@35.00, z=+2, Mono m/z=825.90680 Da, MH+=1650.80632 Da, Match Tol.=0.5 Da



Sequence: VIELFSVCTNEDPK, C8-Carbamidomethyl (57.02146 Da)

Charge: +2, Monoisotopic m/z: 825.90680 Da (+0.95 mmu/+1.15 ppm), MH+: 1650.80632 Da, RT: 63.18 min,

Identified with: Sequest HT (v1.3); XCorr:4.48, Ions matched by search engine: 0/0

Fragment match tolerance used for search: 0.5 Da

Fragments used for search: b; b-H₂O; b-NH₃; y; y-H₂O; y-NH₃

Protein references (1):

- Lymphokine-activated killer T-cell-originated protein kinase OS=Homo sapiens GN=PBK PE=1 SV=3 - [TOPK_HUMAN]

5448 identified proteins

100		Q96KB5	Lymphokine-activated killer T-cell-originated protein kinase OS=Ho...	41.78	60.87 %	1	12	12	13	322	31
A2	Sequence	# PSMs	# Proteins	# Protein Groups	Protein Group Accessions	Modifications	ΔCn	q-Value	PEP	XCorr	Charge
1	IcDVGVSLPLDENMTVDPE...	1	1	1	Q96KB5	C2(Carbamidomethyl); C22...	0.0000	0	1.17e-07	5.10	3
2	VIELFSVcTNEIDPK	1	1	1	Q96KB5	C8(Carbamidomethyl)	0.0000	0	5.96e-07	4.48	2
3	SVLcSTPTINIPASPFMQK	1	1	1	Q96KB5	C4(Carbamidomethyl)	0.0000	0	3.75e-05	3.24	3
4	AFTEANDGSLcLAMEYGGBK	1	1	1	Q96KB5	C11(Carbamidomethyl)	0.0000	0	9.89e-05	3.22	2
5	INPIcNDHYR	1	1	1	Q96KB5	C5(Carbamidomethyl)	0.0000	0	0.000356	3.09	3
6	SLHHPNIVGYR	1	1	1	Q96KB5		0.0000	0	0.00076	2.86	3
7	SLNDLIEER	1	1	1	Q96KB5		0.0000	0	0.00187	2.85	2
8	ASQDPFPAAILK	1	1	1	Q96KB5		0.0000	0	0.00016	2.71	2
9	TFDESDFDDEAYYAALGTRP...	1	1	1	Q96KB5	Q32(Deamidated)	0.0000	0.001	0.0153	3.34	4
10	TFDESDFDDEAYYAALGTRP...	1	1	1	Q96KB5	N23(Deamidated); Q32(De...	0.0000	0.001	0.0198	2.40	3
11	VALNMAR	1	1	1	Q96KB5		0.0000	0.001	0.00852	2.14	2
12	EAVEENGVITDK	1	1	1	Q96KB5		0.0000	0.004	0.048	2.78	2
13	DRPSAAHIVEALETDV	1	1	1	Q96KB5		0.0000	0.006	0.0779	3.58	3

	Accession	Description	Score	Coverage	# Proteins	# UniquePeptides	# Peptides	# PSMs	# AAs	MW [kDa]
101	A0AVT1	Ubiquitin-like modifier-activating enzyme 6 OS=Homo sapiens GN=...	43.86	14.83 %	1	12	12	14	1052	111
102	O00116	Alkyl dihydroxyacetone phosphate synthase, peroxisomal OS=Homo...	40.54	25.08 %	1	11	11	11	658	77
103	Q12802	A-kinase anchor protein 13 OS=Homo sapiens GN=AKAP13 PE=1...	32.11	9.14 %	1	11	11	13	2813	307
104	O43684	Mitotic checkpoint protein BUB3 OS=Homo sapiens GN=BUB3 PE=...	38.40	44.51 %	1	11	11	11	328	33
105	O60832	H/ACA ribonucleoprotein complex subunit 4 OS=Homo sapiens GN=...	32.86	24.71 %	1	11	11	11	514	55
106	P19525	Interferon-induced, double-stranded RNA-activated protein kinase...	34.26	24.68 %	1	11	11	11	551	66
107	Q8N3D4	EH domain-binding protein 1-like protein 1 OS=Homo sapiens GN=...	43.89	11.36 %	1	11	11	13	1523	166
108	P60228	Eukaryotic translation initiation factor 3 subunit E OS=Homo sapie...	41.87	32.13 %	1	11	11	12	445	55
109	P62495	Eukaryotic peptide chain release factor subunit 1 OS=Homo sapien...	53.24	37.30 %	1	11	11	16	437	44
110	P15170	Eukaryotic peptide chain release factor GTP-binding subunit ERF3A...	45.47	36.27 %	1	11	11	12	499	55

ready 4681/4776 Protein Group(s), 5448/18724 Protein(s), 25168/152137 Peptide(s), 31700/190665 PSM(s), 88527/88527 Search Input(s)

Key questions in proteomics

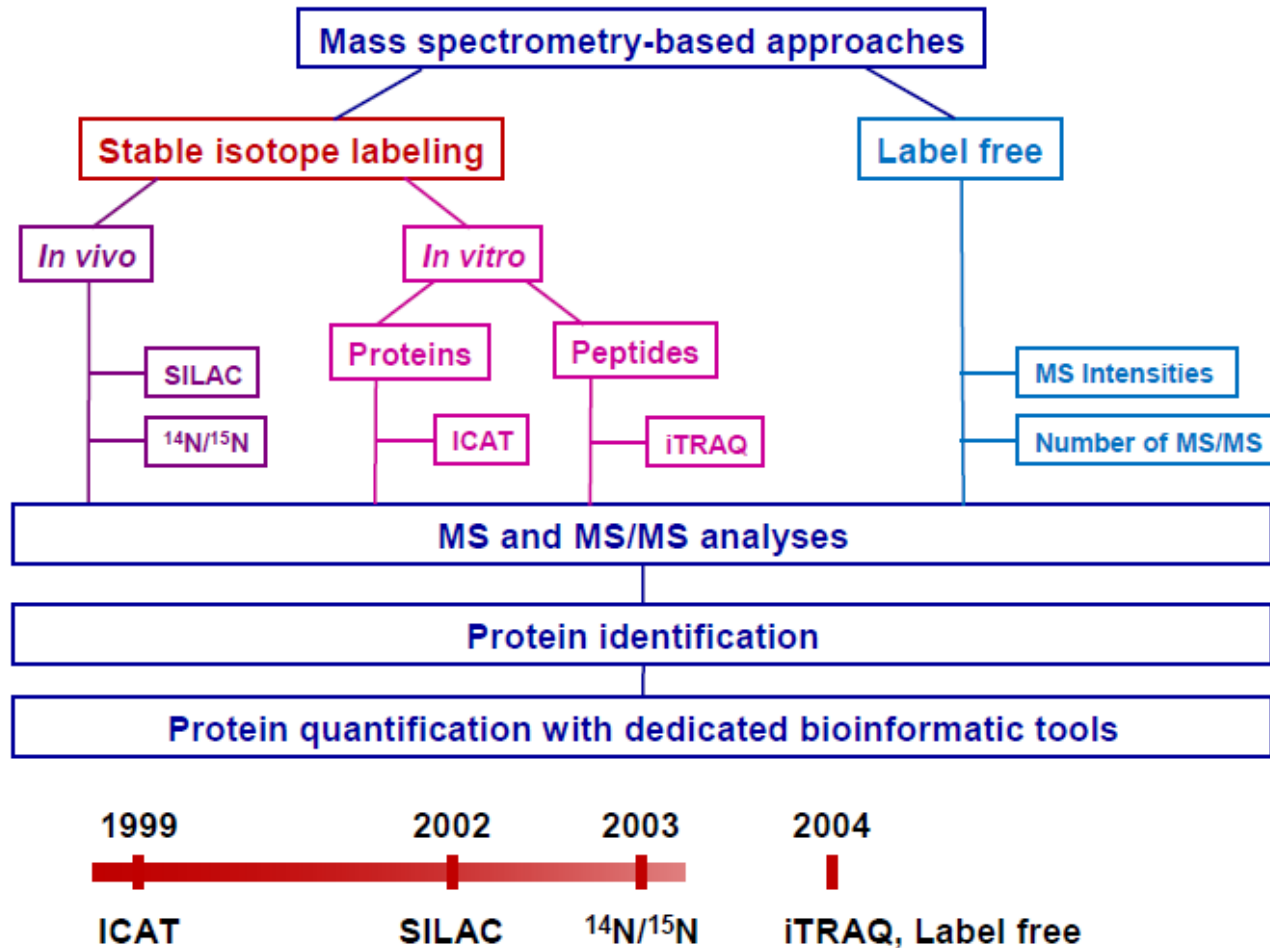
- “ What is the protein content of my biological sample?
=> problem of identification
- “ What is the abundance of my protein of interest?
=> **quantification problem**
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ What are the partners of my protein of interest?
- “ Are there any signature proteins related to a particular biological process?

=> biomarkers identifications and quantifications

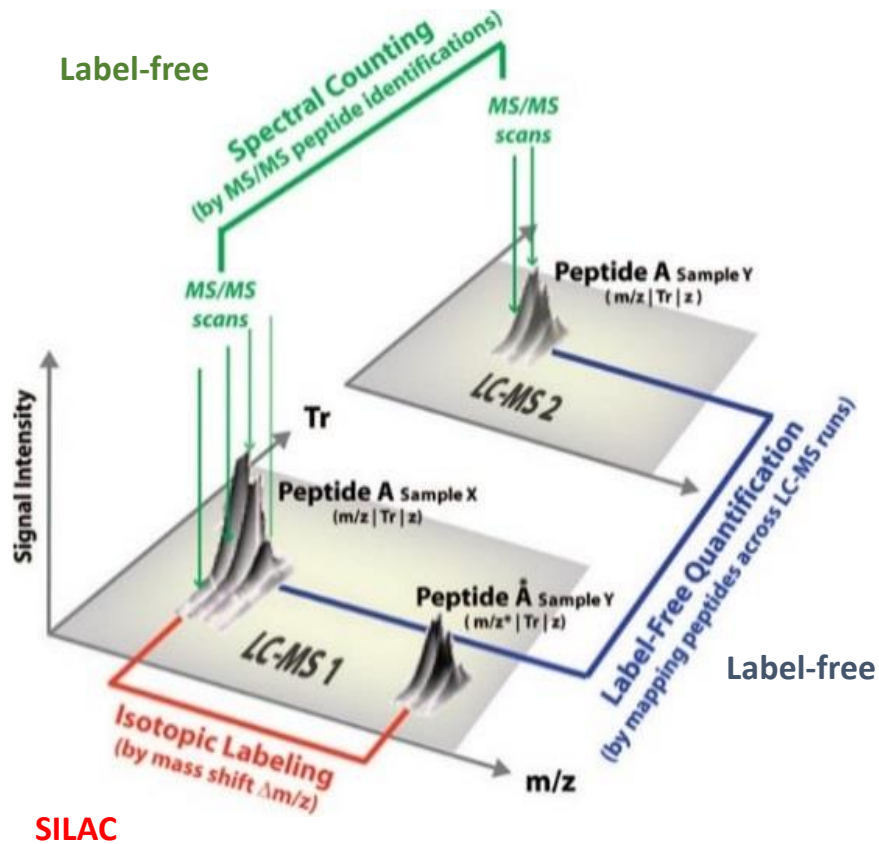
Quantitative proteomics

- “ Relative quantification
 - “ Stable isotopes labelling
 - “ Label-free
 - “ Metabolic labeling
- “ Absolute quantification

Quantitative proteomics

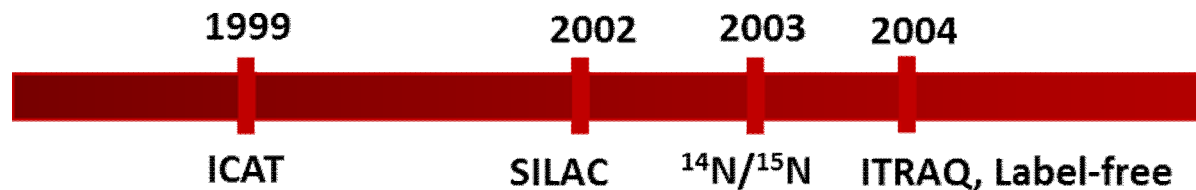


Quantitative proteomics in bottom-up

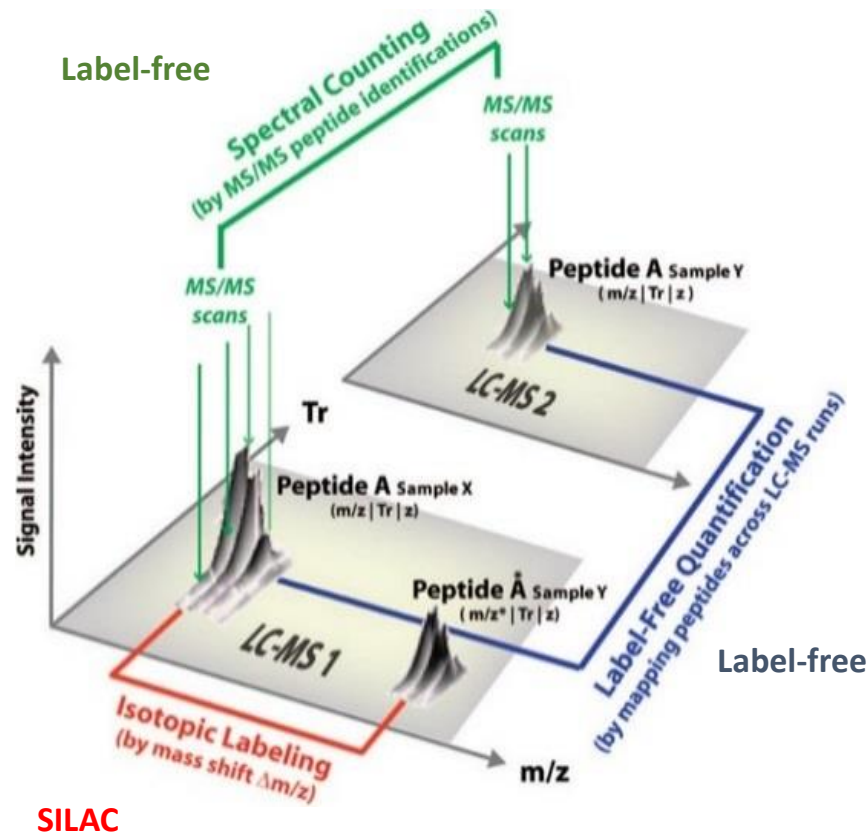


Advantages/Limitations:

- Label-free
- Metabolic Labeling (SILAC, $^{14}\text{N}/^{15}\text{N}$ – ^{13}C labeling)
- Chemical labeling (TMT, ITRAQ)



Quantitative proteomics: label-free

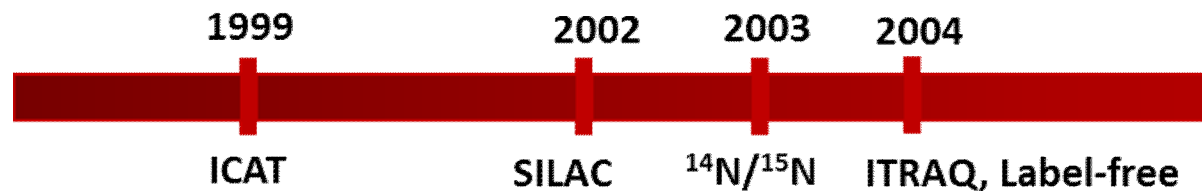


Advantages/Limitations:

Label-free:

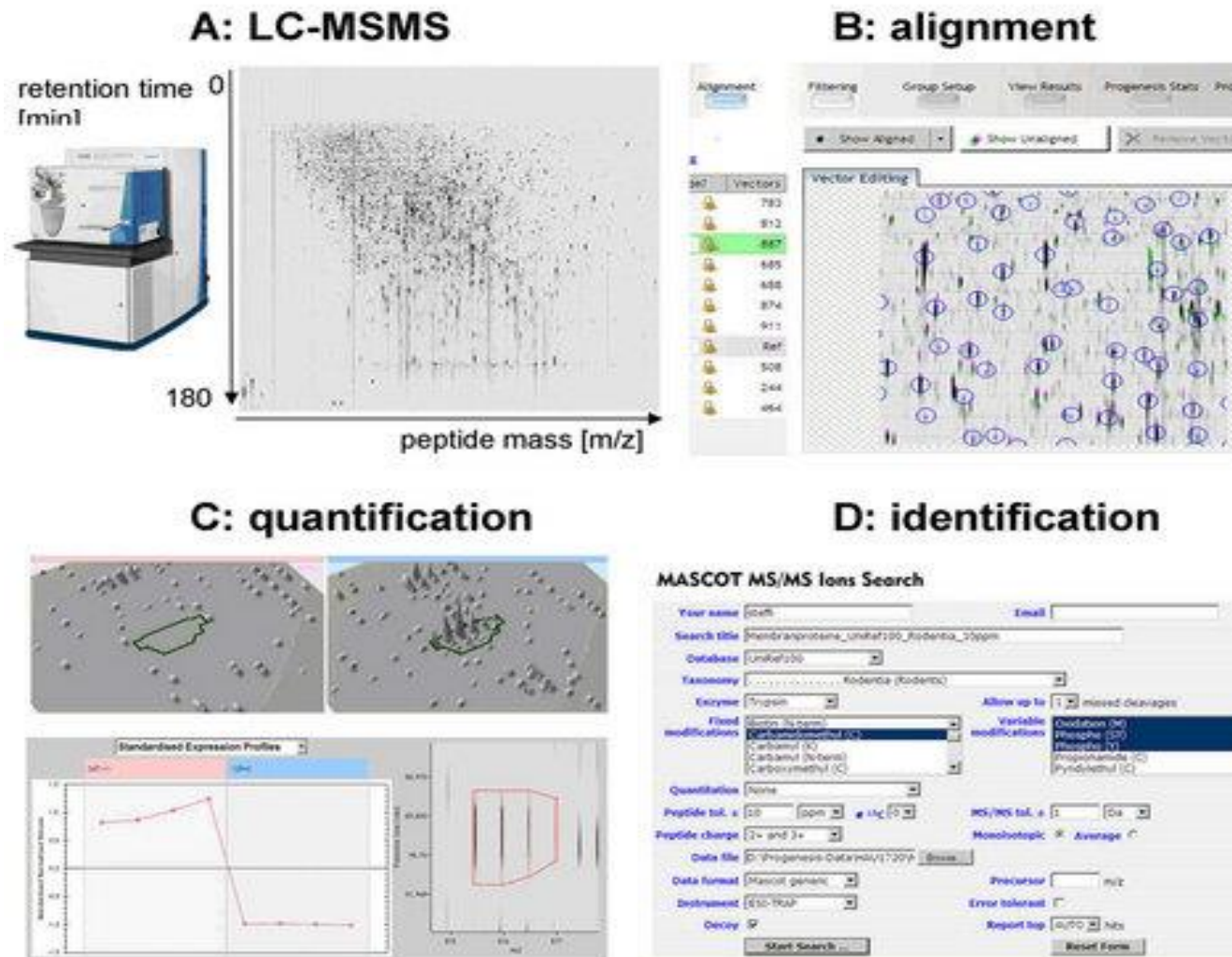
- Simplicity
- Number of identifications
- Reproducibility between runs
- Number of samples to run

Review for Label-free and yeasts:
Leger et al. Methods Mol Biol (2016)



Quantitative proteomics without labeling

Quantification label-free basée sur les intensités MS



Quantitative proteomics without labeling : results

Experiment Design

Condition	WT	1003	1006	1215	1443
Replicates	3	3	3	3	3

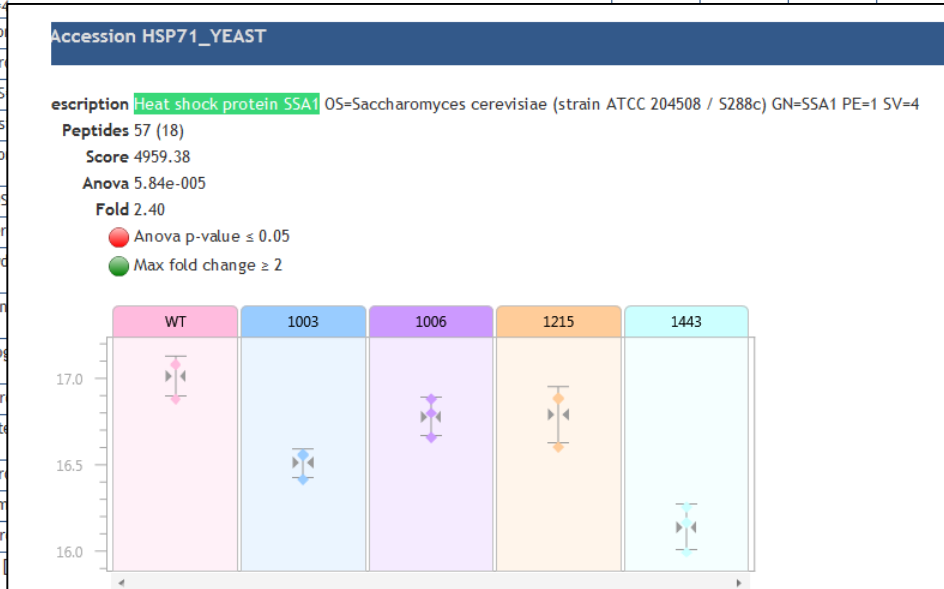
Proteins

Protein building options

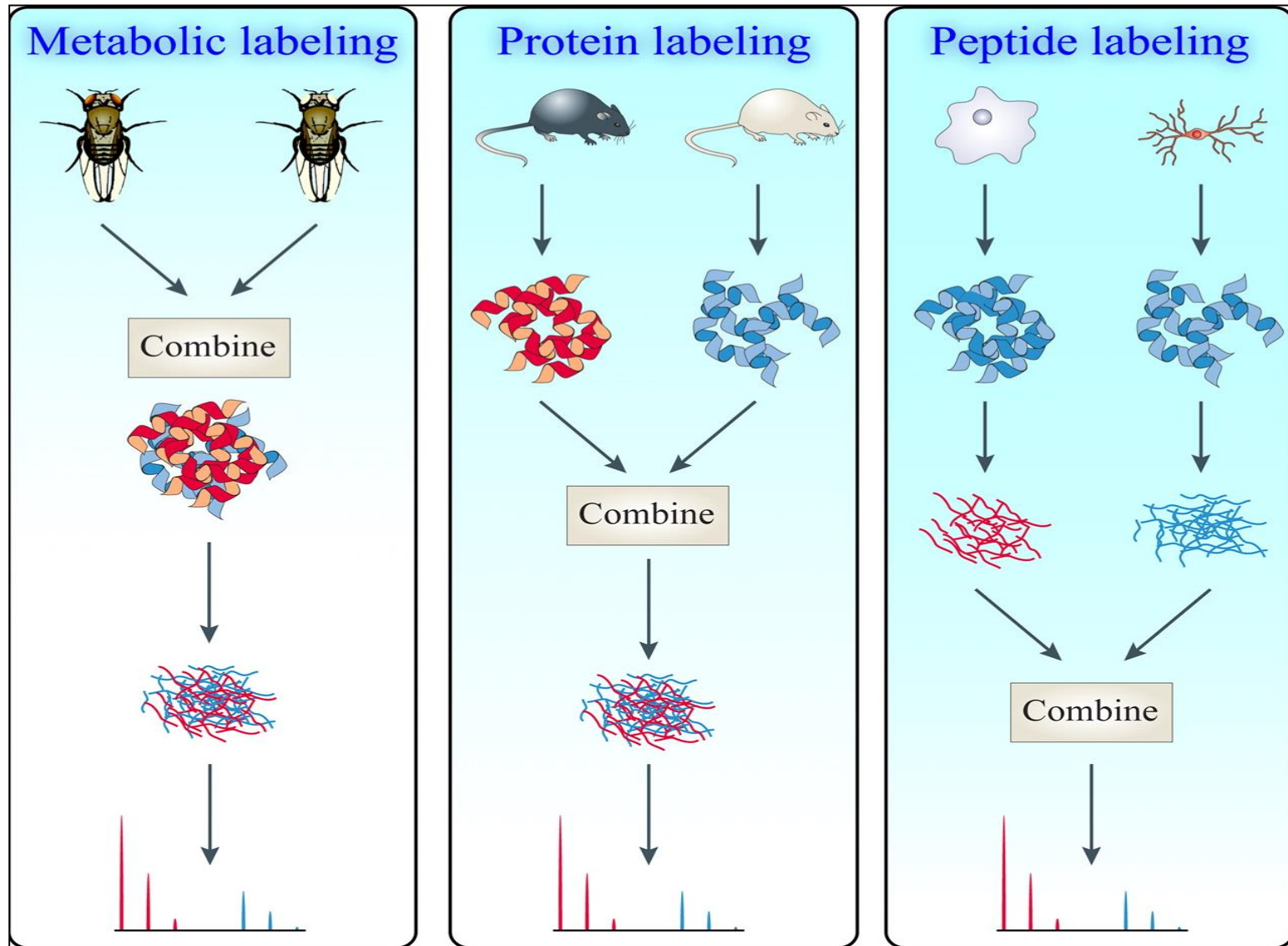
Protein grouping **Group similar proteins**

Protein quantitation **Using only features with no protein conflicts**

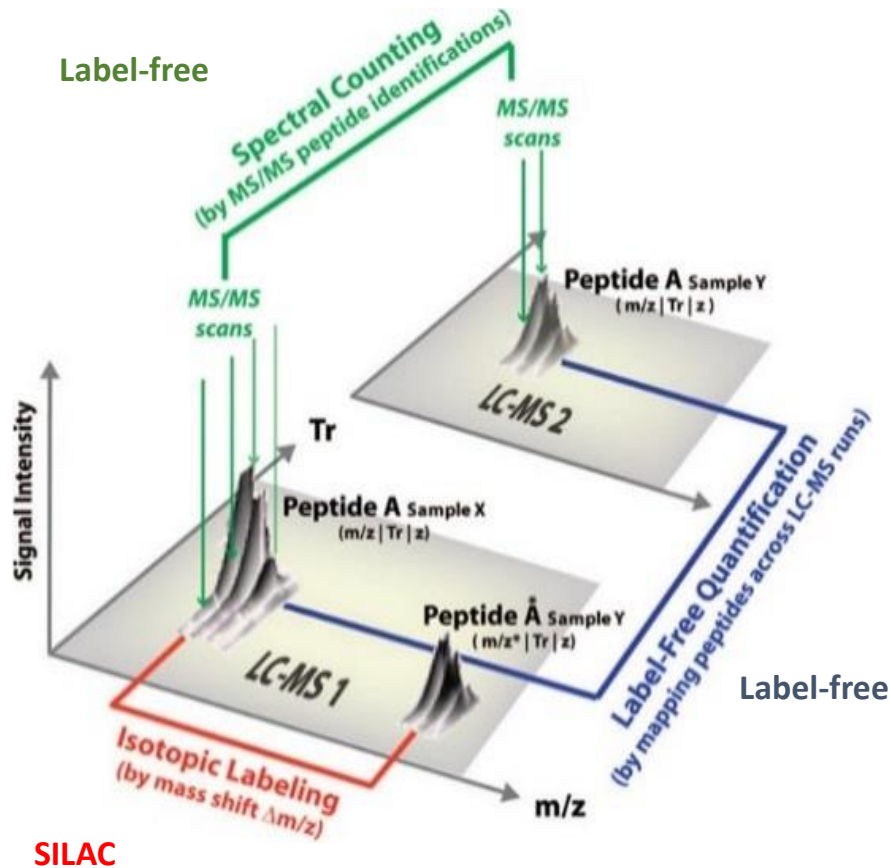
Accession	Peptides	Score	Anova (p)*	Fold	Tags	Description	Average Normalised Abundances				
							WT	1003	1006	1215	1443
HSP71_YEAST	57 (18)	4959.38	5.84e-005	2.40		Heat shock protein SSA1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=SSA1 PE=1 SV=4	1.23e+007	7.42e+006	9.72e+006	9.87e+006	5.13e+006
EF2_YEAST	69	4650.11	7.26e-004	2.16		Elongation factor 2 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=EFT1 PE=1 SV=1	3.40e+007	2.39e+007	2.89e+007	3.28e+007	1.58e+007
FAS1_YEAST	74 (71)	4506.21	9.72e-003	2.33		Fatty acid synthase subunit beta OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=FAS1 PE=1 SV=2	1.03e+007	6.55e+006	8.51e+006	1.01e+007	1.53e+007
EF3A_YEAST	58 (44)	3816.91	7.06e-006	3.32		Elongation factor 3A OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=YEF3 PE=1 SV=4	2.88e+007	1.29e+007	2.33e+007	2.72e+007	8.66e+006
METE_YEAST	46	3373.13	1.85e-006	13.40		5-methyltetrahydropteroyltriglutamate--homocysteine methyltransferase OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=MET6 PE=1 SV=4	4.34e+006	5.55e+006	3.77e+006	5.16e+006	5.05e+007
HS104_YEAST	53	3190.19	6.84e-004	2.29		Heat shock protein 104 OS=Saccharo					3.51e+006
HSP75_YEAST	40 (1)	3062.55	1.21e-006	28.16		Heat shock protein SSB1 OS=Sacchar					1.05e+004
HSP7F_YEAST	39 (32)	2658.69	2.58e-004	2.01		Heat shock protein homolog SSE1 OS					3.50e+006
ENO1_YEAST	31 (15)	2367.12	6.07e-005	2.19		Enolase 1 OS=Saccharomyces cerevis					1.46e+007
ATPA_YEAST	32	2341.09	3.17e-006	2.59		ATP synthase subunit alpha, mitochon					3.66e+006
SYLC_YEAST	37	2176.12	1.52e-006	2.01		Leucine--tRNA ligase, cytoplasmic OS					1.82e+006
HXKA_YEAST	29 (28)	2162.35	3.17e-004	2.88		Hexokinase-1 OS=Saccharomyces cer					4.61e+006
ALDH6_YEAST	30	2091.58	4.85e-004	2.15		Magnesium-activated aldehyde dehyd					2.50e+006
ATPB_YEAST	28	2015.82	4.45e-006	2.39		ATP synthase subunit beta, mitochon					4.05e+006
G3P1_YEAST	31 (21)	1986.15	8.75e-005	4.16		Glyceraldehyde-3-phosphate dehydrog					1.79e+007
HSP74_YEAST	26 (12)	1750.55	0.04	2.68		Heat shock protein SSA4 OS=Sacchar					5.24e+005
PUR92_YEAST	28 (22)	1725.94	6.21e-007	7.07		Bifunctional purine biosynthesis prot					9.63e+006
ADH1_YEAST	24 (17)	1689.13	5.88e-004	2.62		Alcohol dehydrogenase 1 OS=Sacchar					1.03e+007
HSP26_YEAST	18	1538.64	2.83e-006	2.31		Heat shock protein 26 OS=Saccharom					8.43e+006
SAHH_YEAST	27	1535.76	2.79e-006	3.51		Adenosylhomocysteinase OS=Sacchar					1.31e+007
PCKA_YEAST	20	1515.31	3.42e-009	9.67		Phosphoenolpyruvate carboxykinase					6.25e+005



Quantitative proteomics with labeling



Quantitative proteomics: metabolic labeling



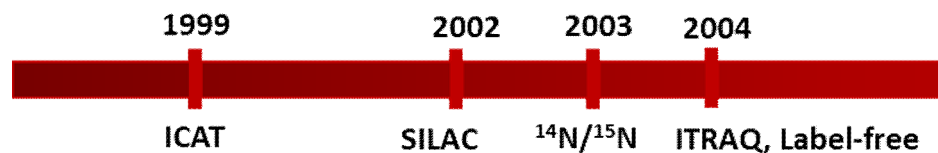
Advantages/Limitations:

□ SILAC:

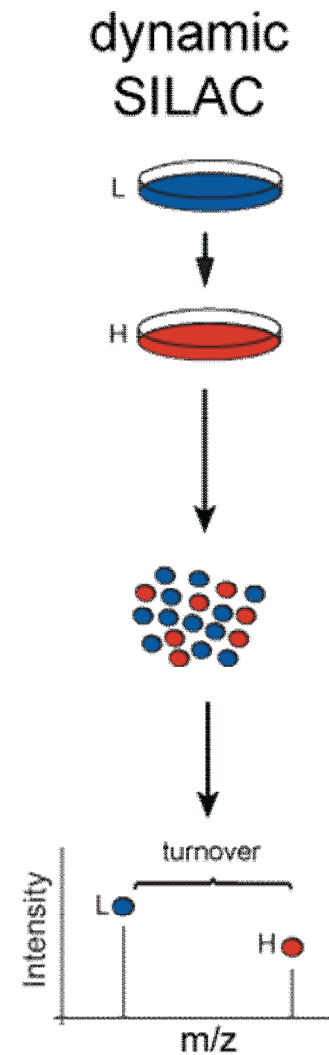
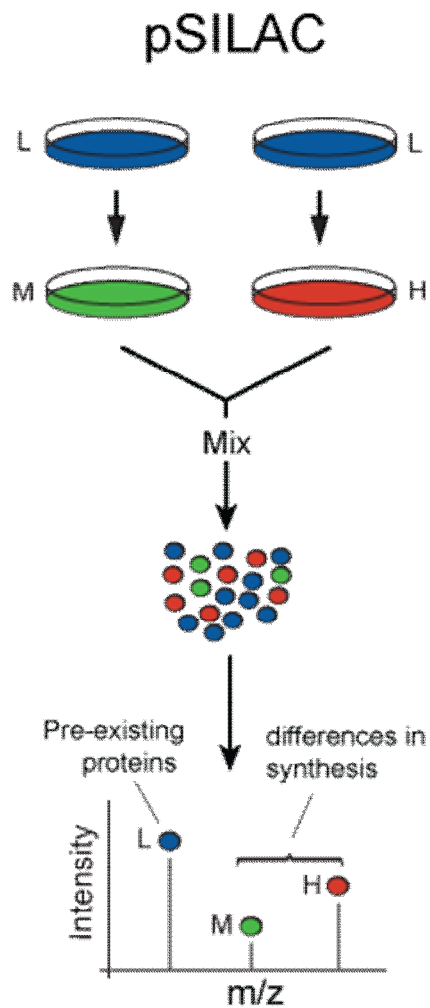
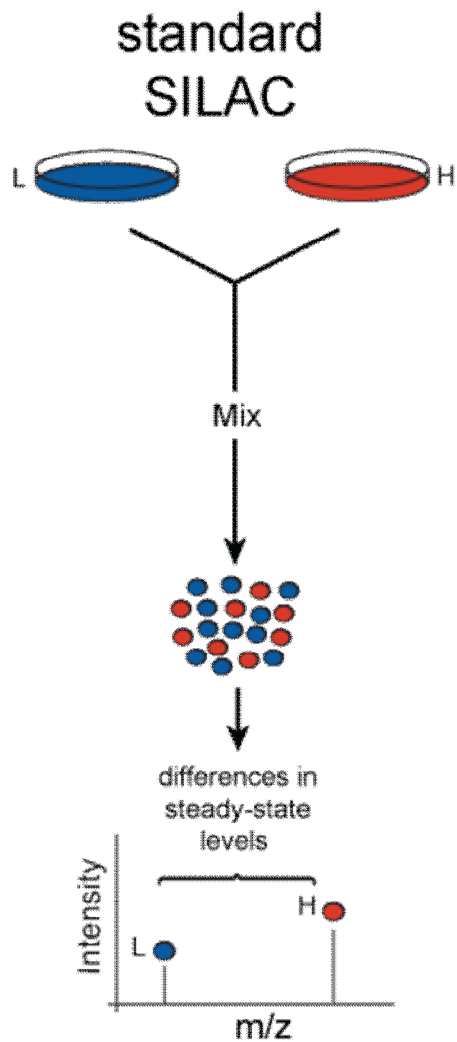
- Multiplexing
- Reproducibility
- **2 peaks instead of 1 to analyze by the MS (for 2 samples)**
- Less identifications
- Partial labeling
- Arginine/proline conversion (use of mutants)
- Trypsin exclusively

□ $^{14}\text{N}/^{15}\text{N} - ^{13}\text{C}$ labeling:

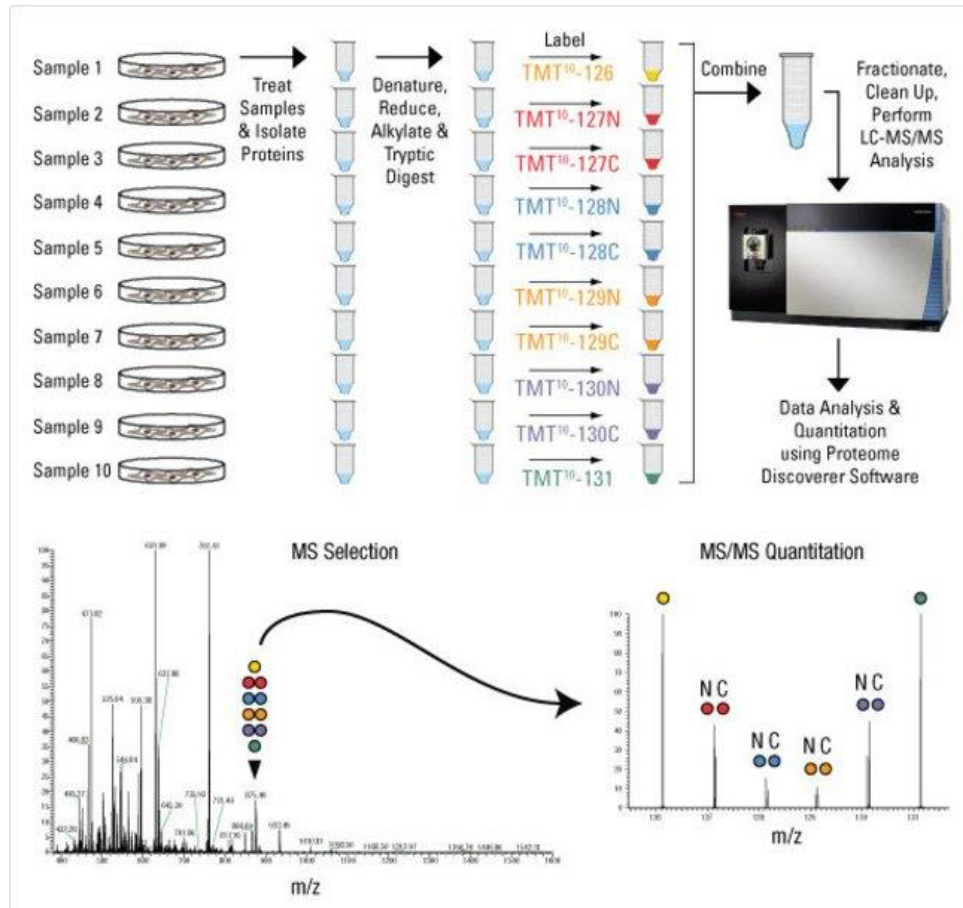
- Multiplexing
- Reproducibility
- **2 peaks instead of 1 to analyze by the MS (for 2 samples)**
- Less identifications and quantifications
- Partial labeling
- **Variable mass shift between heavy and light forms**



SILAC approaches

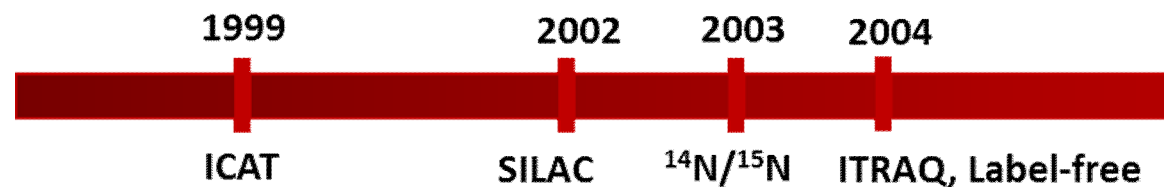


Quantitative proteomics: chemical labeling

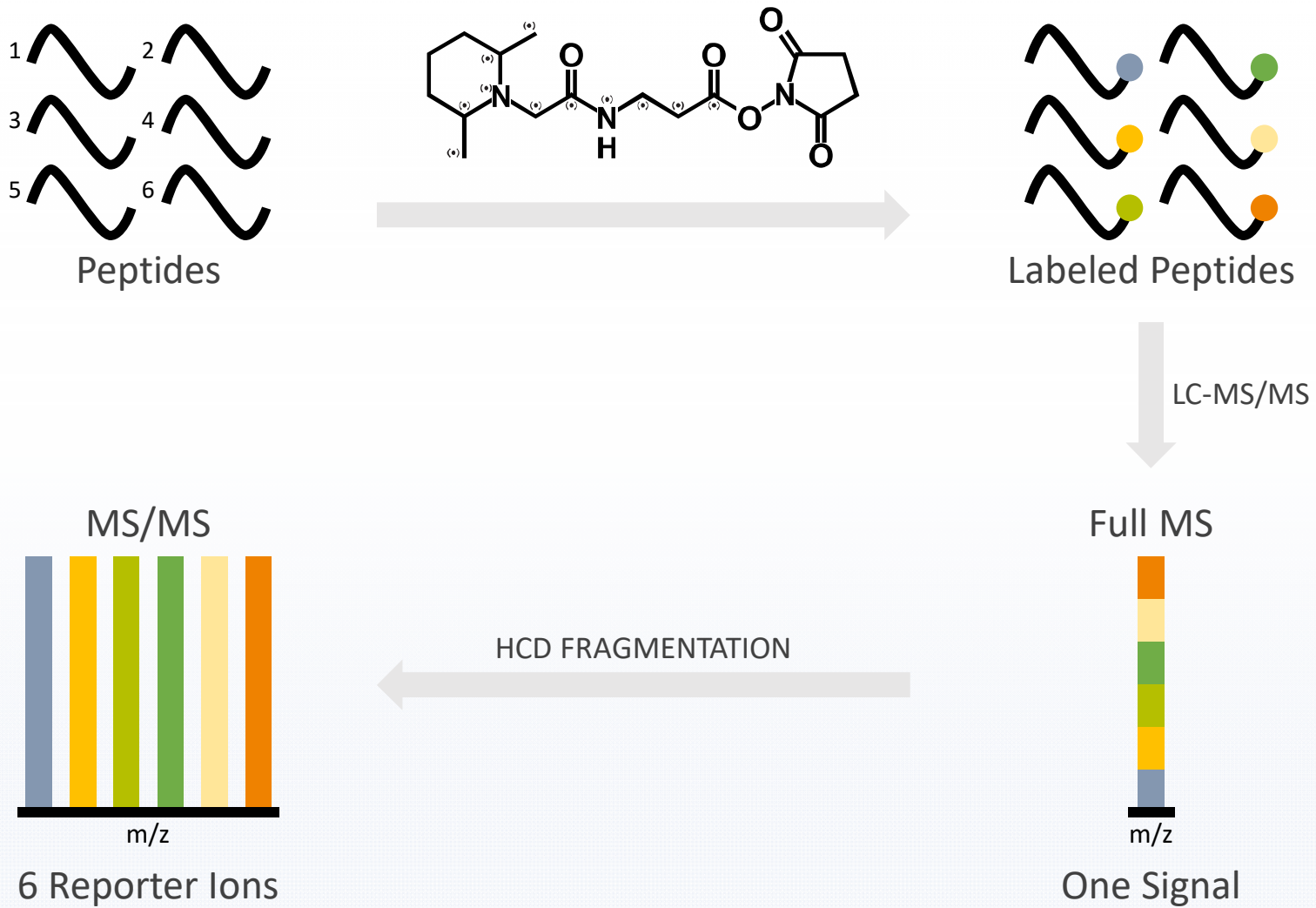


Advantages/Limitations:

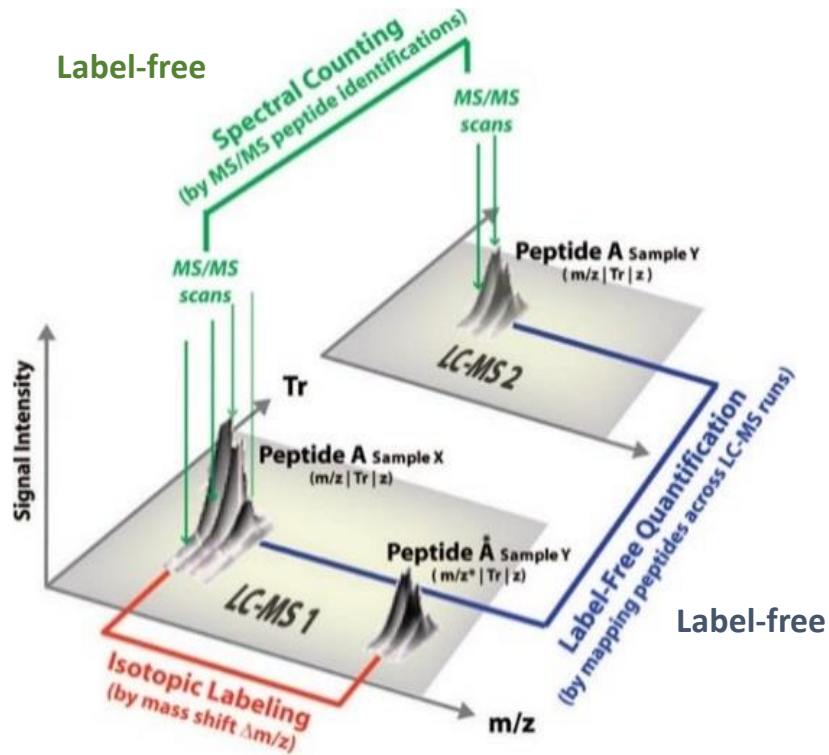
- ❑ Chemical labeling (TMT, ITRAQ)
 - Multiplexing (until 11plex)
 - Reproducibility
 - Quantification in MS2 or MS3
 - **1 peak instead of N (for N samples) to analyse in MS**
 - Amount of materials for the peptide labeling
 - Need of resolution in MS2 for quantifications
 - Incomplete labeling
 - Less identifications and quantifications



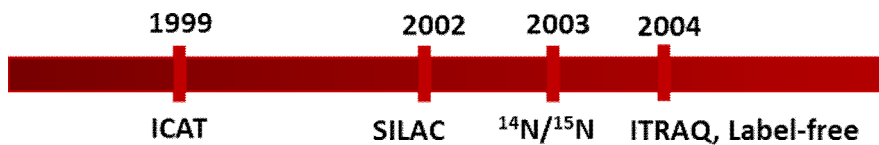
TMT labeling : principles



Quantitative proteomics in bottom-up



SILAC



Advantages/Limitations:

Label-free:

- Simplicity
- Number of identifications
- Reproducibility between runs
- Number of samples to run

SILAC:

- Multiplexing
- Reproducibility
- **2 peaks instead of 1 to analyze by the MS (2 samples)**
- Less identifications
- Partial labeling
- Arginine/proline conversion (use of mutants)
- Trypsin exclusively

$^{14}\text{N}/^{15}\text{N} - ^{13}\text{C}$ labeling:

- Multiplexing
- Reproducibility
- **2 peaks instead of 1 to analyze by the MS (2 samples)**
- Less identifications and quantifications
- Partial labeling
- **Variable mass shift between heavy and light forms**

Metabolic labeling

Chemical labeling (TMT, ITRAQ)

- Multiplexing
- Reproducibility
- **1 peak instead of N to analyse in MS (N samples)**
- Amount of materials for the peptide labeling
- Need of resolution in MS2 for quantifications
- Incomplete labeling
- Less identifications and quantifications

Quantitative proteomics without labeling : export

Peptide level!



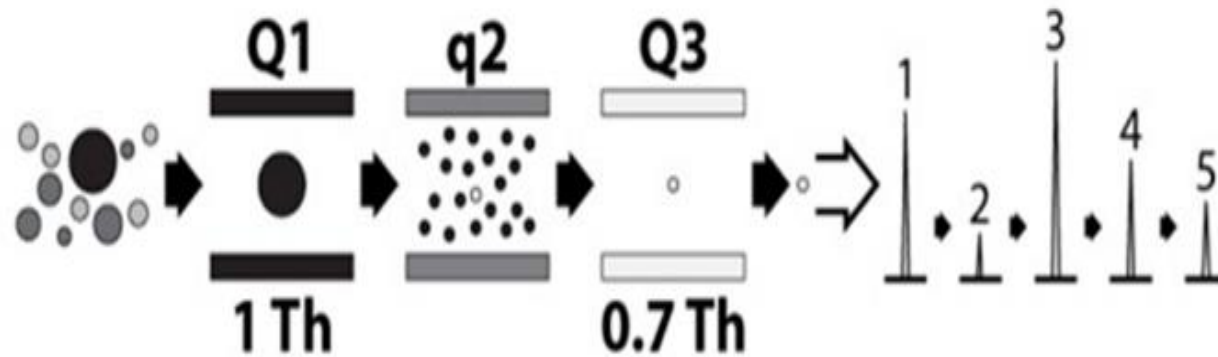
ID	Peptide	Ions used	Deconvolu	Deconvolu	Retention	Neutral ma	Score	Best identification	Sequence	Modification	Accession	Description	Max fold of	Highest m	Lowest m	Normalized abundance		Raw abundance		Spectral counts									
																P2Y2-IFN	NT-P2Y2	NT-P2Y2	P2Y2-IFN	NT-P2Y2	NT-P2Y2								
1	72.73_130	2	2	#115;#1147 2,3	72.73305	1301.7079	75.54	SLLDLSIAEK		P04264	Keratin,	1.0524126	P2Y2-IFN	NT-P2Y2	0.78116031	106.87214	5429311.51	23369307.2	6511636.4	117442590.3	117444671.5	2102461.81	3616651.21	23369307.2	3625390.6	4227100.8	2766726.5	4354021.4	1
2	60.30_130	0	1	#6540	60.296133	1301.7094	20.29	SLLDLSIAEK		P04264	Keratin,	1.14065223	P2Y2-IFN	NT-P2Y2	0.8070285	50.874645	32369.596	24424.675	44472.255	28914.657	6718.7124	27474.576	21561.3117	24424.675	24389.321	59032.895	14397.925	56837.534	0
3	60.51_130	0	1	#26858	60.54783	1301.7088	16.04	SLLDLSIAEK		P04264	Keratin,	1.2179871	P2Y2-IFN	NT-P2Y2	0.3895835	123.23264	0	233.60288	915.21675	695.04198	253.57423	537.22822	0	239.60288	501.91967	51.00625	53.735206	111.55443	0
4	39.00_138	1	1	#150	39.001683	1178.5339	71	YELQITAGR		P04264	Keratin,	1.2378820	P2Y2-IFN	NT-P2Y2	0.3403633	32.299321	2168831.75	1268884.3	671002.91	1752814.0	3032025.1	1843956.7	1444307.2	1268884.3	915964.34	3948417.1	734025.95	3818673.1	1
5	56.96_138	0	1	#153	56.96445	1382.6833	87.18	SLNNGFASFDIK		P04264	Keratin,	0.20711715	P2Y2-IFN	NT-P2Y2	0.0601139	39.471061	5158842.5	1173669.8	1667027.21	1919750.7	3133234.7	1956241.5	1038341.2	1173669.8	9835808.7	31429354.6	712653.66	4051211.4	1
6	55.63_147	0	1	#17867	55.687866	1474.7409	66.14	WELLQVDTSTR		P04264	Keratin,	1.9501543	NT-P2Y2	P2Y2-IFN	0.0332066	52.594856	1494.515	11166.4226	18652.09	4235.6873	11984.023	6503.1219	9654.7621	11166.4226	10223.1142	3299.7283	2263.1166	31467.4168	0
7	56.63_147	0	1	#200	56.633916	1474.7414	86.01	WELLQVDTSTR		P04264	Keratin,	1.5717223	NT-P2Y2	NT-P2Y2	0.0755519	27.940603	1417558.4	1012105.52	1125228.17	1469256.3	2452542.6	1665466.5	944232.26	1012105.52	617093.72	3229396.1	1577753.2	10449040.9	1
8	63.32_127	0	1	#215	63.3216	1276.7040	64.55	LALDLEIATR		P04264	Keratin,	1.5188866	P2Y2-IFN	NT-P2Y2	0.9260536	107.53163	864082.21	724826.8	1671152.21	521744.34	3127963.1	536437.4E	575563.0E	742826.8E	640085.5E	1145523.7	736864.0C	110916.80	0
9	63.32_127	0	1	#34344	63.315488	1276.7034	27	LALDLEIATR		P04264	Keratin,	Infinity	NT-P2Y2	P2Y2-IFN	0.3739005	173.20508	14.238816	0	0	0	0	0	0	0	0	0	0	0	0
10	26.18_103	0	1	#292	26.184133	1032.5095	34.35	TLLGEESR		P04264	Keratin,	1.9482378	P2Y2-IFN	NT-P2Y2	0.037473E	36.507447	565077.21	253522.0E	435785.6E	746444.7E	855097.4C	854063.0E	376396.57	253522.0E	238992.0E	1638868.2	201437.95	1768862.6	0
11	32.36_873	0	1	#300	32.362833	873.49232	27.04	SLVNLGGSK		P04264	Keratin,	1.3133291C	NT-P2Y2	P2Y2-IFN	0.489357E	94.323936	973040.0C	428278.9C	1248380.4	310007.98	1405023.6	302512.44	648193.61	426278.9C	684632.4E	680642.77	7330985.7E	628477.8C	0
12	43.15_156	0	1	#323	43.146133	1356.6889	57.58	LNLDELALQAK		P04264	Keratin,	51.472069	P2Y2-IFN	NT-P2Y2	0.0309571	80.583775	16964.421	13808.1491	11363.3921	1045477.9	50893.905	1033393.3	11299.9604	13808.1491	62318724	3821261.61	11989.2364	2139987.8	0
13	70.37_193	2	2	#6877;#35 2,3	70.369506	1932.9637	70.42	THILPEYFSPINLR		P04264	Keratin,	1.2724553	P2Y2-IFN	NT-P2Y2	0.7562446	60.67555	1656628.7	1358222.1	2568785.5	1666749.1	4225490.5	1241989.3	1104808.6	1358222.1	1408784.6	3659456.5	995411.94	2570525.2	2
14	40.10_126	0	1	#418	40.09796	1264.6928	48.42	ITNAEFTIK		P04264	Keratin,	1.263207	P2Y2-IFN	NT-P2Y2	0.601753E	33.807571	200263.61	25635.08	7372.152	365349.31	688108.31	133394.70	15235.86	107093.05	161858.5E	248666.467	1420669.3	1	
15	58.13_163	0	2	#3340;#68 2,3	58.132484	1631.8523	35.56	SLNNGFASFDIKR		P04264	Keratin,	2.597494E	P2Y2-IFN	P2Y2-IFN	0.835334	189.06068	1727366.1	5368070.05	2632931.7	12001.384	1835856.5	14564.298	1150594.4	536070.05	1443989.0	219550.0E	443666.33	20161.4368	1
16	57.42_163	0	1	#23689	57.47463	1637.8526	7.32	SLNNGFASFDIKR		P04264	Keratin,	4.4531173	P2Y2-IFN	NT-P2Y2	0.0425537	128.06406	55.862671	51.810944	1046.3398	1631.5291	6695.6456	503.47831	51.810944	573.85934	371.49734	327.80688	13686.1162	0	
17	23.27_112	0	1	#661	23.274633	1124.5353	23.8	AEAESLYGSK		P04264	Keratin,	1.3469793	P2Y2-IFN	NT-P2Y2	0.3103307	58.585204	307010.56	88304.354	170421.451	265433.31	233707.32	262893.9E	204501.6E	88304.354	93461.941	52776.79	55055.16	5144443.67	0
18	23.02_119	0	2	#2369;#75 2,3	23.020275	1192.7249	33.58	ITNAEFTIKK		P04264	Keratin,	2.1442453	NT-P2Y2	P2Y2-IFN	0.2880216	138.35432	70194.03	284797.07	913521.64	129647.01	793707.22	419.82833	526349.67	284797.07	500990.9E	284648.6E	187824.14	869.42911	0
19	33.21_114C	0	1	#745	33.209283	1140.5125	37.44	DYQELMNTK		P04264	Keratin,	11.7282151	P2Y2-IFN	NT-P2Y2	0.0468781	80.410623	22287.805	8061.3943	4603.8936	170095.05	25034.00E	214878.75	14845.856	8061.3943	2528.1419	373454.8C	5897.3393	444995.8C	0
20	42.91_171E	0	2	#806;#181 2,3	42.91125	1715.8444	97.44	QISNLQQSDAQR		P04264	Keratin,	0.9315362	P2Y2-IFN	NT-P2Y2	0.0145680	54.749022	80525.022	48208.510	29468.530	632154.40	175286.52	621765.60	53637.524	48208.510	15612.507	1387936.2	41252.791	1287624.2	0
21	61.26_121	0	1	#24914	61.2566	1215.8456	53.58	QISNLQQSDAQR		P04264	Keratin,	18.12896	P2Y2-IFN	NT-P2Y2	0.0951607	87.852537	906.37633	572.13906	116134721	19403.903	1078.6449	29007.632	603.73513	572.13906	636.91683	333920.42E	254.09375	60072.48E	0
22	26.92_101	0	1	#4913	26.923366	1014.4330	37.9	DQDGYAY (7)		P04264	Keratin,	1.9520103	NT-P2Y2	NT-P2Y2	0.704596C	42.33310	214773.11	119855.00C	21134.766	15135.333	311778.96	165837.86	143059.85	119855.00C	115888.50C	33222.7C	73446.74C	343436.2E	0
23	44.18_129	0	1	#963	44.177	1299.5227	37.02	NMQDMVEYR		P04264	Keratin,	5.409689E	P2Y2-IFN	NT-P2Y2	0.1051817	72.759371	41620.224	39418.937	4937.4123	107284.34	46089.08E	246894.21	27723.131	39418.937	25212397	373870.4C	10857.349	511297.18	0
24	17.30_133	0	2	#2112;#123 2,3	17.30085	1339.6613	31.13	SKMAEASLYGSK		P04264	Keratin,	1.9486523	NT-P2Y2	P2Y2-IFN	0.3391951	111.941052	513258.24	156505.35	458228.19	83212.1173	575888.62	34574.90C	341880.08	165805.05	251296.81	182697.61	136663.87	195866.67	1
25	44.18_131E	0	1	#1434	44.189488	1315.5173	32.37	NMQDMVE (2)	Oxidat	P04264	Keratin,	2.457723	P2Y2-IFN	NT-P2Y2	0.1043870	61.658569	105909.68	106807.21	22637.96	185121.76	167031.23	232488.3E	7121.617K	106807.21	12447.94	396542.9E	39346.067	481463.80	0
26	30.45_131E	0	1	#18448	30.451381	1315.5176	12.88	NMQDMVE (2)	Oxidat	P04264	Keratin,	1.2507193	P2Y2-IFN	NT-P2Y2	0.8843262	145.31614	694.4236	482.26351	0	4420.9633	538.28895	0	455.89235	482.26351	0	9706.519	126.6059	0	
27	32.97_131E	0	1	#10520	32.96875	1315.5178	31.19	NMQDMVE (5)	Oxidat	P04264	Keratin,	0.1031217	P2Y2-IFN	NT-P2Y2	0.1031217	73.515817	4309.4306	3925.9455	240.77486	1707.8368	2870.5014	3925.9455	132.04491	38000.44E	1708.3234	36652.33C	0		
28	25.24_115E	0	1	#6851	25.25231	1156.5075	21.94	DYQELMNTK (6)	Oxidat	P04264	Keratin,	3.8834292	P2Y2-IFN	NT-P2Y2	0.3200131	106.0885	6169.6506	3879.5554	1660.5098	1124.8844	5422.753C	17567.372	446.05563	3879.5554	101.65103	44675.572	127448.3638	544E	0
29	33.21_115E	0	1	#1463	33.209283	1156.5076	36.97	DYQELMNTK (6)	Oxidat	P04264	Keratin,	2.7419825	P2Y2-IFN	NT-P2Y2	0.005596	40.549866	26723.29	19080.851	11637.3917	185290.107	145420.507	145420.507	17804.247	19080.851	6382.908	406816.6E	17355.857	319934.11	0
30	43.53_153	0	2	#3020;#21 2,3	43.586816	1538.8268	54.06	NKLNDELALQAK		P04264	Keratin,	3.4440257	P2Y2-IFN	NT-P2Y2	0.0214198	56.125894	71152.864	46552.266	22588.207	139778.40	187801.031	185255.27	51391.337	46552.266	12371.293	306892.62	42097.157	383648.0E	0
31	43.53_153	0	1	#2737	43.532145	1538.7880	88.97	SGLLDELALQAK		P04264	Keratin,	2.1066378	NT-P2Y2	P2Y2-IFN	0.487622E	76.268465	110914.26E	126677.55	259589.5	60367.44E	237850.7E	82016.387	73879.72E	126677.55	142363.2	133333.47	56031.24E	1168943.02	1
32	30.44_133	0	1	#11109	30.4389	1331.5123	40	NMQDMVE (2)																					

Quantification output formats

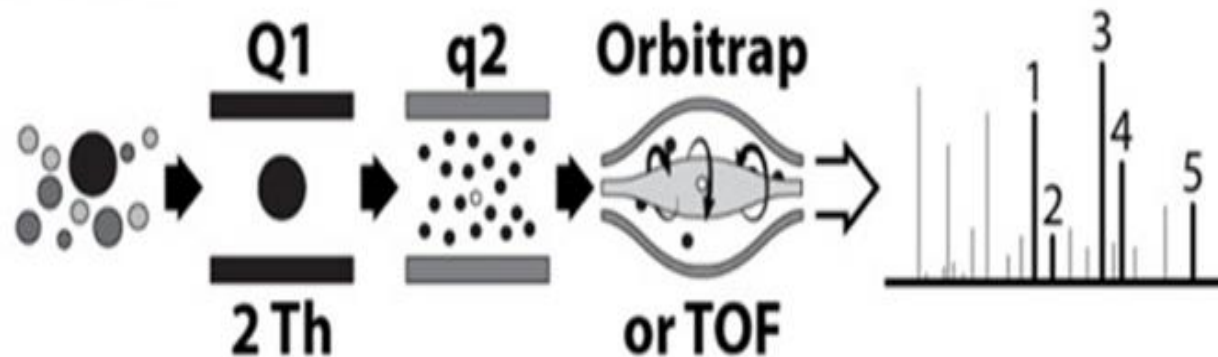
File name	File content
Protein/peptide quantification	Protein/peptide expression values can also be obtained from an MS--based proteomics experiment and then this data and metadata is used for performing the quantification analysis of peptides and proteins.
Metadata	A term used to describe data that provides additional information about a particular data set. This information can include how, when and where the data set was generated and what standards were used. In the proteomics context the addition of metadata such as peptide and protein identifications and quantification of their expression values gives meaning to a simple collection of mass spectra output files.

Targeted proteomics : PRM mode

SRM



PRM



Absolute quantification

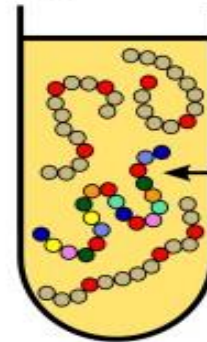
PSAQ standard
(isotope-labeled protein)



● ARG or LYS

● Stable isotope-labeled ARG or LYS

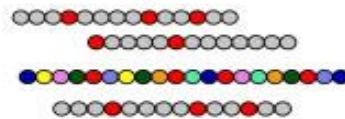
PSAQ



Target protein in
a body fluid

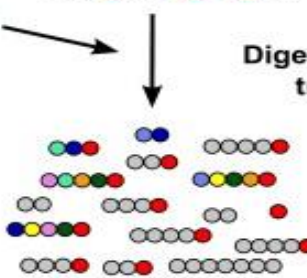
Sample
prefractionation

QconCAT standard
(isotope-labeled concatamer of
proteotypic peptides)



Digestion with
trypsin

AQUA standard
(isotope-labeled proteotypic peptide)



QconCAT

Peptides AQUA

Liquid chromatography and
mass spectrometry analysis

Filtered data?

Peptide filters

- Unique peptides
- FDR <0.01 (or scoring)
- p-value <0.05 (t-tests, Anova)
- Scoring for PTMs localization (e.g ptmRS)

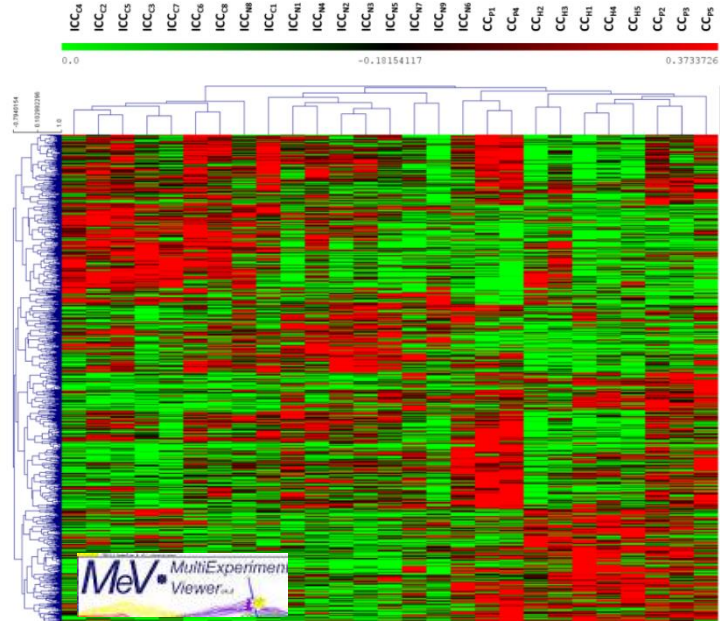
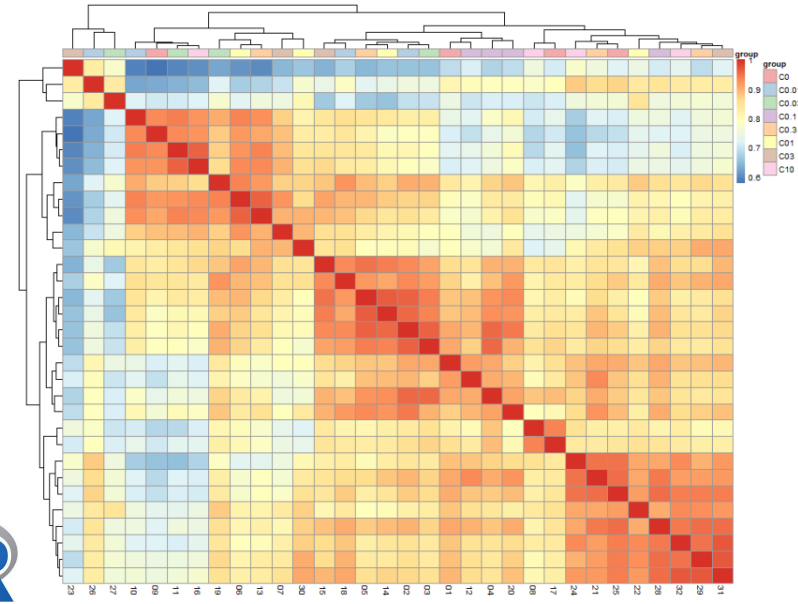
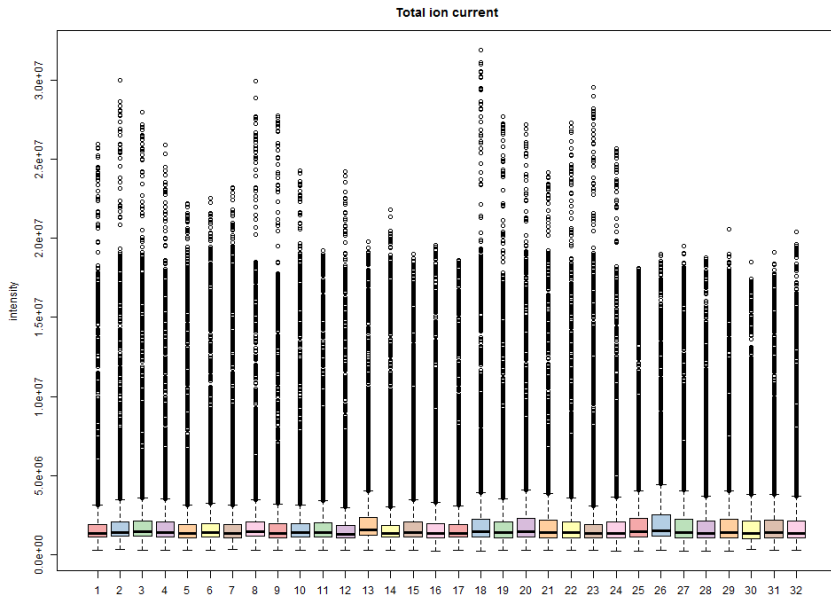
Protein filters

- 2 peptides by protein
- Unique accession
- FDR <0.01 (or scoring)
- p-value <0.05 (t-tests, Anova)

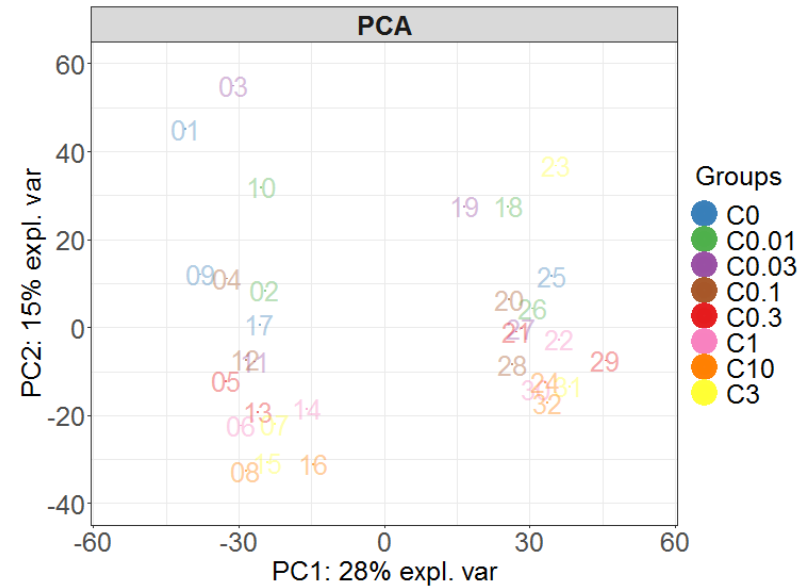
	Peptides						
Case 1	1	2	3	4	5	Parsimony Reported?	Grouping Reported?
Protein A	X	X	X	X	X	Yes	Yes
Protein B	X	X				No	No
Protein C			X	X	X	No	No
Case 2	1	2	3	4	5	Parsimony Reported?	Grouping Reported?
Protein D	X	X				Yes	Yes
Protein E		X	X			No	Yes
Protein F			X	X		Yes	Yes

No score ratio to compare a protein abundance between 2 conditions!
No score ratio to compare two different proteins in 1 sample!

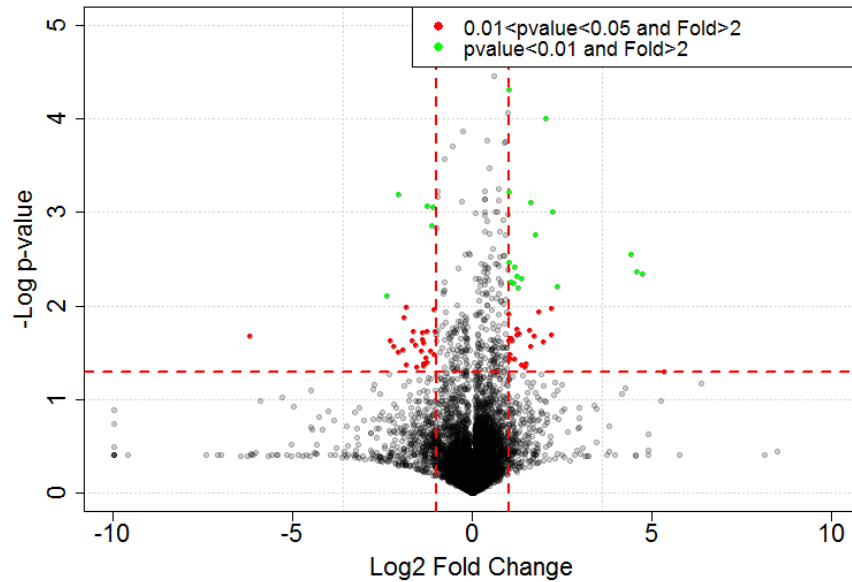
Unsupervised analyses



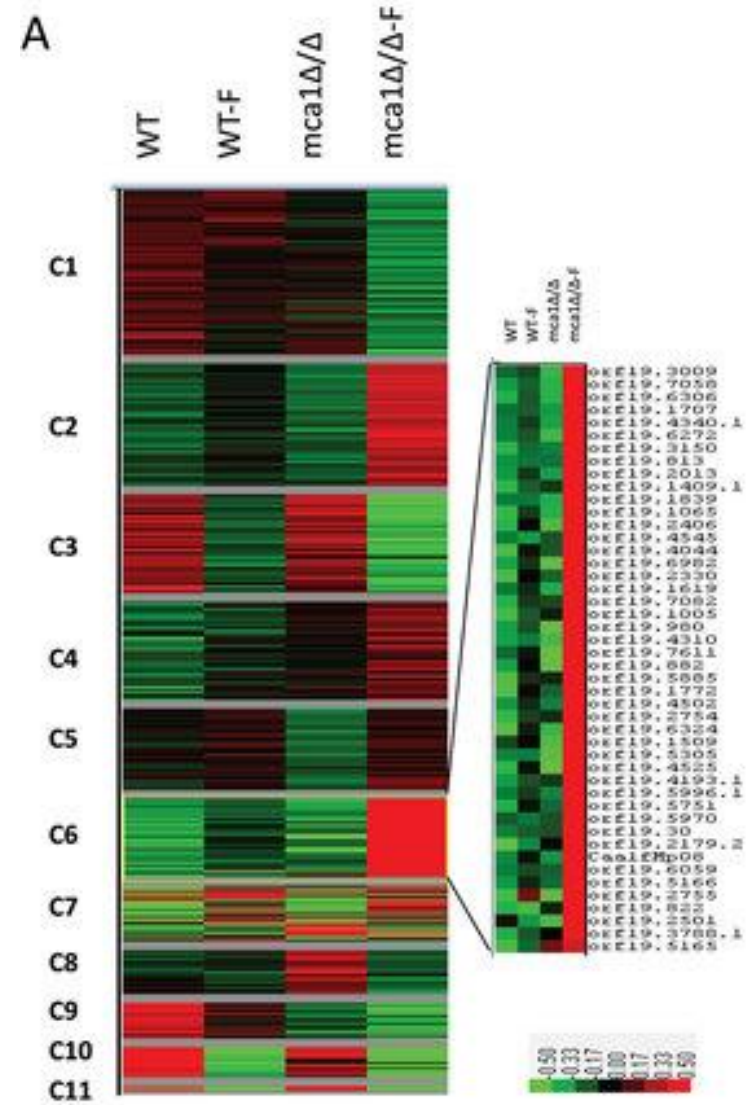
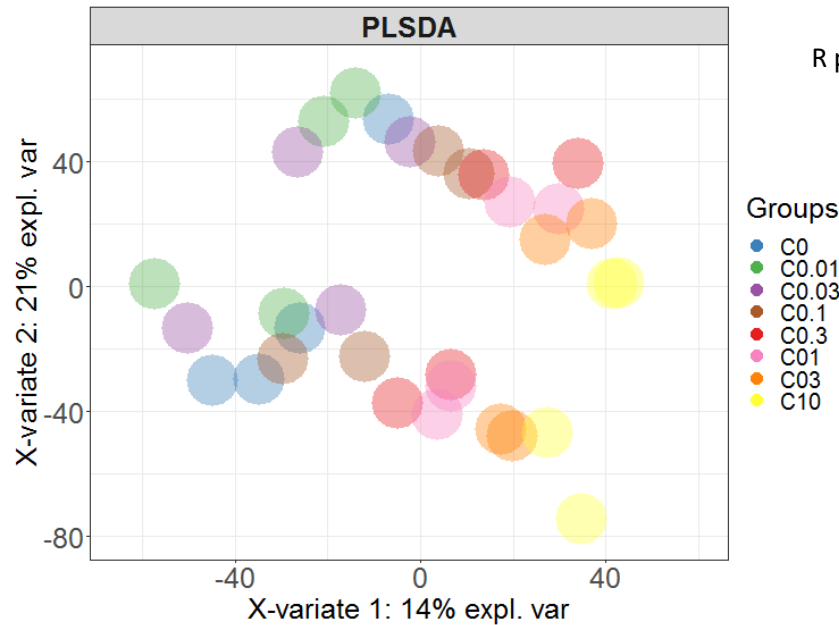
R packages:
pcaMethods
mixOmics
factoextra



Supervised analyses



R packages:
ropls

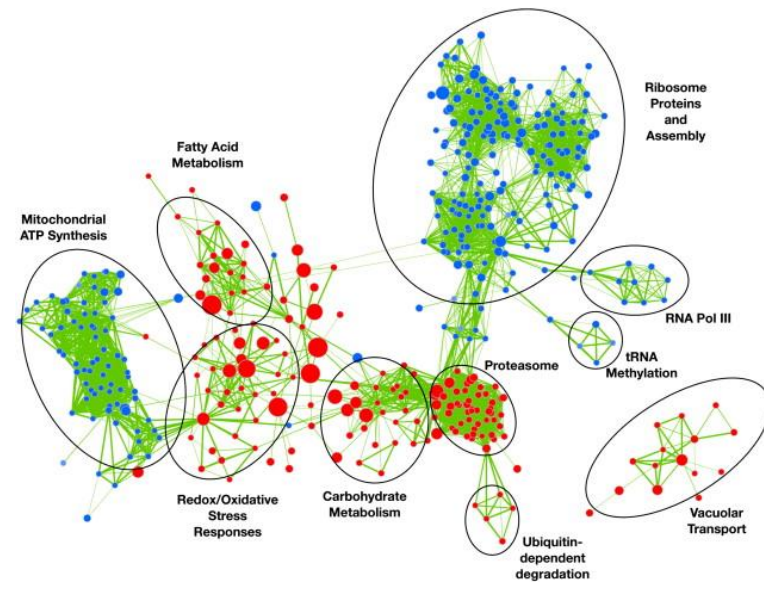
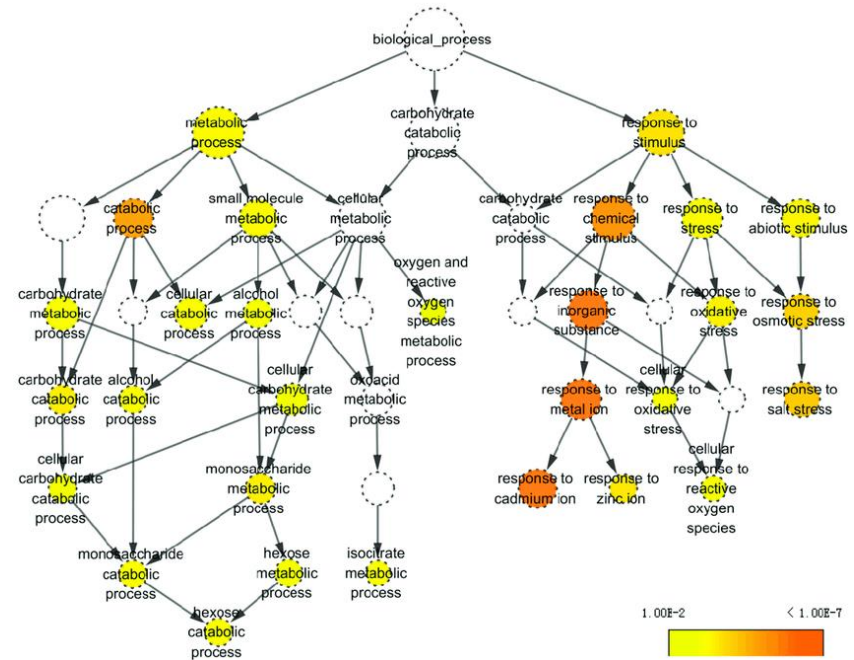


Leger et al. Mol Cell Proteomics 2015

Gene Ontology analyses

- Cytoscape plugins: BINGO, CLUEGO
- PANTHER (Protein ANalysis THrough Evolutionary Relationships) Classification System
- GSEA (Gene Set Enrichment Analyses)
- David software
- Tools dedicated to specific species:
 - “ Candida Genome database (CGD)
 - “ Saccharomyces Genome Database (SGD)

Fisher's exact test / hypergeometric test

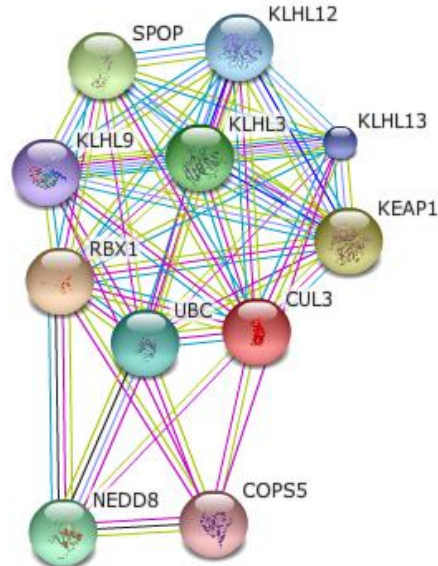
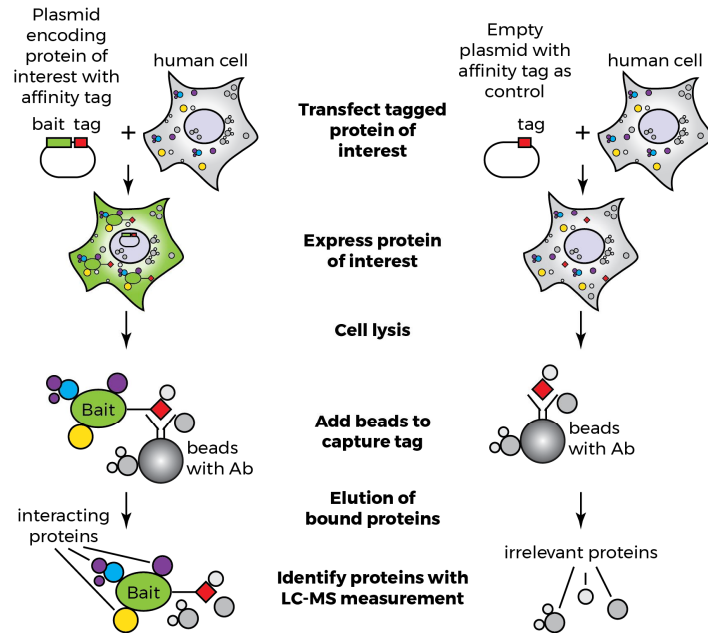


Key questions in proteomics

- “ What is the protein content of my biological sample?
=> problem of identification
- “ What is the abundance of my protein of interest?
=> quantification problem
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ **What are the partners of my protein of interest?**
- “ Are there any signature proteins related to a particular biological process?

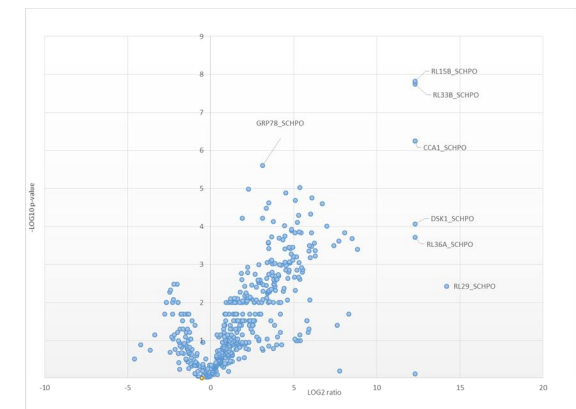
=> biomarkers identifications and quantifications

Affinity purification MS (AP-MS)



Description	Score A3	Score B3	Score C3
Cullin-3 OS=Homo sapiens GN=CUL3 PE=1 SV=2-[CUL3...		1172.08	547.08
Cullin-associated NEDD8-dissociated protein 1 OS=Homo...	0.00	394.42	0.00
E3 ubiquitin-protein ligase RBX1 OS=Homo sapiens GN=R...		251.21	123.68
Tubulin beta chain OS=Homo sapiens GN=TUBB PE=1 SV...		199.36	136.63
Kelch-like protein 20 OS=Homo sapiens GN=KLHL20 PE=...		164.62	78.37
Kelch-like protein 13 OS=Homo sapiens GN=KLHL13 PE=...		158.68	86.54
Tubulin beta-4B chain OS=Homo sapiens GN=TUBB4B PE...		150.00	143.23
Kelch-like protein 42 OS=Homo sapiens GN=KLHL42 PE=...		149.87	62.55
ADP/ATP translocase 2 OS=Homo sapiens GN=SLC25A5 P...		148.12	57.72
COP9 signalosome complex subunit 5 OS=Homo sapiens...		142.70	29.32
COP9 signalosome complex subunit 8 OS=Homo sapiens...		135.68	37.43
Tubulin alpha-1B chain OS=Homo sapiens GN=TUBA1B P...	37.89	135.00	103.98
Kelch-like protein 9 OS=Homo sapiens GN=KLHL9 PE=1 S...		131.61	110.90
ADP/ATP translocase 3 OS=Homo sapiens GN=SLC25A6 P...		131.31	78.50
Kelch-like protein 15 OS=Homo sapiens GN=KLHL15 PE=...		130.71	35.01
Kelch-like protein 12 OS=Homo sapiens GN=KLHL12 PE=...		127.87	23.09
Elongation factor Tu, mitochondrial OS=Homo sapiens GN...		112.52	119.93
Kelch-like protein 8 OS=Homo sapiens GN=KLHL8 PE=2 S...		110.71	28.11
COP9 signalosome complex subunit 6 OS=Homo sapiens...		106.43	25.42

Ilektra Kouranti (HEGP)

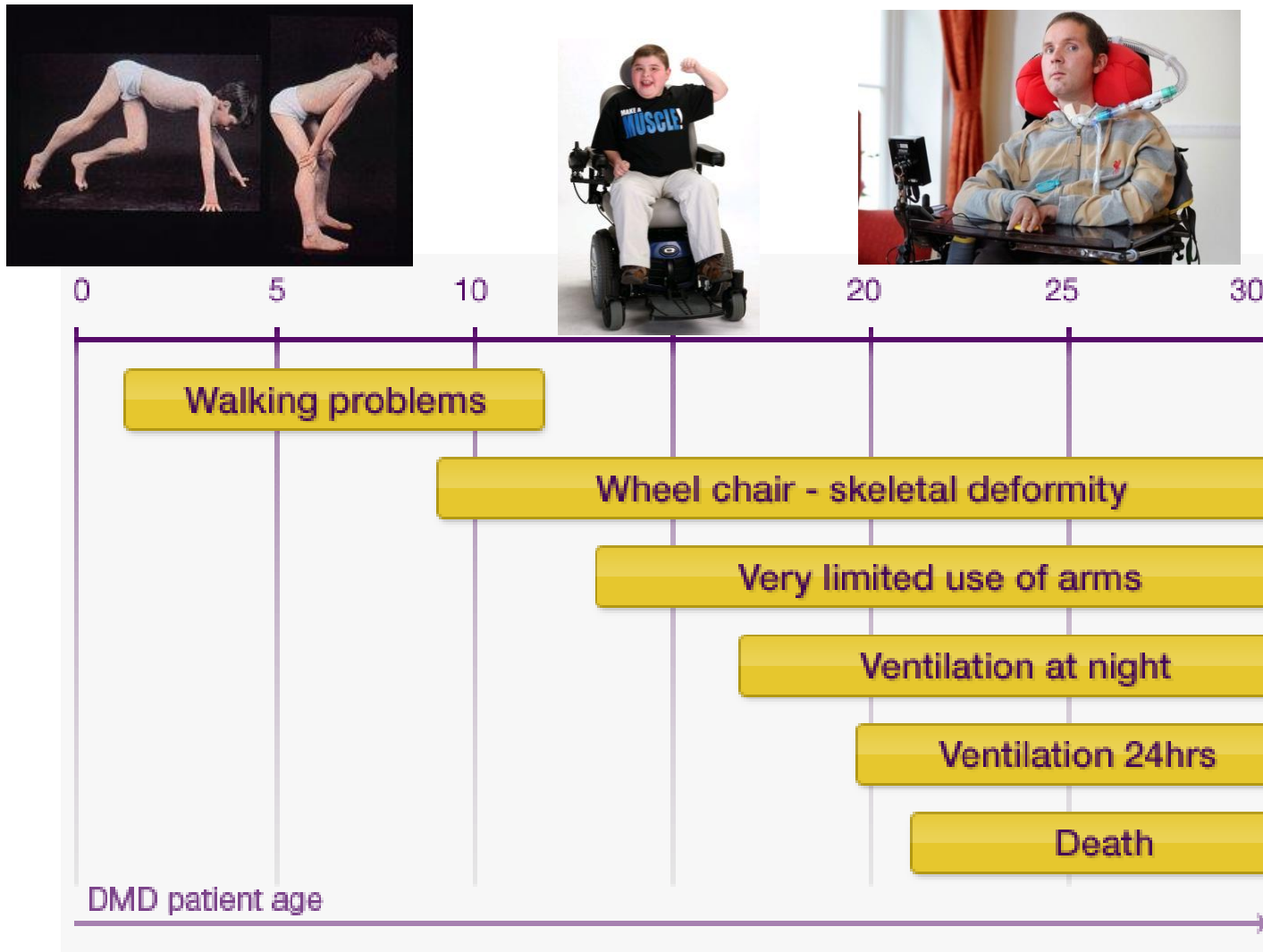


Key questions in proteomics

- “ What is the protein content of my biological sample?
=> problem of identification
- “ What is the abundance of my protein of interest?
=> quantification problem
- “ Relative question: What are the protein abundance variations of the proteomes studied?
- “ What are the partners of my protein of interest?
- “ Are there any signature proteins related to a particular biological process?

=> **biomarkers identifications and quantifications**

Biomarkers: applications to Duchenne dystrophy



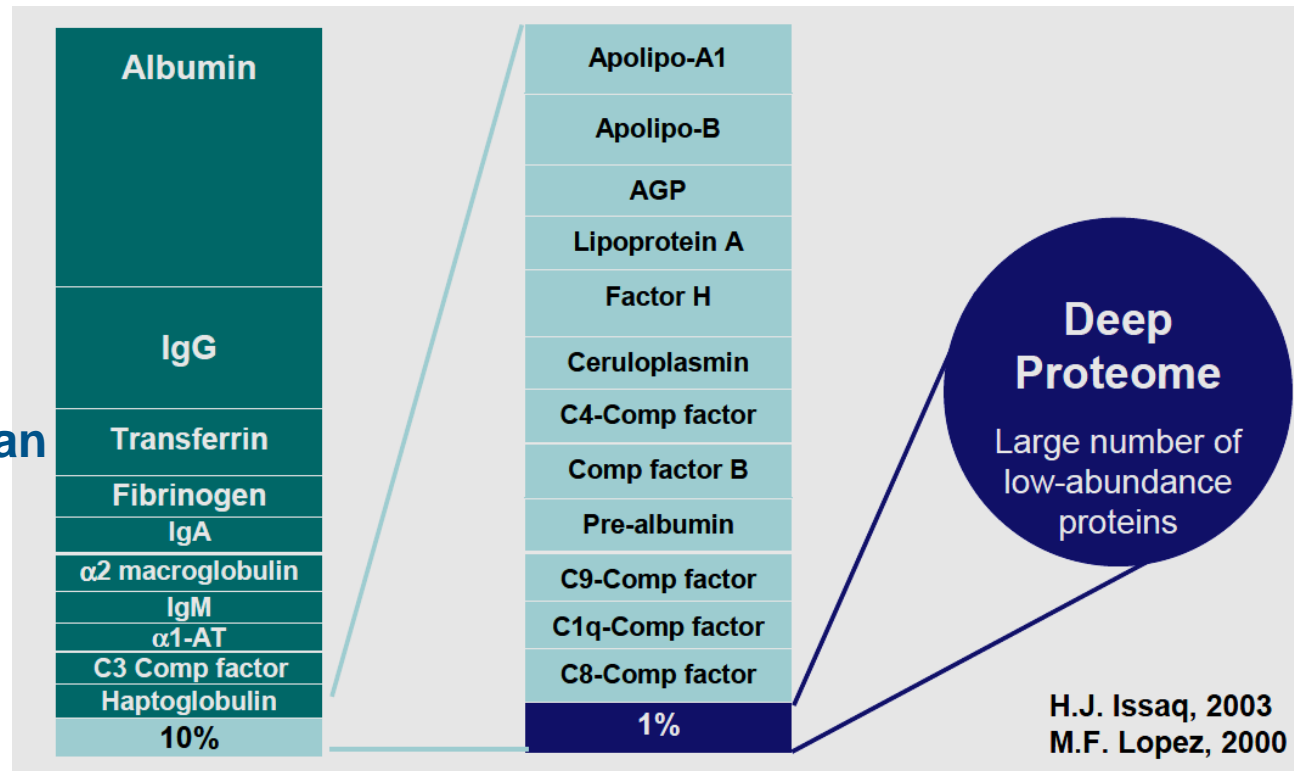
~ Death of DMD patients usually occurs ~ 30

Serum: a tricky fluid for Mass Spectrometry

Serum : Mixture of proteins with different ranges of proteins concentration
(from mg/ml to pg/ml)

99% of serum
proteome = 20 major
proteins

1% remaining = more than
one thousand proteins



- Albumin : ~40 mg/ml (60% of serum proteome)
- C-reactive protein: ~1 µg/ml (40 000 times less than albumin)
- FGF-9 : ~400 pg/ml (100 000 000 times less than albumin)

Biomarkers: applications to Duchenne dystrophy

Description	No. of peptides	Score	ANOVA (<i>p</i> -value)	Fold change DMD/Healthy
Titin	23	1469.0	3.88E-06	37.4
Uromodulin	13	777.4	3.96E-03	5.5
Cubilin	10	576.8	2.55E-03	-2.3
Nuclear transport factor 2	5	356.9	1.05E-04	5.8
TNF-receptor superfamily member 16	4	308.7	4.07E-05	3.3
Myosin-1	3	265.3	8.66E-04	39.4
Fibulin-2	3	256.7	1.75E-03	2.9
β-galactosidase	6	253.7	1.23E-03	-2.4
Complement C1r subcomponent-like protein	5	235.8	3.47E-05	2.7
Aminopeptidase	3	213.7	2.71E-03	2.4

From: **“Proteomics profiling of urine reveals specific titin fragments as biomarkers of Duchenne muscular dystrophy”**, J. Rouillon, A. Zocevic, T. Leger, C. Garcia, J-M. Camadro, B. Udd, B. Wong, L. Servais, T. Voit, F. Svinartchouk, 2014 *Neuromuscular Disorders*

Biomarkers: applications to Duchenne dystrophy

No. accession	Description	Localization	Peptides	Score	ANOVA (P-value)	Fold change
MYG_HUMAN	Myoglobin	Cytoplasm	4	195	2.7e-03	234.8
MYOM2_HUMAN	MYOM2	Myofibril	10	390	9.8e-05	100.1
MYOM3_HUMAN	MYOM3	Myofibril	11	491	1.5e-05	49.7
TPIS_HUMAN	Triosephosphate isomerase	Cytoplasm	3	128	2.3e-03	48.4
AATC_HUMAN	Aspartate aminotransferase	Cytoplasm	3	75	4.7e-04	45.7
KCRM_HUMAN	CK-M	Cytoplasm	15	849	2.9e-05	39.8
MYH7_HUMAN	Myosin-7	Myofibril	11	520	2.2e-05	38.3
ENOB_HUMAN	β -enolase	Cytoplasm	4	178	7.4e-05	34.8
G6PI_HUMAN	Glucose-6-phosphate isomerase	Cytoplasm/Secreted	4	130	1.6e-03	29.5
CAH3_HUMAN	Carbonic anhydrase 3	Cytoplasm	5	182	8.6e-05	23.9
FLNC_HUMAN	Filamin-C	Myofibril	4	145	4.3e-04	19.4
ALAT1_HUMAN	Alanine aminotransferase 1	Cytoplasm	4	127	3.0e-05	15.6
ALDOA_HUMAN	Fructose-bisphosphate aldolase A	Cytoplasm	15	729	9.3e-05	14.2
KPYM_HUMAN	Pyruvate kinase PKM	Cytoplasm	16	845	1.1e-05	12.8
TITIN_HUMAN	Titin	Myofibril	14	495	1.9e-03	10.8
VINC_HUMAN	Vinculin	Cytoplasm/Membrane	2	74	7.2e-05	10.3
PYGM_HUMAN	Glycogen phosphorylase, muscle form	Cytoplasm	8	257	6.1e-04	9.9
LDHA_HUMAN	L-lactate dehydrogenase A chain	Cytoplasm	8	378	9.1e-04	9.5
HPT_HUMAN	Haptoglobin	Secreted	29	1867	1.5e-04	7.6
HBD_HUMAN	Haemoglobin subunit δ	Cytoplasm	3	100	5.1e-03	6.2
LDHB_HUMAN	L-lactate dehydrogenase B	Cytoplasm	10	598	2.4e-05	5.4
HBB_HUMAN	Haemoglobin subunit β	Cytoplasm	7	552	8.0e-03	3.6
HBA_HUMAN	Haemoglobin subunit α	Cytoplasm	7	407	5.3e-03	3.4
TPM2_HUMAN	Tropomyosin β chain	Myofibril	5	170	2.0e-02	2.6

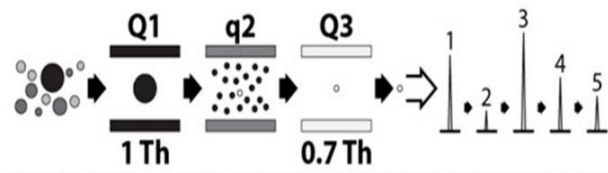


Rouillon, J., Zocevic, A., Poupiot, J., Amor, F., Léger, T., Garcia, C., Camadro, J.M., Wong, B., Cosette, J., ML Coenen-Stass, A., McClorey, G., C Roberts, T., JA Wood, M., Servais, L., Voit, T., Richard, I., Svinartchouk, F. (2015). *Serum proteomic profiling reveals specific MYOM3 fragments as biomarkers of Duchenne muscular dystrophy with applications for the follow-up of gene therapy treatment in a mouse model of muscular dystrophies.* *Human Mol. Genetics*

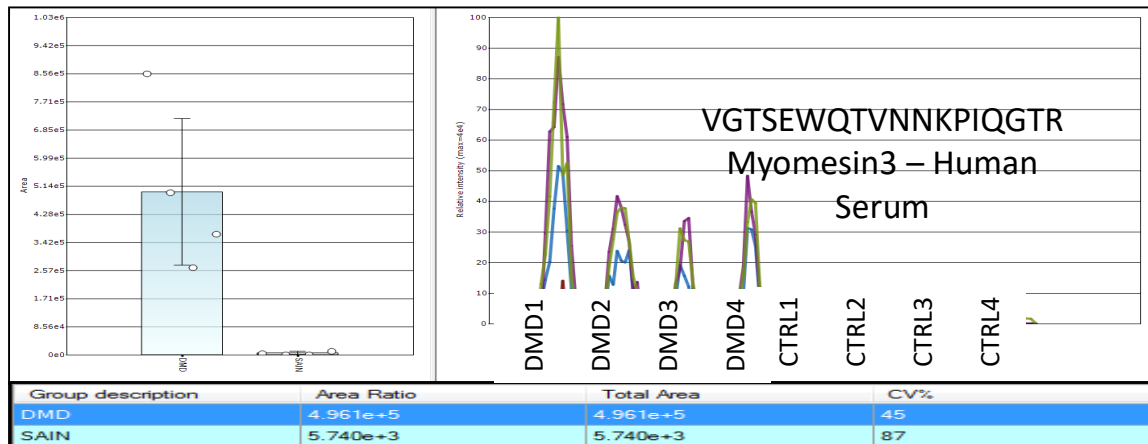
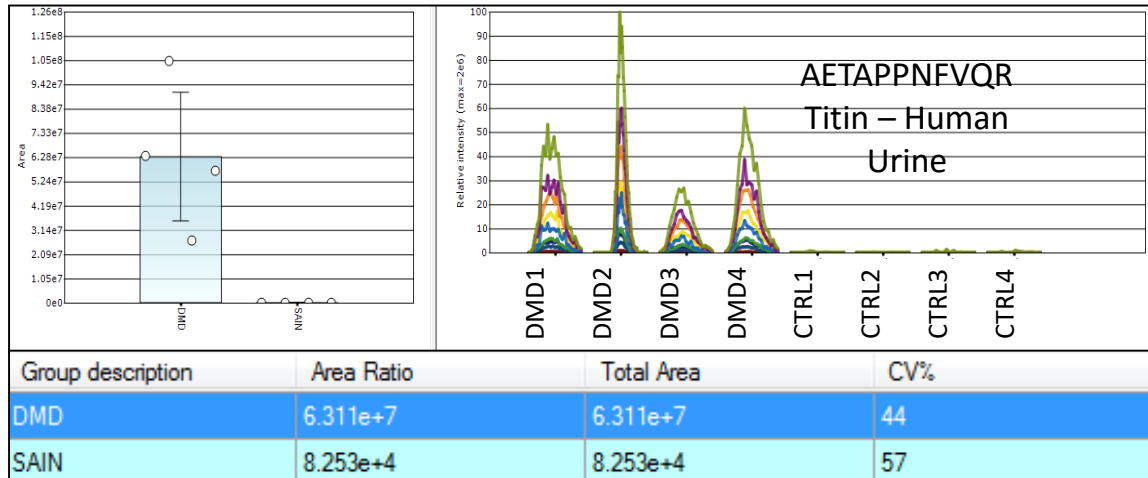
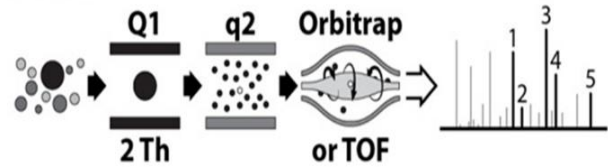
Jeremy Rouillon; Aleksandar Zocevic; **Thibaut Léger**; **Camille Garcia**; **Jean-Michel Camadro**; Bjarne Udd; Laurent Servais; Thomas Voit; Fedor Svinartchouk. (2014). *Proteomics profiling of urine reveals specific titin fragments as biomarkers of Duchenne muscular dystrophy.* *Neuromuscular disorders.*

Targeted proteomics PRM

SRM

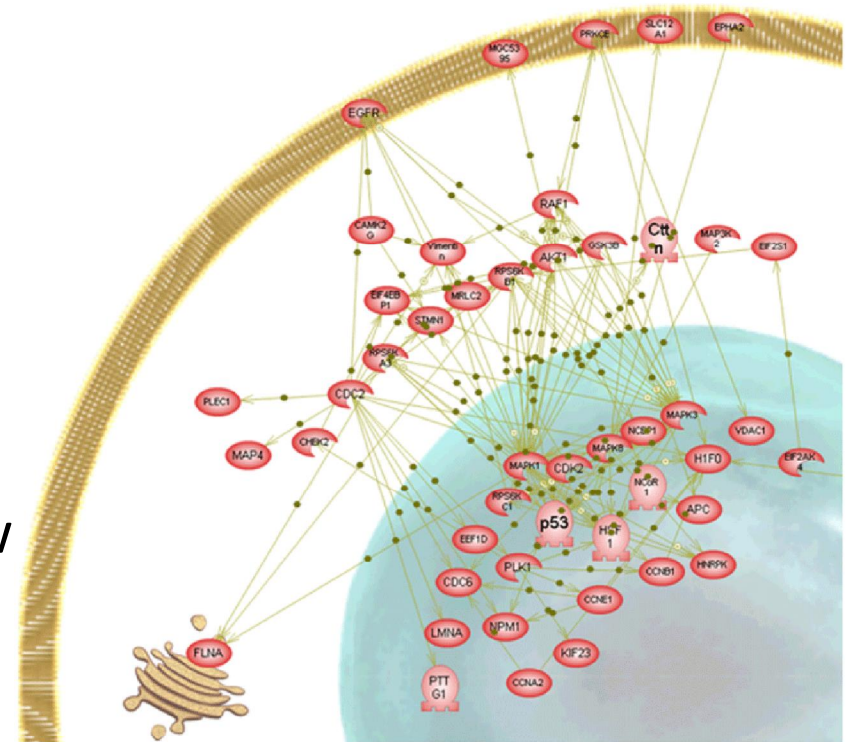


PRM



Why study PTMS?

- “ Cells can rapidly respond to stimuli and perturbations
- “ Important cellular mechanisms are tightly controlled
- “ Often, diseases (e.g. cancer) are due to aberrantly activated proteins
 - “ Protein expression is much too slow for quick adaption
 - “ PTMs are crucial regulator
 - “ MS-based proteomics allows to analyze complex networks of post-translationally modified proteins



PTMs *in vivo*

- “ **Phosphorylation** (Ser, Thr, Tyr; +80 Da)
 - “ Phosphorylation is one of the most important PTMs
 - “ A key event in signaling
 - “ Catalyzed by kinases/phosphatases
- “ **Glycosylation** (Asn, Ser, Thr)
 - “ marks proteins for degradation
 - “ s for degradation
- “ **Glycation** (Asn, Ser, Thr)
 - “ marks proteins for degradation
 - “ s for degradation
- “ **Ubiquitination** (Lys; +114 Da)
 - “ marks proteins for degradation
- “ **Proteolytic cleavage**
- “ **Acetylation** (N-termini and Lys +42 Da)
 - “ often combined with removal of protein initial Met

Others: oxidations, methylations, sumoylations, glutathionylations...

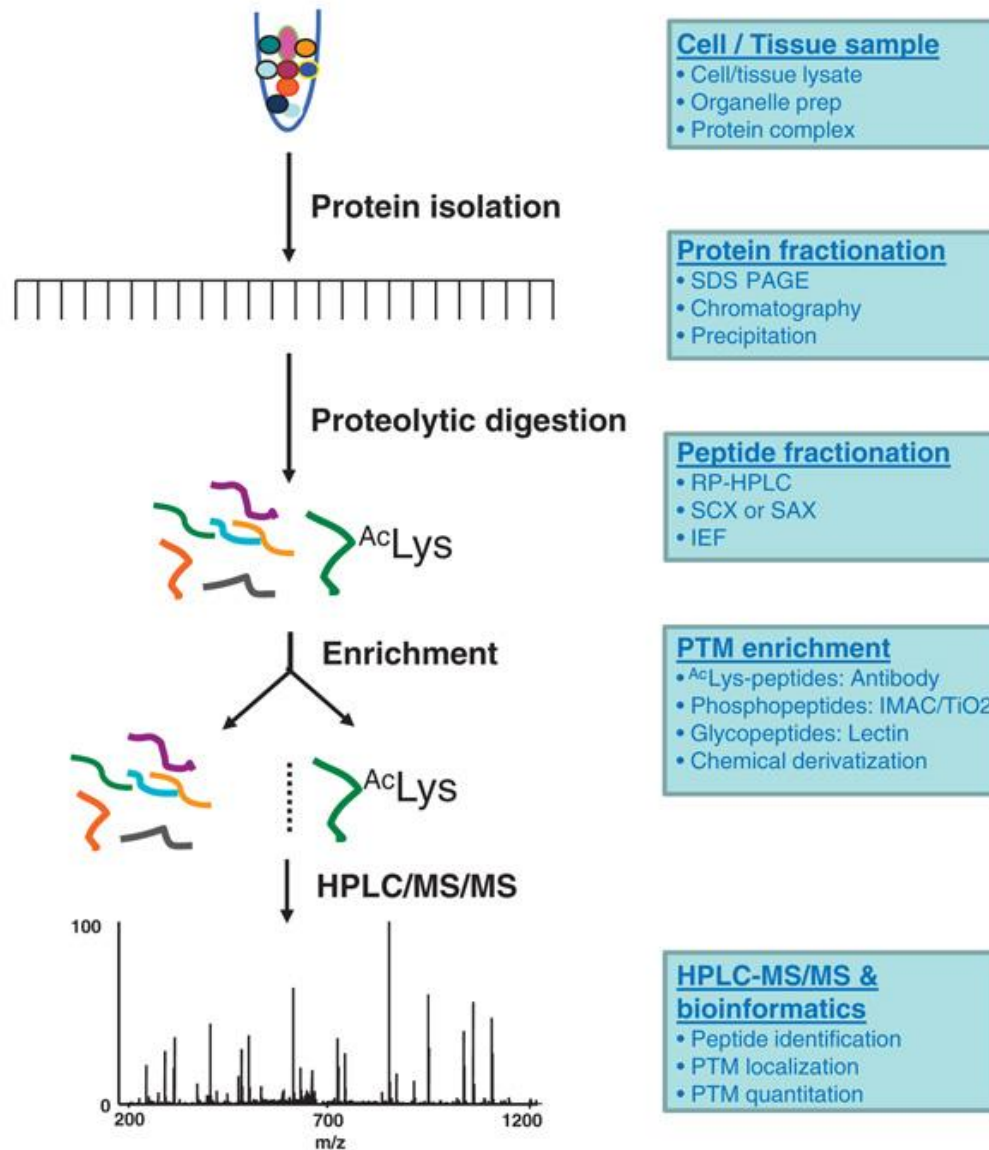
PTMs characterization: techniques

Techniques for detection and identification of PTM substrates

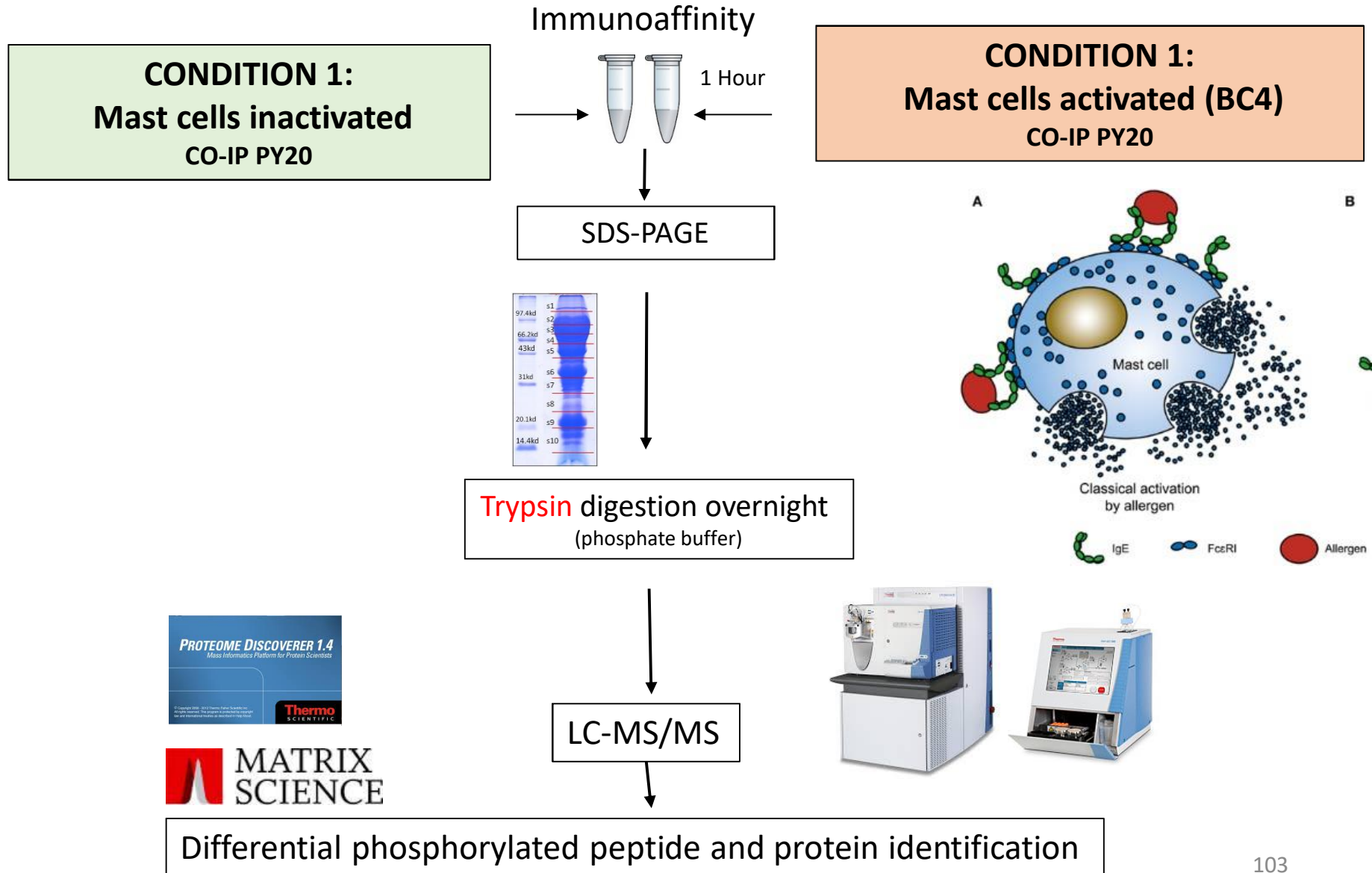
Method	<i>In vitro</i> or <i>in vivo</i>	Case studies	Advantages	Disadvantages
Radioactive isotope labeling	<i>In vitro</i> or <i>in vivo</i>	^{32}P (pSer, pThr, pTyr) ^3H , or ^{14}C for AcLys or MeK	Reagents accessible	Inconvenience/hazard low sensitivity
Western blotting	<i>In vitro</i> or <i>in vivo</i>	pTyr, AcLys or MeK	Good affinity	Moderate sensitivity
Peptide/protein array	<i>In vitro</i>	pSer/Thr/Tyr, AcLys or MeK	Rapid, global scale	Possibly non-specific, low sensitivity, requires verification
MS-proteomics	<i>In vitro</i>	pSer/Thr/Tyr, AcR or MeK	Specific, global scale	Need enrichment methods

AcR , MeK , pSer, pThr, and pTyr, represent acetylated arginine, methyllysine, phosphorylated serine, threonine, and tyrosine residues, respectively.

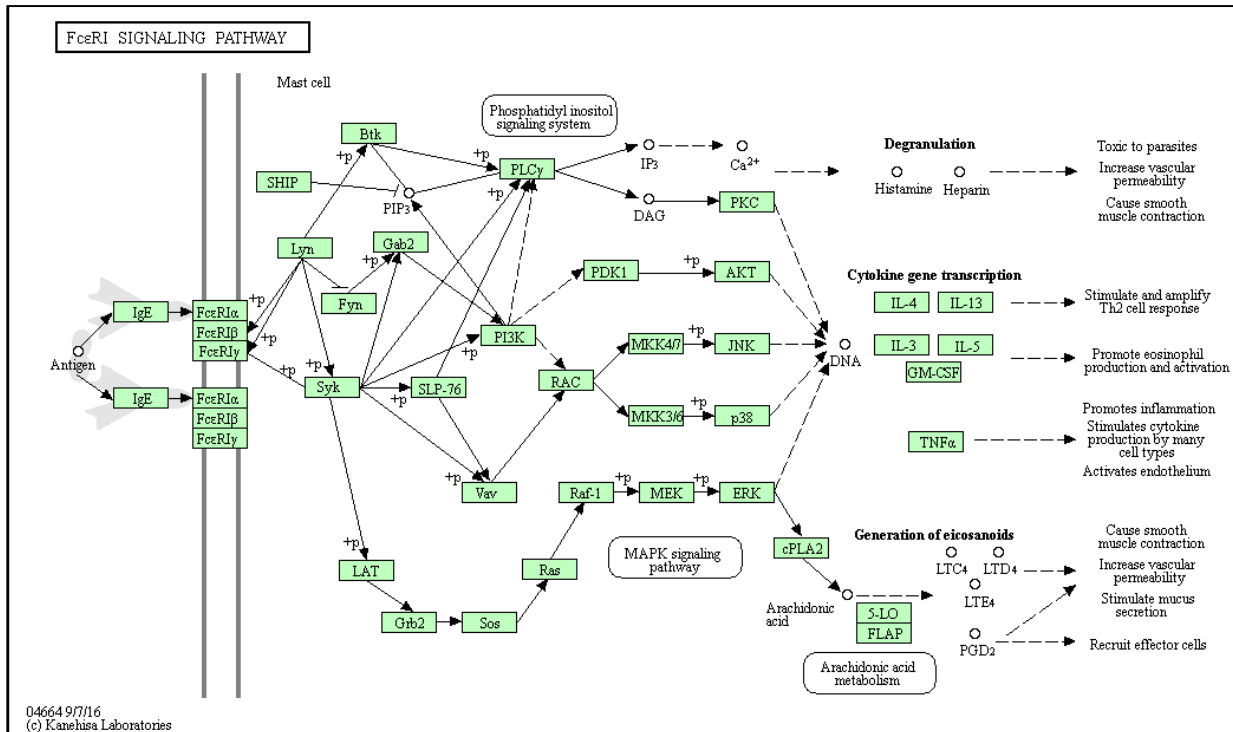
Workflow for PTMs characterization



Quantitative proteomics and phosphorylations



Accession	Protein Name	Gene	Species	Cellular Component	Molecular Function	Biological Process	PF0017	PF0069	PF07714	763.10	676.62	45.9	4.81							
953	P20411	High affinity immunoglobulin epsilon receptor subunit gamma	Chn1	transducer activity	cell surface; membrane	to stimulus; transport	PF11628			573.65	352.21		9.8							
963	Q64725	Tyrosine-protein kinase SYK OS=Rattus norvegicus GN=Syk	Fynal	transducer activity	cleus; organelle lumen	to stimulus; transport	PF00017; PF00069; PF07714			835.28	323.20		71.5							
964		Sequence	# PSMs	# Proteins	# Protein Groups	Protein Group Accessions	Modifications	MH+ [Da]	phosphoRS Site Probabilities	A4	IonScore A4	Exp Value A4	B4	IonScore B4						
965		LLTLEDNELGSGNFGTVK	2	1	1	Q64725		1906.97726	High		107	1.50125E-10								
966		DESEQTVLIGSK	4	1	1	Q64725		1305.65435	High		84	2.49757E-08	-high	67						
967		ELNGTYAISGGR	7	1	1	Q64725		1237.61919	High		55	2.38967E-05	-high	58						
968		ADENYYK	3	1	1	Q64725		902.38894	High		46	0.000100972	-high	23						
969		NVLLVTQHYAK	8	1	1	Q64725		1285.72683	High		44	0.000137239	-high	31						
970		ISDFGLSK	6	1	1	Q64725		866.46208	High		42	0.000234007	-high	40						
971		LIATTAHEK	7	1	1	Q64725		983.55124	High		37	0.000403124	-high	37						
972		LRNYYVDVNV	4	1	1	Q64725		1318.64326	High		37	0.001486192	edu	24						
973		YLEESNFVHR	4	1	1	Q64725		1293.62261	High		36	0.00176677	-high	27						
974		MGCPGGCPR	4	1	1	Q64725	M1(Oxidation); C3(Carbamidon)	1047.41679	High		35	0.000734469	-high	41						
975		GSEVTAMLEK	2	1	1	Q64725		1064.52982	High		34	0.002136328								
976		GSEVTAMLEK	4	1	1	Q64725	M7(Oxidation)	1080.52370	High		34	0.002154235	edu	20						
977		EVYLRK	2	1	1	Q64725		922.49926	High		33	0.001493862								
978		ALRADENYYK	2	1	1	Q64725		1242.61182	High		30	0.007003353								
979		VLTVPQCK	5	1	1	Q64725	C6(Carbamidomethyl)	944.52278	Medium		25	0.02099256								
980		GSEVTAMLEKGER	1	1	1	Q64725	M7(Oxidation)	1422.68938	Medium		25	0.027162273								
981		LRNYYVDVNV	6	1	1	Q64725	Y4(Phospho)	1398.61077	Y(4): 93.9; Y(5): 5.7; Y(6): 0.4	Medium	24	0.023646873								
982		TGPFEDLKENLR	2	1	1	Q64725		1531.81073	Medium		22	0.025122601								
983		GSEVTAMLEKGER	2	1	1	Q64725		1406.69446	Medium		20	0.067444564								
984		KPFNRPPGVQPK	1	1	1	Q64725		1364.77961	Medium		19	0.035807446								
985		LLTLEDNELGSGNFGTVK	1	1	1	Q64725		2035.07124	Low		19	0.093821803								
986		WYAPECINYFK	1	1	1	Q64725	C6(Carbamidomethyl)	1490.67815	Low		17	0.171057347								
987		NYYVDVNV	2	1	1	Q64725		1049.45671	Low		16	0.121475657								
988		MGCPGGCPR	1	1	1	Q64725	C3(Carbamidomethyl); C7(Car)	1031.42131	Low		15	0.108098357								
989		MPWFFHGNISR	1	1	1	Q64725	M1(Oxidation)	1260.59497	Low		13	0.348397318								
990		YLQQR	1	1	1	Q64725		821.42674	Low		12	0.268819476								
991		NYLGGFALSAVHR	1	1	1	Q64725	Y2(Phospho)	1598.76055	Y(2): 99.9; S(9): 0.1									Low	11	
992	Q5U2U2	Crk-like protein OS=Rattus norvegicus GN=Crkl PE=1 SV=1		protein binding m; cytosol; membrane	development	PF00017; PF00018; PF07653				234.55	96.32		33.8	6.74						
1005	P60868	40S ribosomal protein S20 OS=Rattus norvegicus GN=Rps20 F: tural molecule activity	ism; cytosol; ribosome	metabolic process	PF00338					172.28	36.35		13.4	9.94						



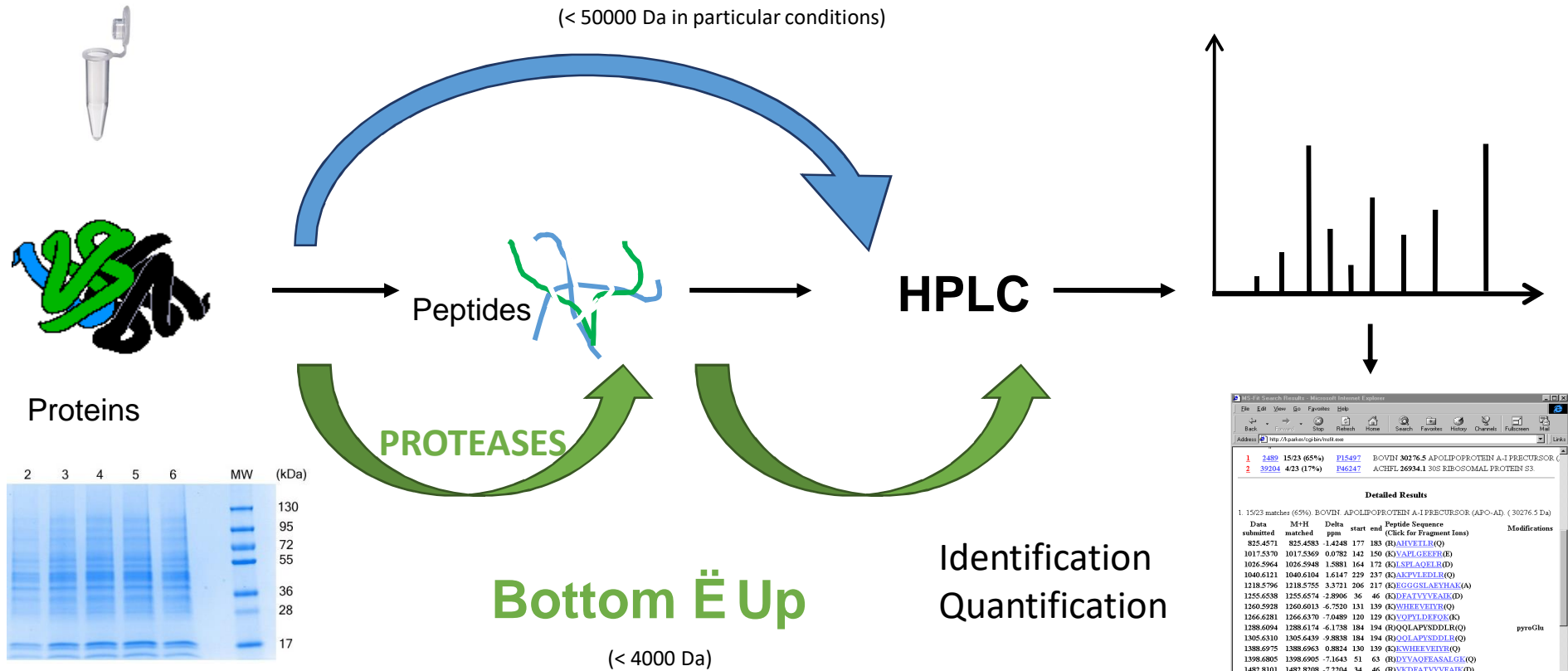
Proteomics workflows

Top Æ Down

(< 50000 Da in particular conditions)

MS acquisition

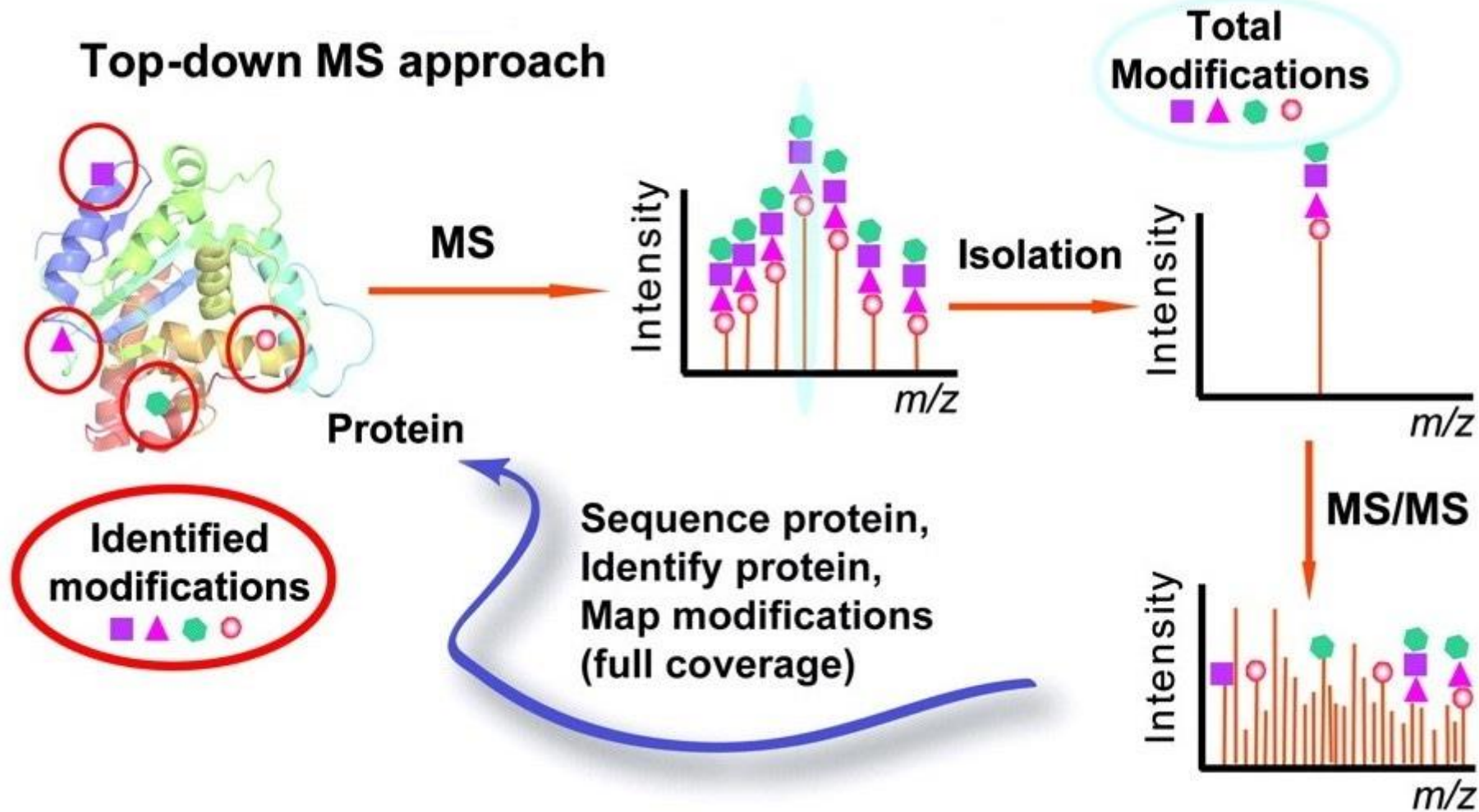
- ~ MALDI TOF/TOF
- ~ Orbitrap



The strategy is dependent of the sample complexity

Search engine (Mascot, Sequest, Peaks, Maxquant, OMSSA, ProSight, Byonik)

TOP DOWN proteomics for PTMs characterization



Challenges in TOP-DOWN proteomics

Challenges

Innovations

1. Protein solubility

Conventional surfactant (e.g. SDS) not compatible with MS

Develop new top-down MS compatible surfactant

2. Proteome complexity

Intact protein chromatography underdeveloped

Develop novel multi-dimensional chromatography for intact protein separation

3. Proteome dynamic range

Difficulty in detecting low abundant proteins

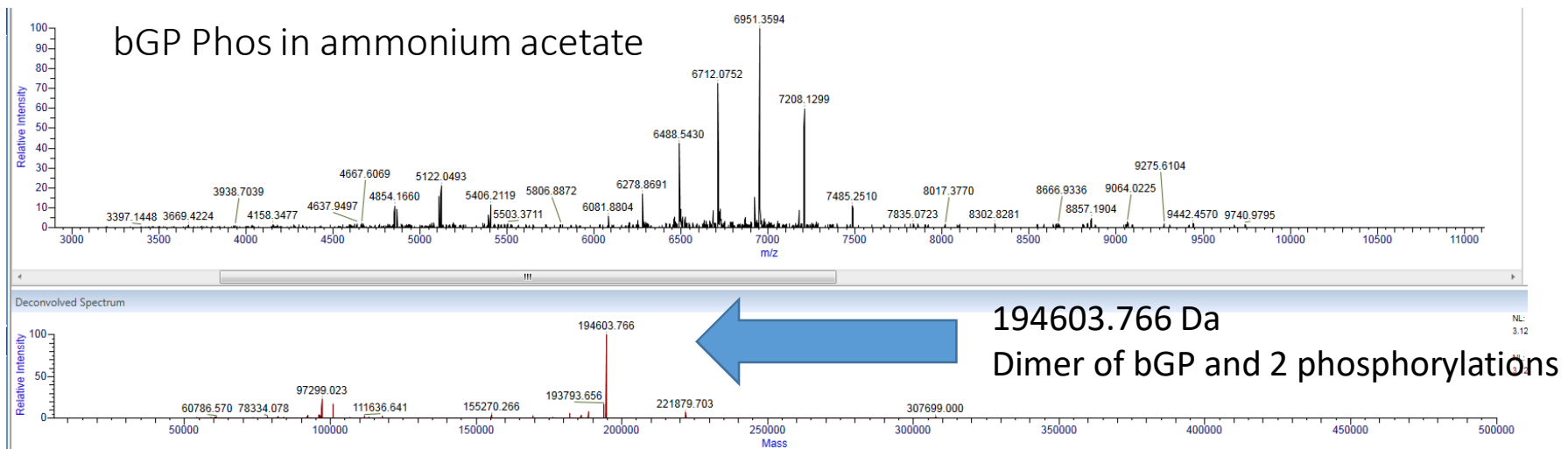
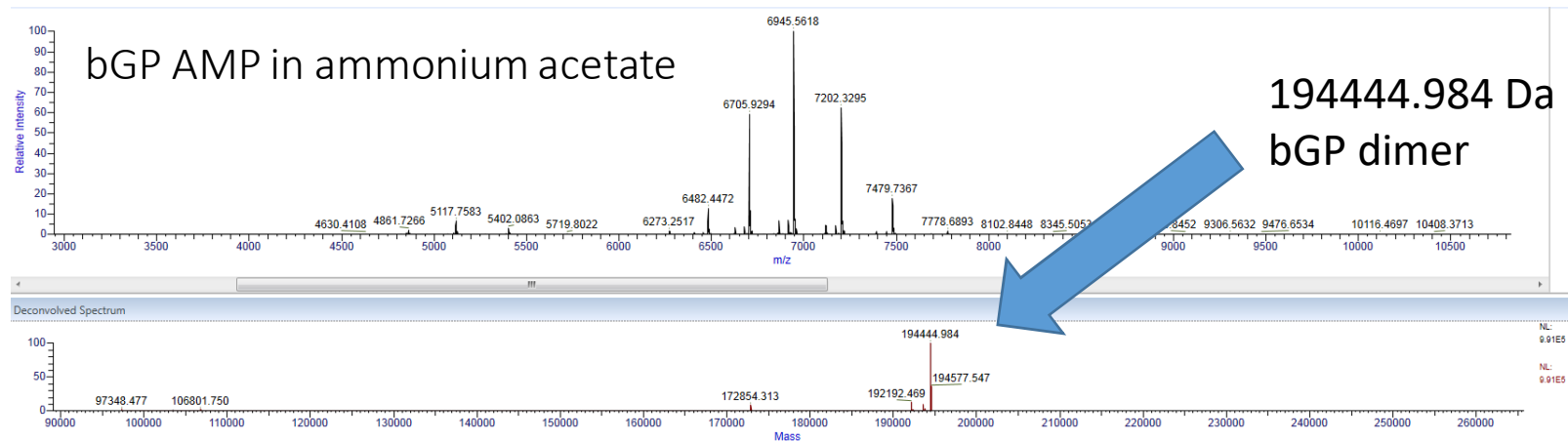
Develop novel nanomaterials for enriching low abundant proteins

4. Protein MS data interpretation

Software for top-down proteomics underdeveloped

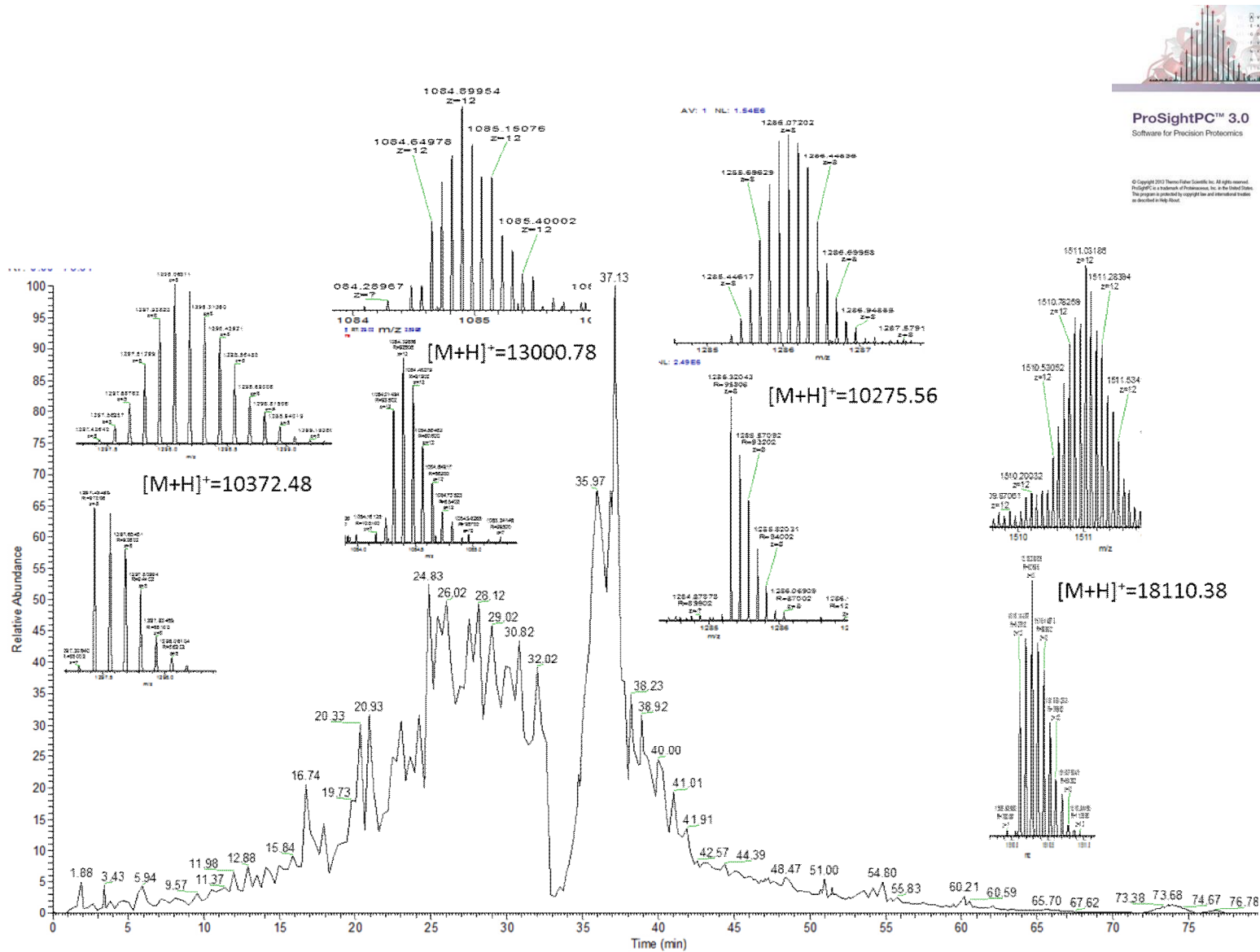
Develop user-friendly and versatile software interface

Analysis in intact protein mode: human brain glycogen phosphorylase

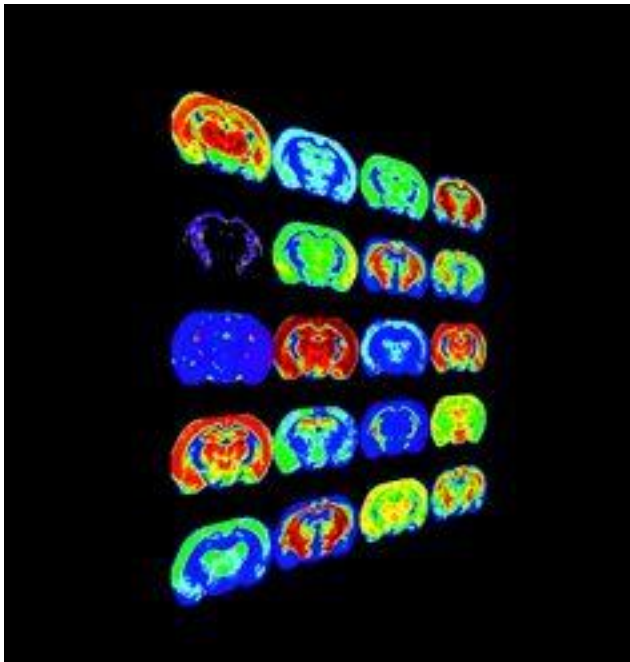


Crystal structure of human brain glycogen phosphorylase. Cécile Mathieu, Ines de la Sierra-Gallay, Romain Duval, Ximing Xu, Angélique Cocaïgn, Thibault Léger, Jean-Michel Camadro, Catherine Etchebest, Ahmed Haouz, Jean-Marie Dupret, Fernando Rodrigues-Lima. Under review.

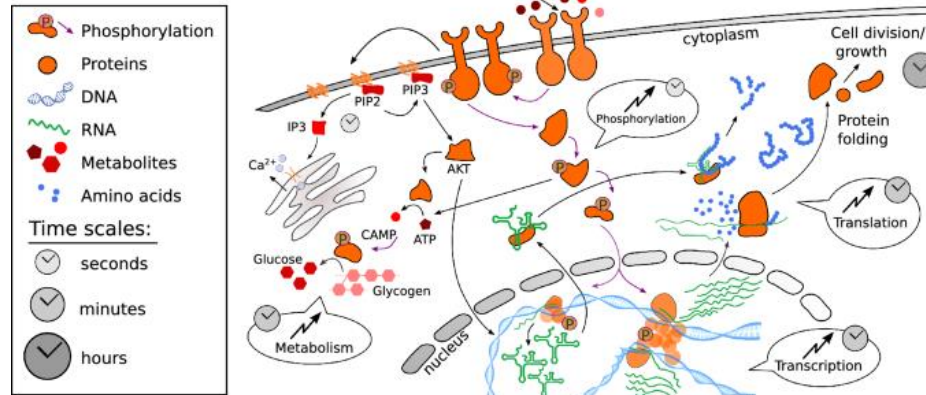
Analysis in intact protein mode



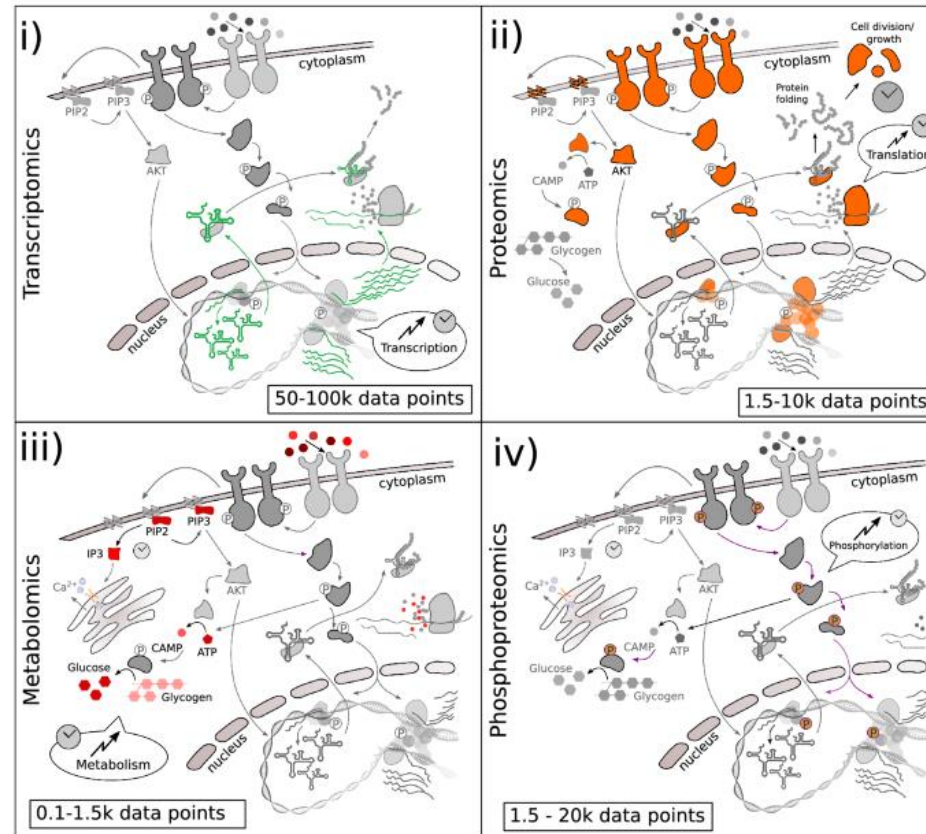
Emerging MS technologies



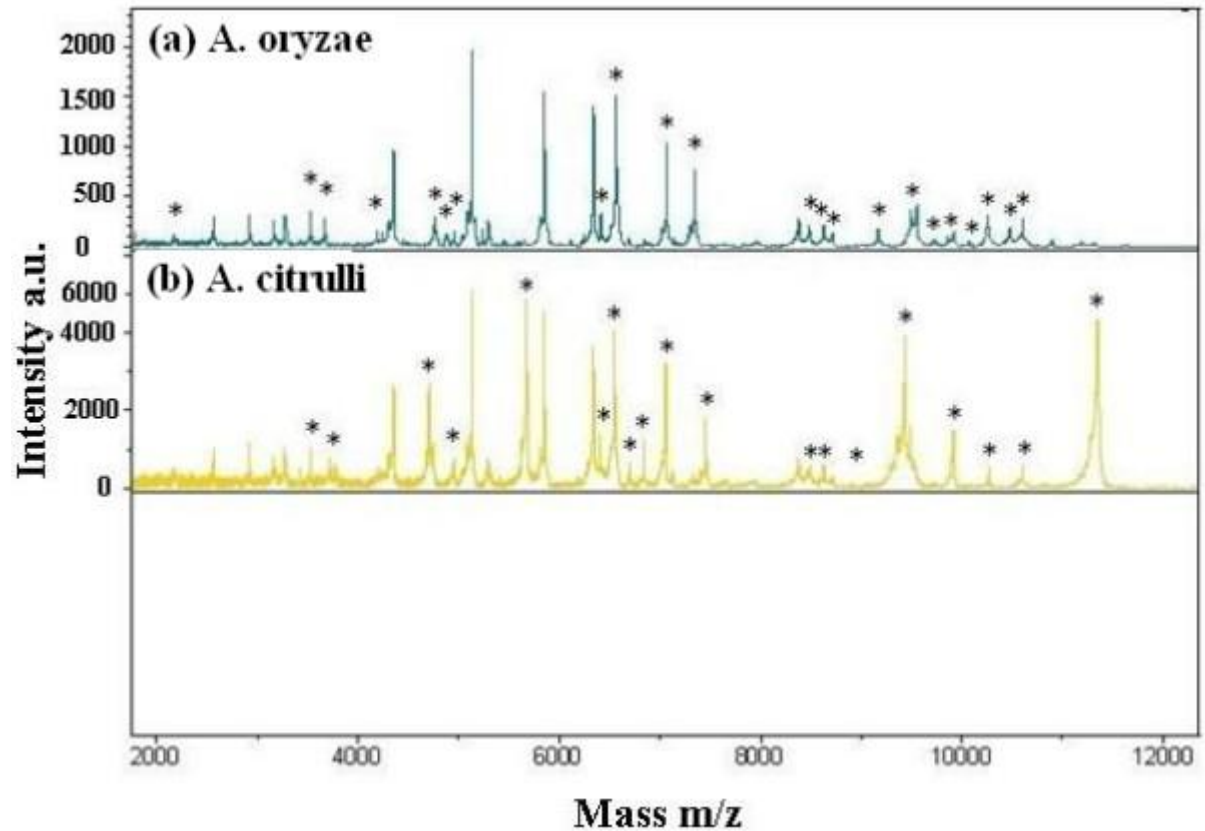
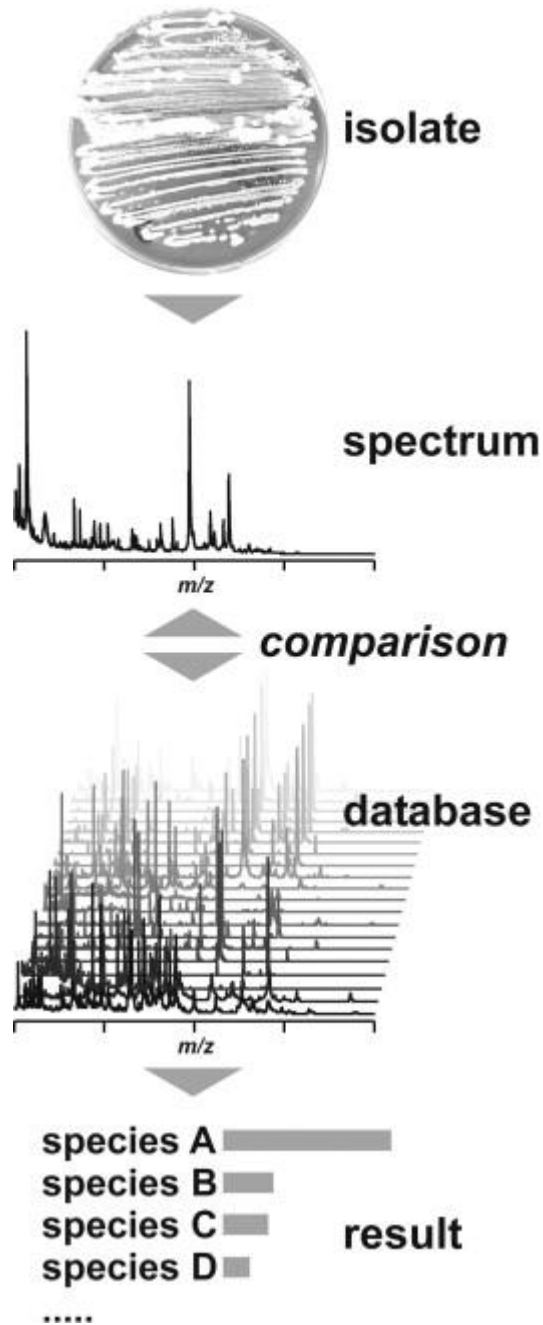
Multi-omics



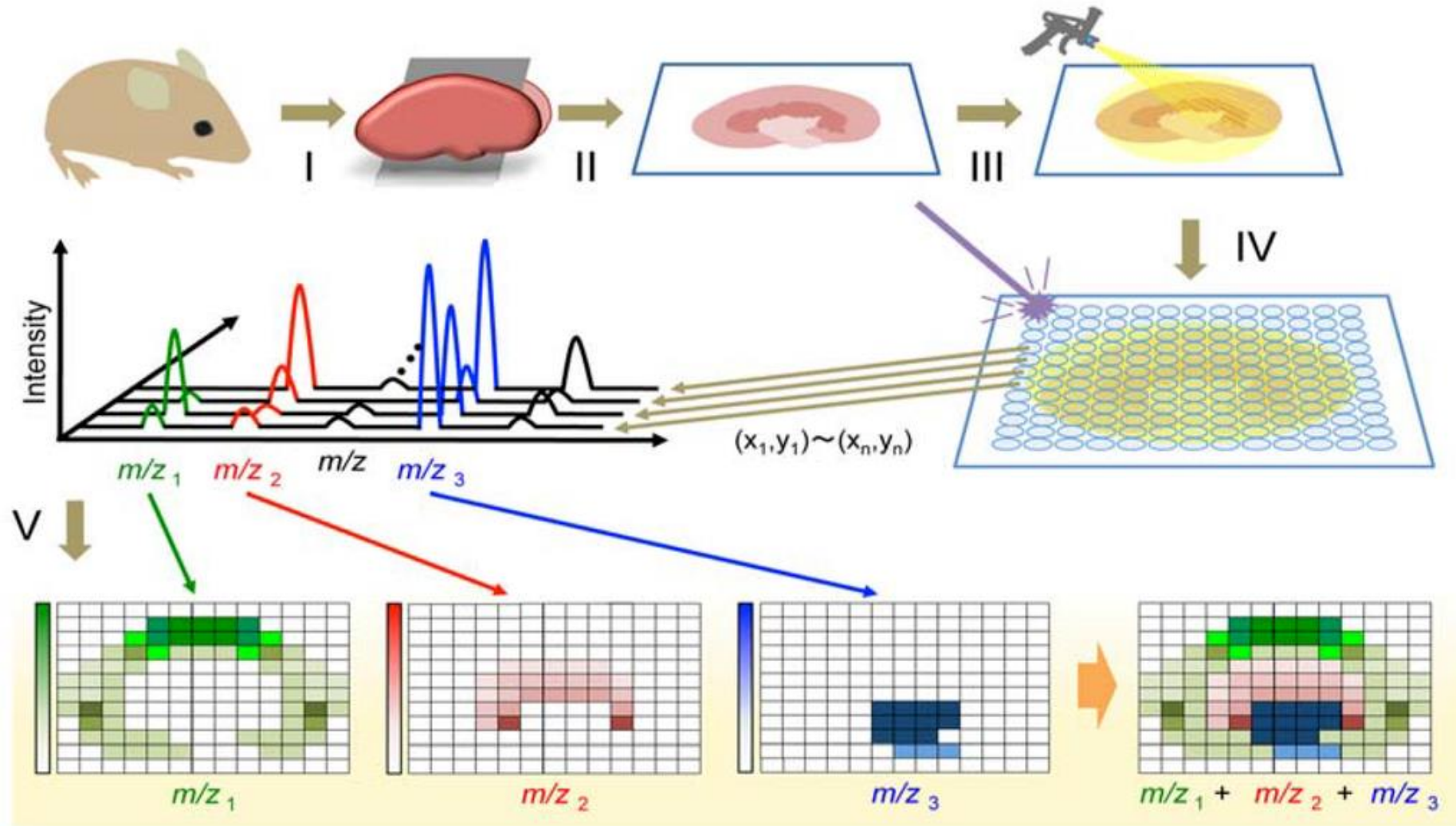
(a)



MS identification in Bacteriology (Biotyper)



Mass spectrometry imaging



I Sacrifice and organ dissection

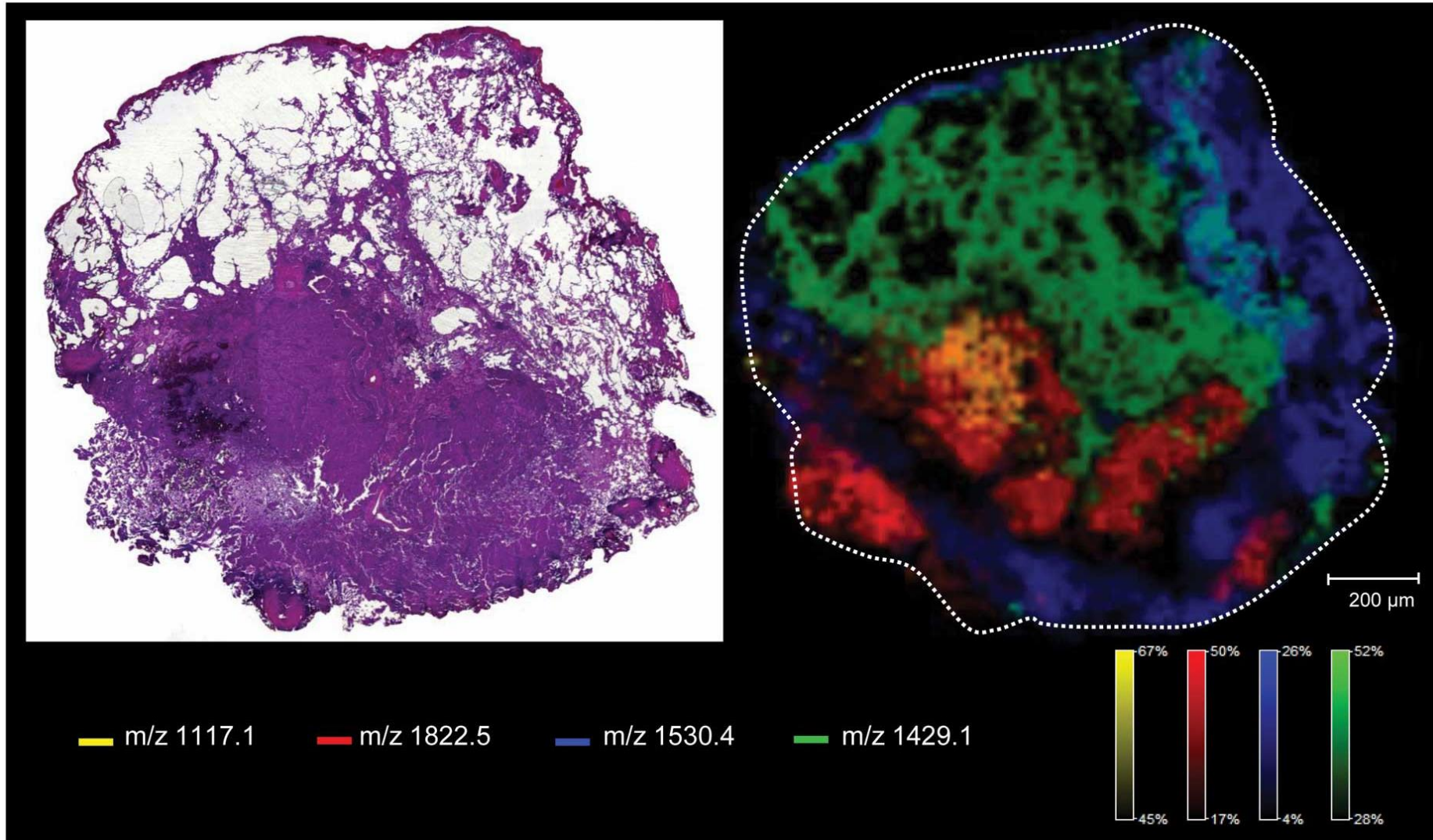
II Cryosectioning and moving to ITO glass slide

III Matrix deposition

IV MALDI laser 2D scanning

V Reconstruction of intensity image

Mass spectrometry imaging

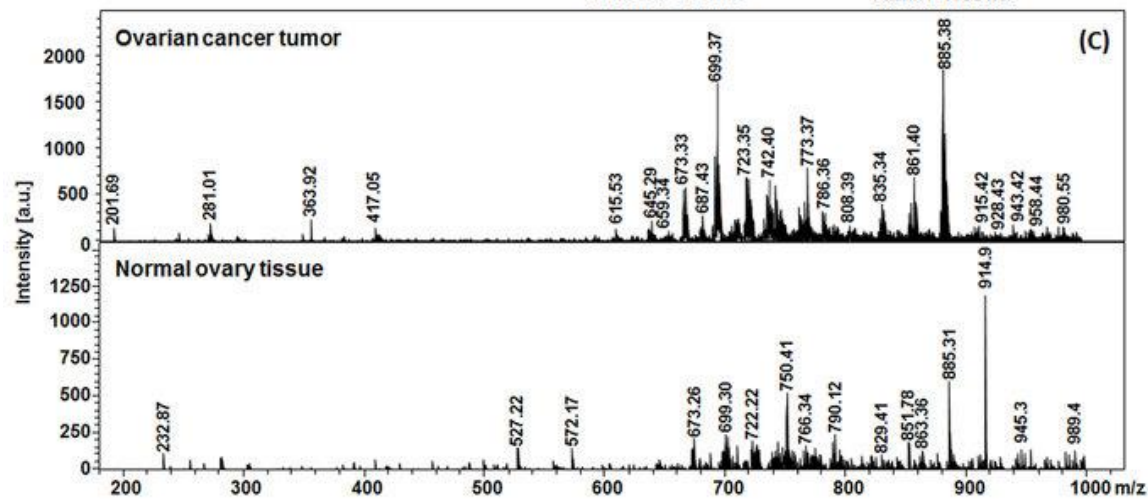
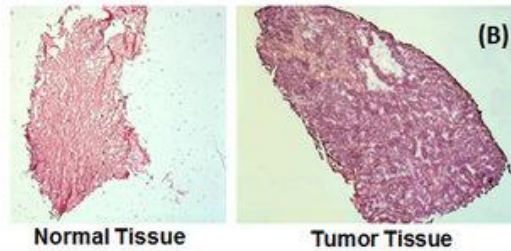
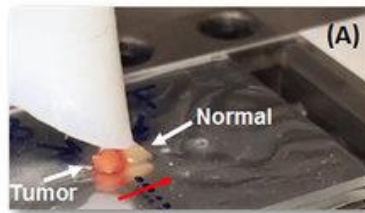


SPIDERMASS



PRISM

Protéomique • Réponse Inflammatoire • Spectrométrie de Masse
INSERM U 1192 – Université de Lille



Thank you for your attention!

